

isolation , processing of specimen for isolation
, isolation in culture , pure culture
Techniques

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Specimen collection

Many different specimens are sent for microbiological examination from patients with suspected bacterial infection.

Common specimens include **urine, faeces, wound swabs, throat swabs, vaginal swabs, sputum, and blood.**

cerebrospinal fluid, pleural fluid, joint aspirates, **tissue, bone and prosthetic material** (e.g. line tips).

Methods of isolating pure culture

1. Mixed culture

A culture contains more than one species of microbes.

2. Pure culture

A culture containing only one species of microbes.

The following methods are used to isolate pure culture.

A pure culture is usually derived from a mixed culture by transferring a small sample into new, sterile culture medium by dispersing or thinning the mixed culture multiple times before inoculating them. This technique was developed by German bacteriologist **Robert Koch**.

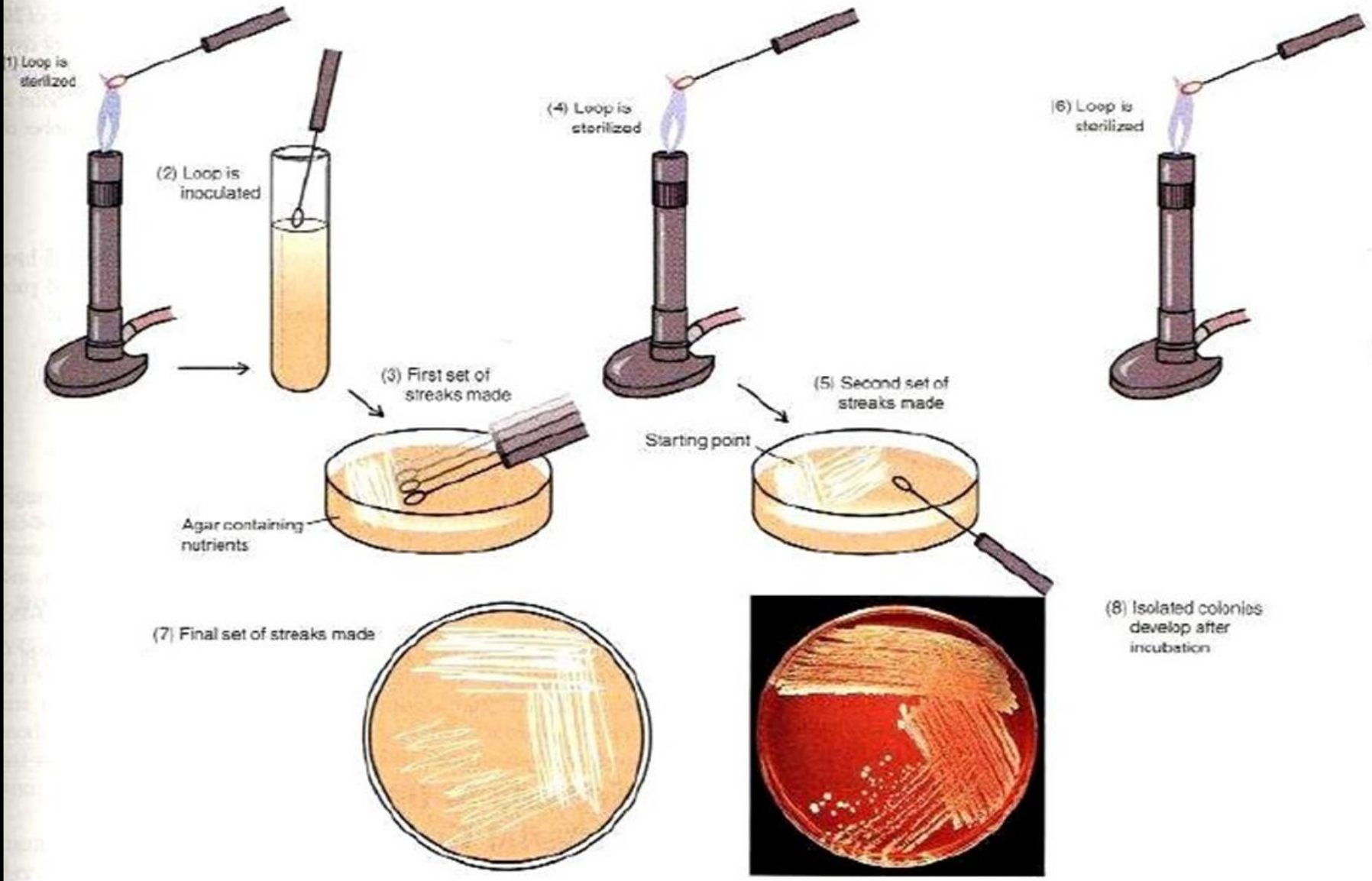
- **NEED OF PURE CULTURE:**
- To identify and study the characters of microbes.
- To detect the causative agent of particular disease.
- In order to find out antibiotic sensitivity and bacteriophage & bacteria susceptibility.
- Mutation rate in pure culture is low and clone is 99% identical.

Isolation

- Microorganisms occur in huge numbers.
- *Isolation of single species (pure culture) is done by :
 - **Streak plate method**
 - **Spread plate method**
 - **Pour plate method**
- *Based on diluting sample out to a point where a single cell will give rise to a single colony

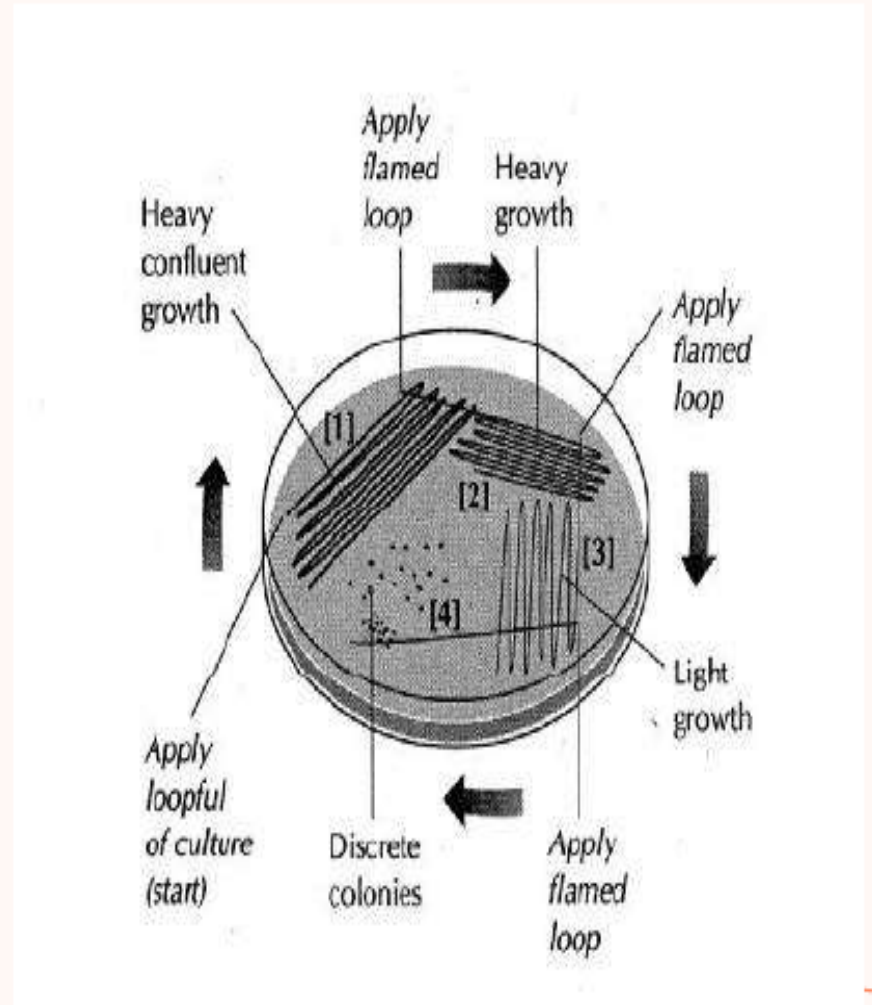
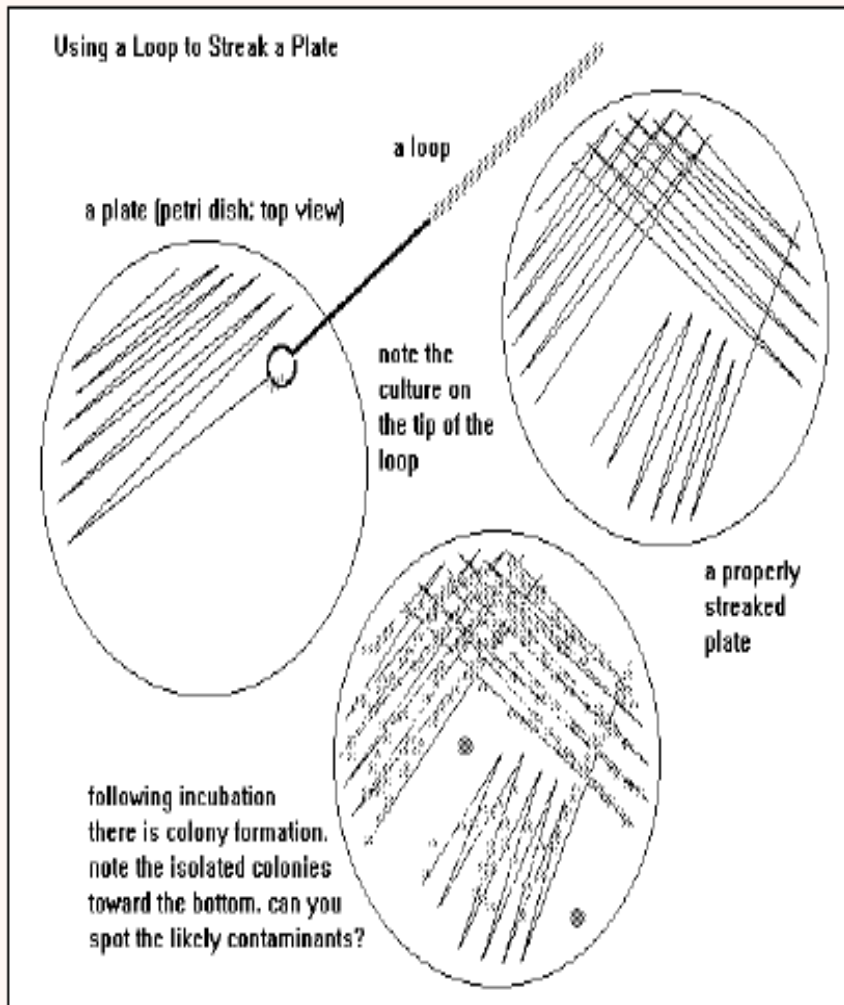
STREAK PLATE METHOD OF ISOLATION

- ❖ Most common way of separating bacterial cells on the agar surface to obtain isolated colonies
- ❖ After incubation
 - area at the beginning of the streak pattern -confluent growth
 - area near the end of the pattern should show discrete colonies

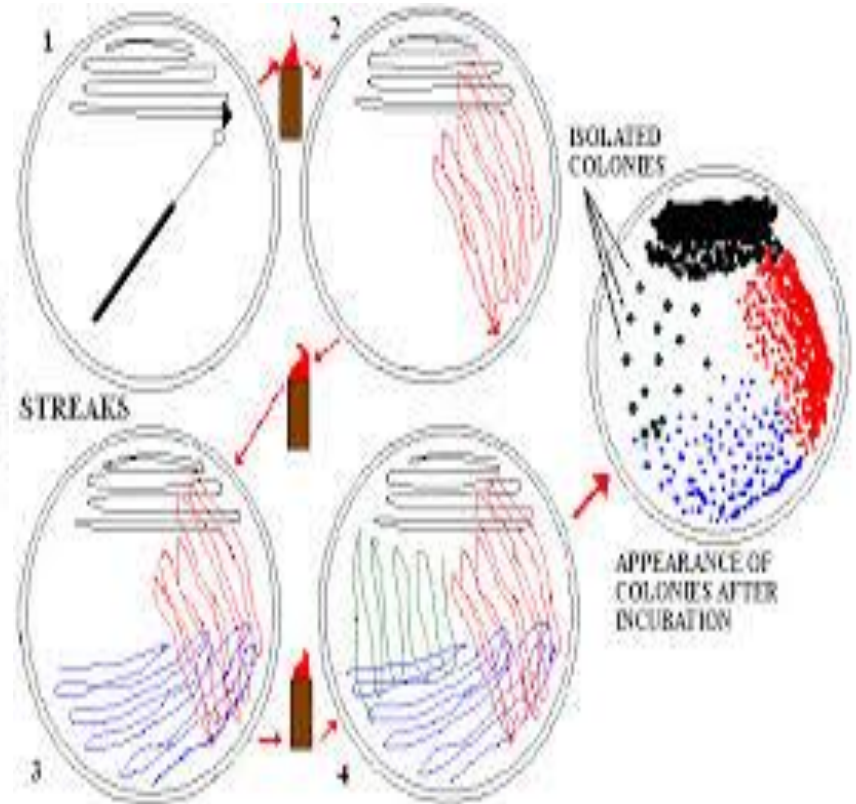
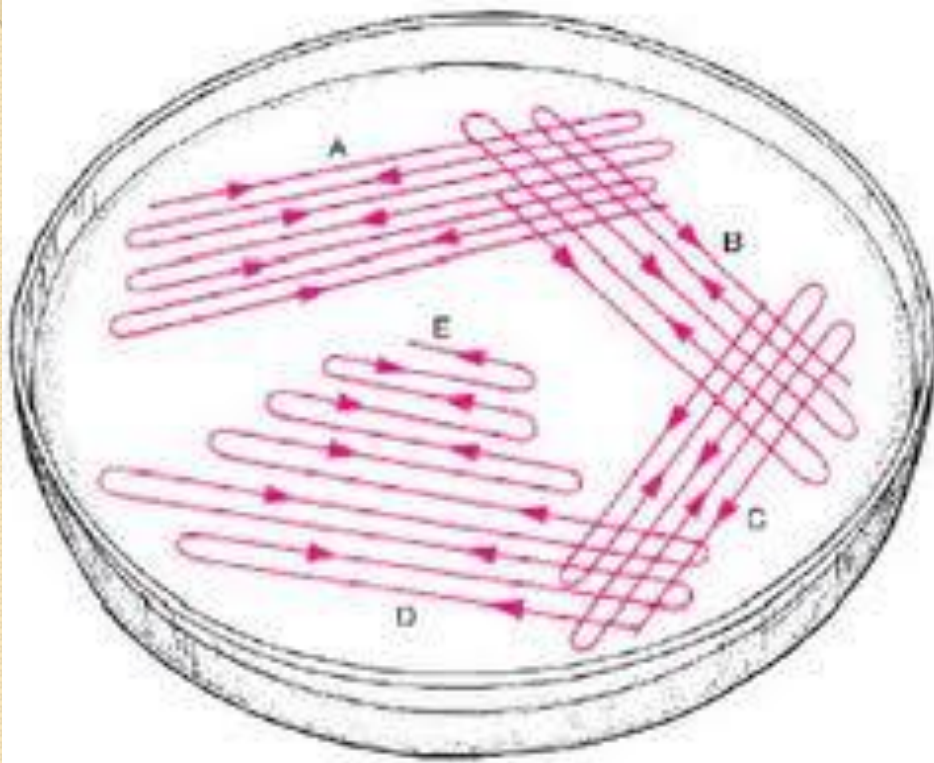


Streak-plate method

Pure culture – Streak Plates

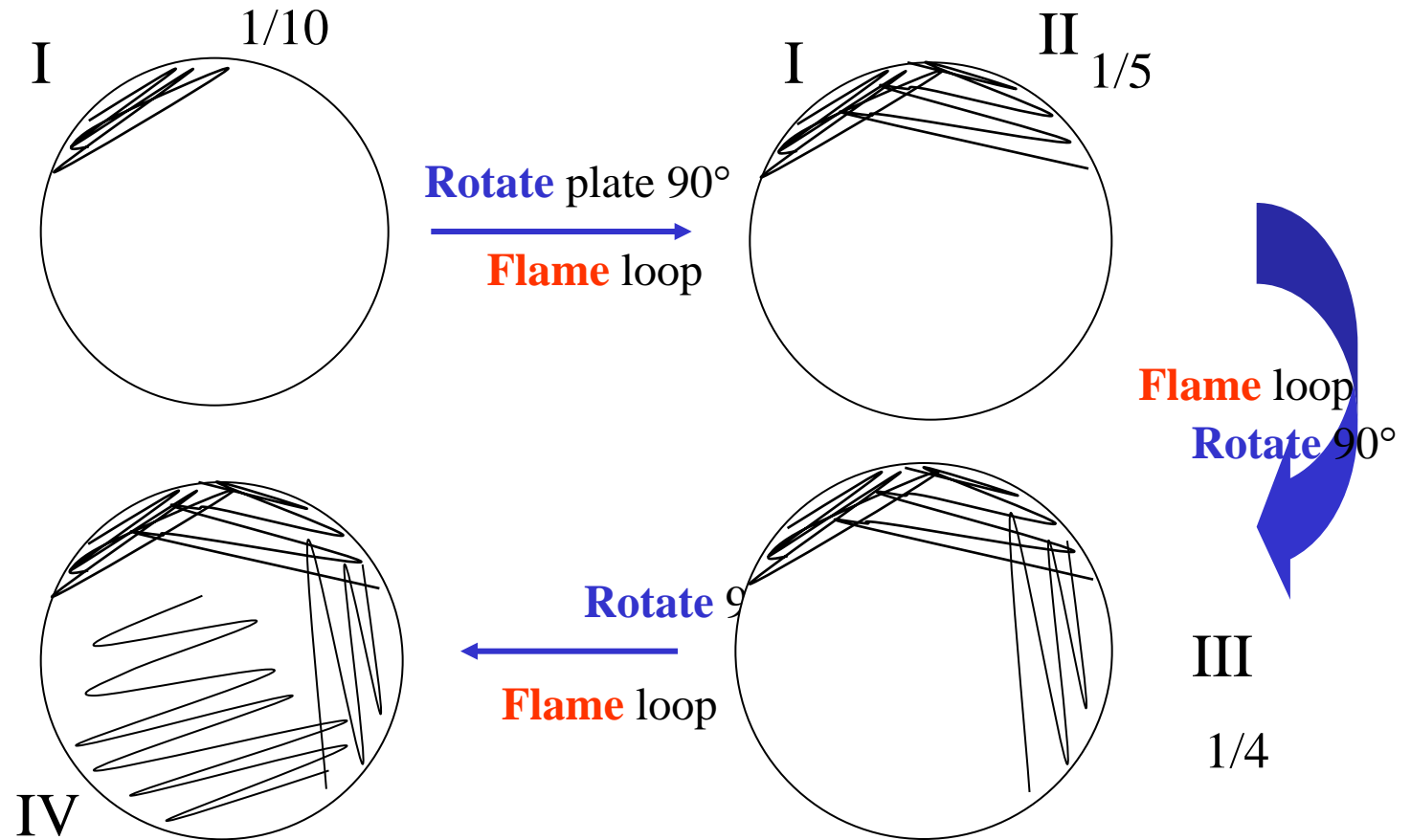


streak – plate technique

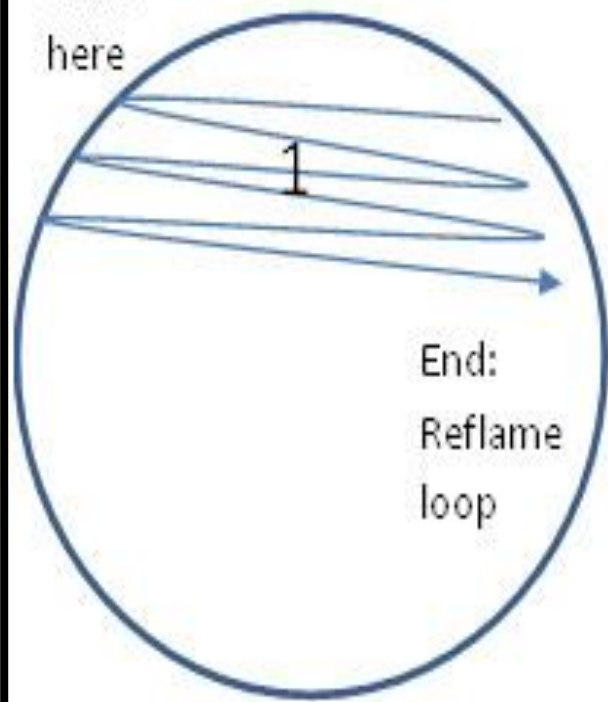


Streak-plate technique

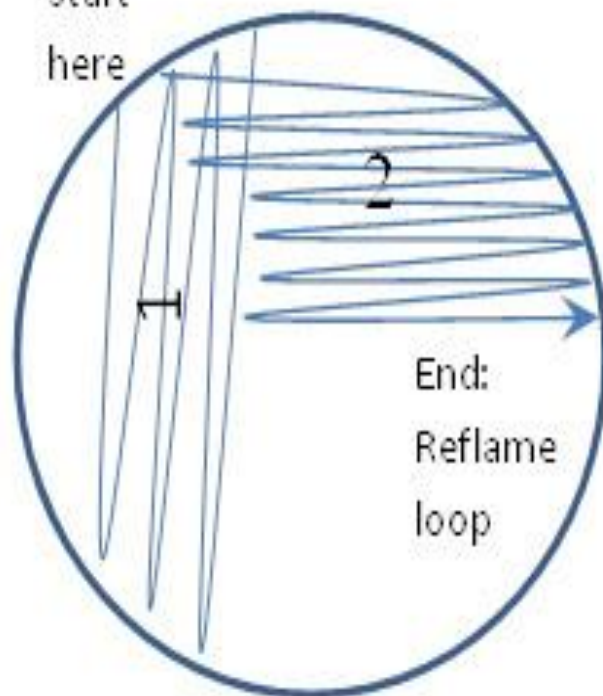
four-area streak plate technique



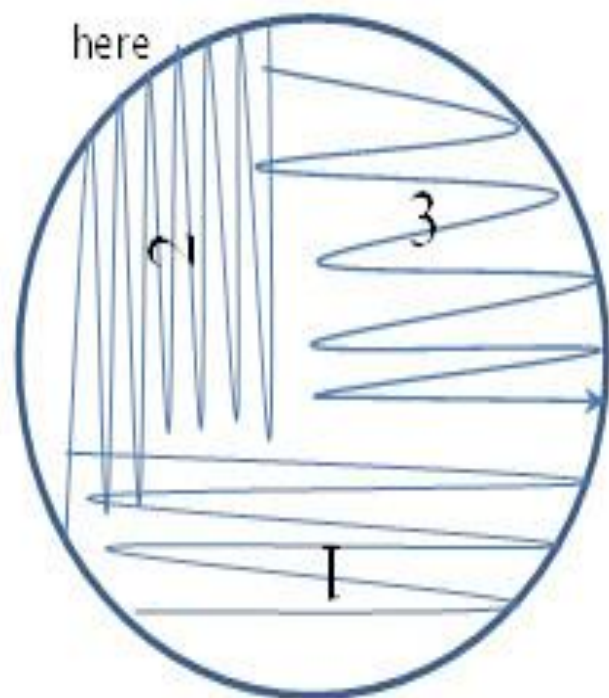
Start
here



Start
here



Start
here



Step 1: Streak plate across the top sector, using continuous motion, but do not cross lines.

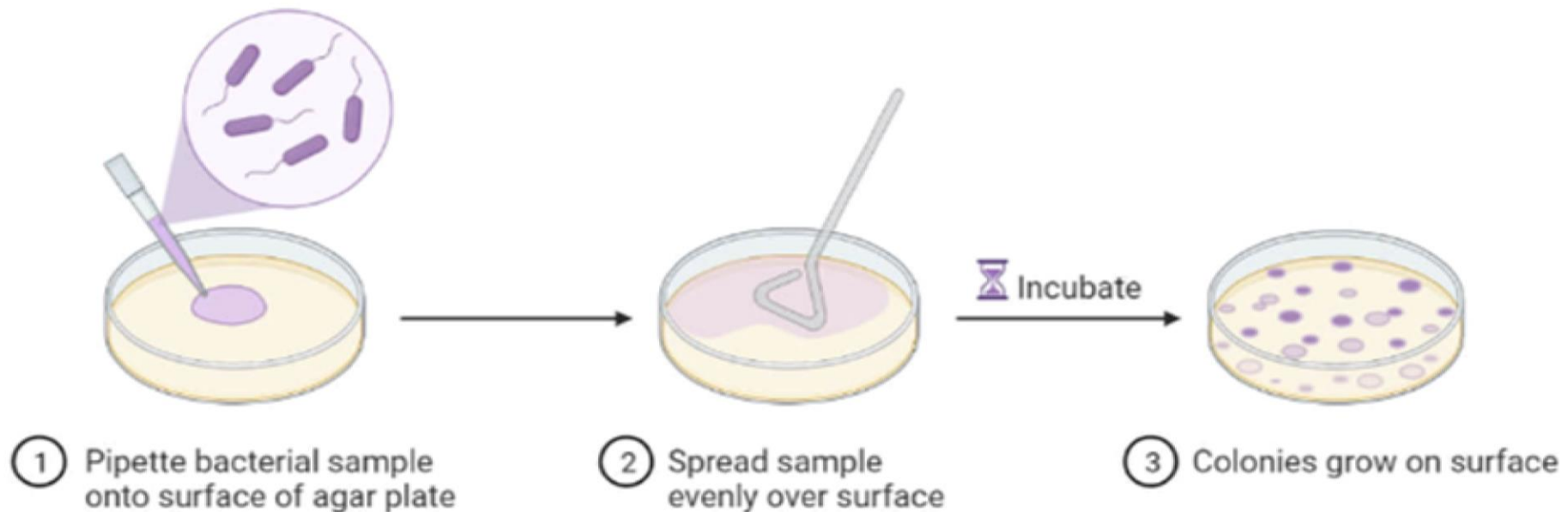
Step 2: Turn plate 90 degrees. Streak plate across the second sector, using continuous motion, making sure to cross into the first sector during the first streak or two.

Step 3: Repeat step two, but streak across sector 3.

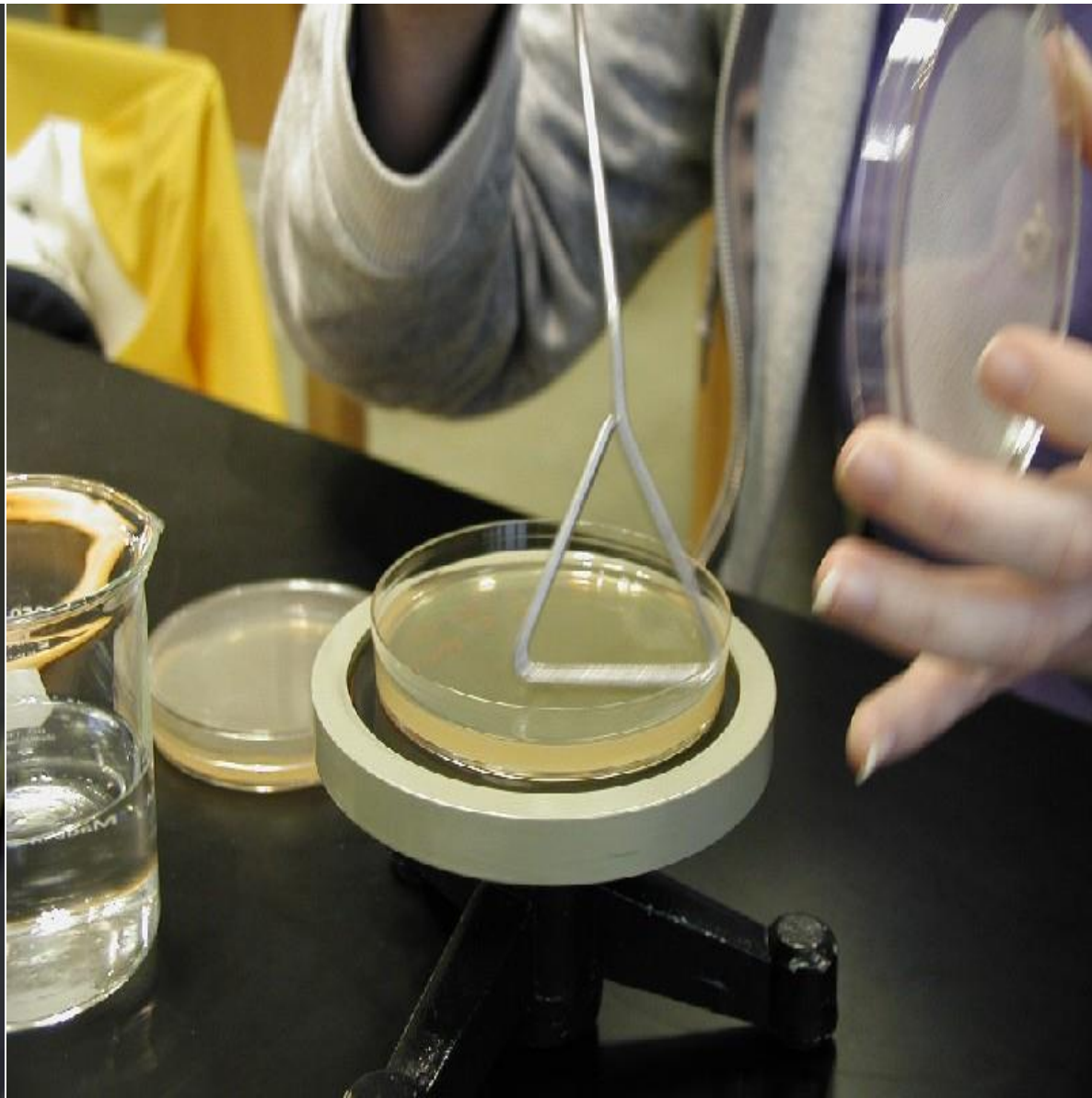
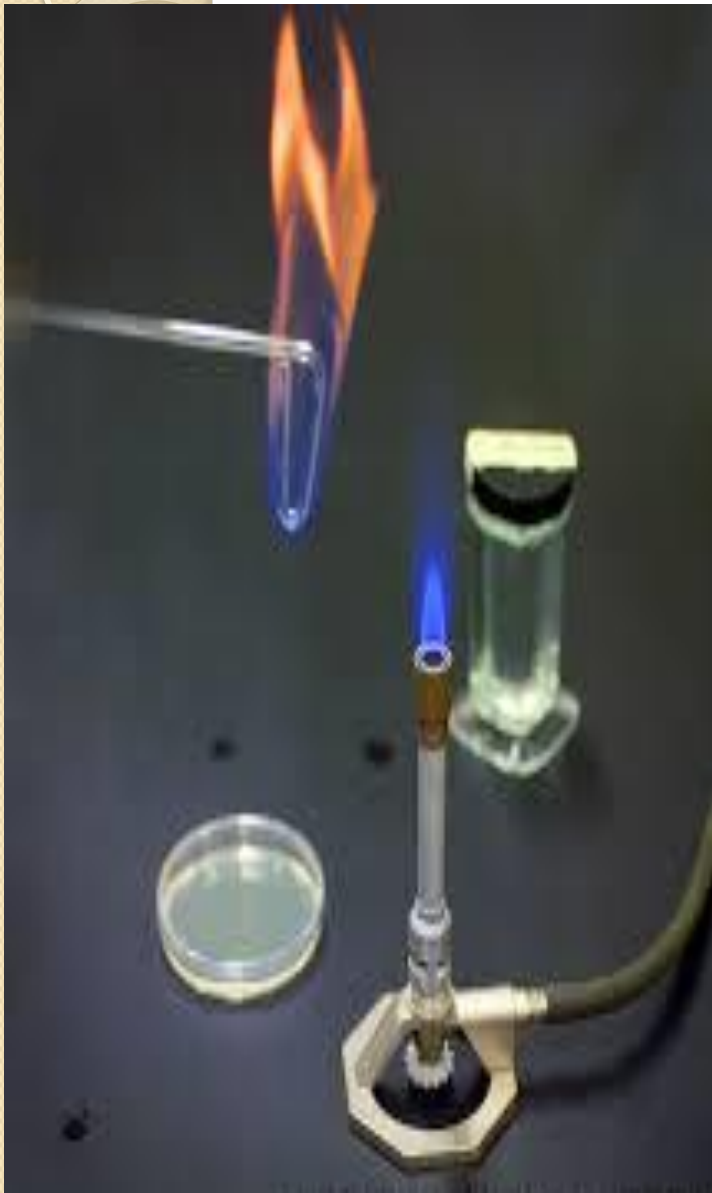
Spread Plate method

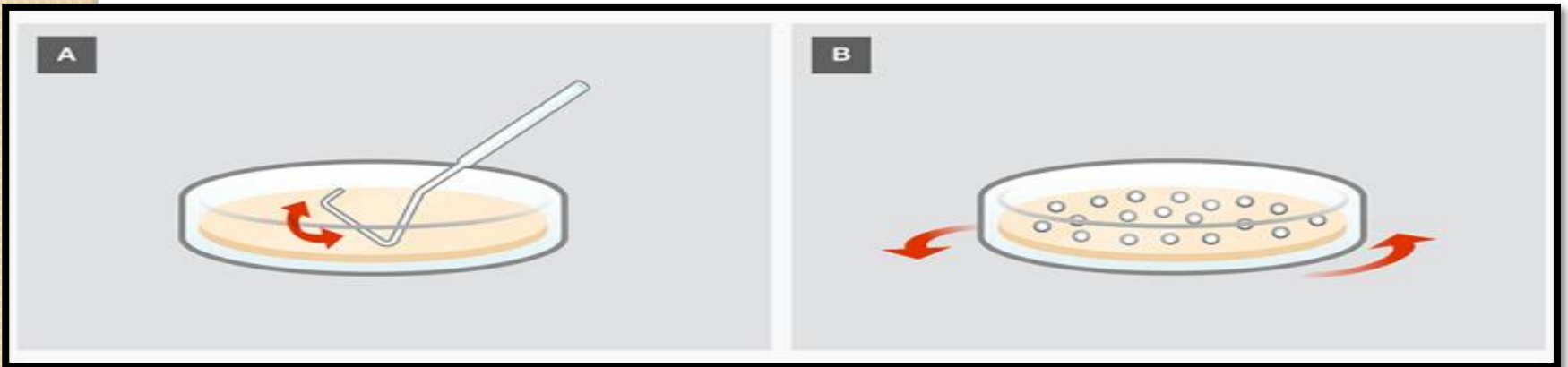
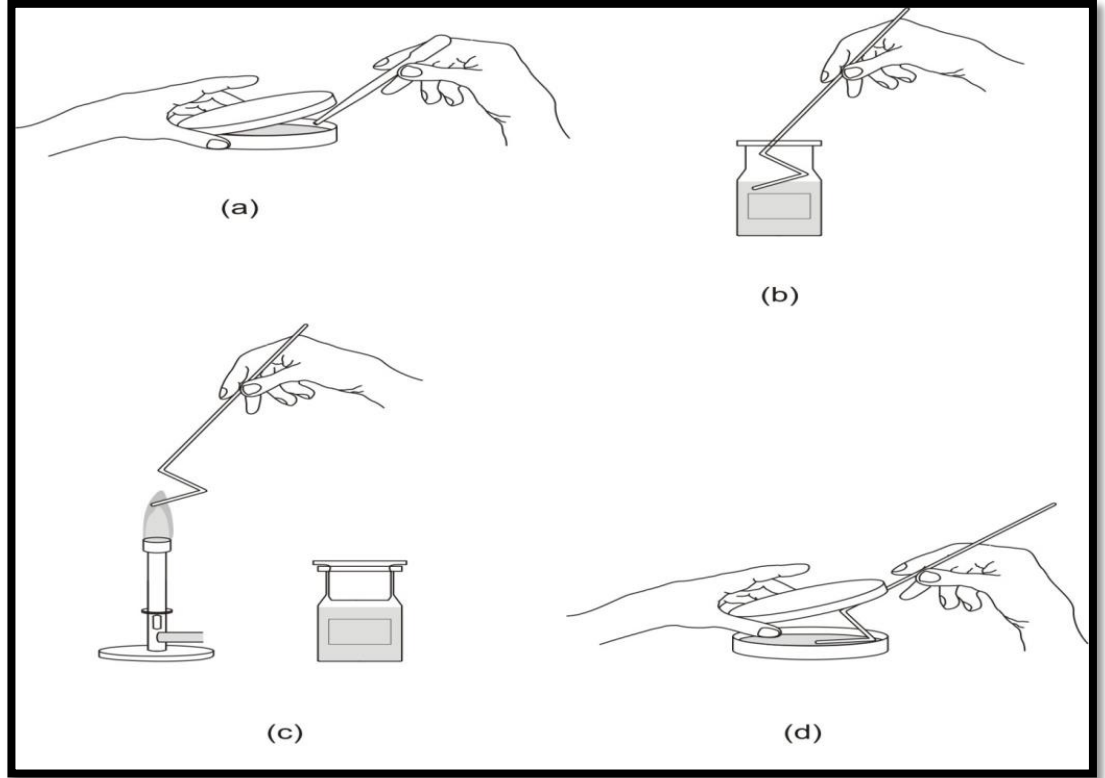
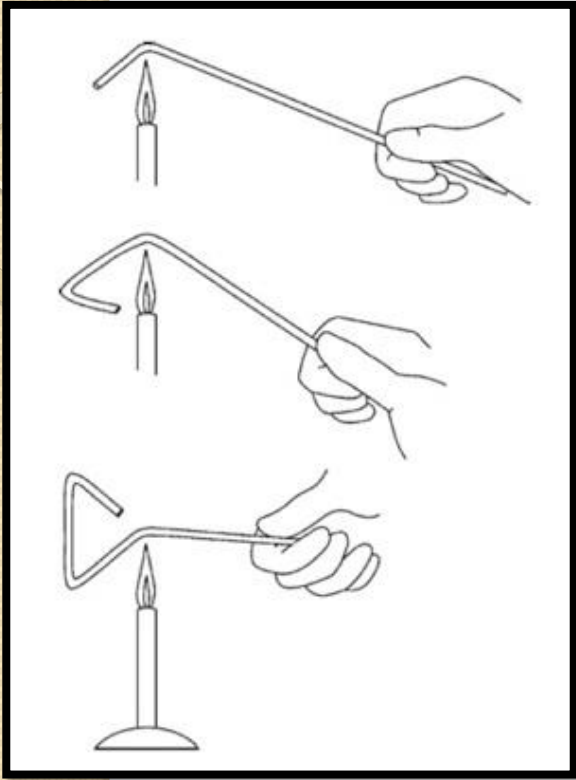
- ❖ Serially diluted specimen spread over the solidified agar media plates as a thin layer with the help of a sterile L-shape glass rod (Spreader)

Spread Plate Method



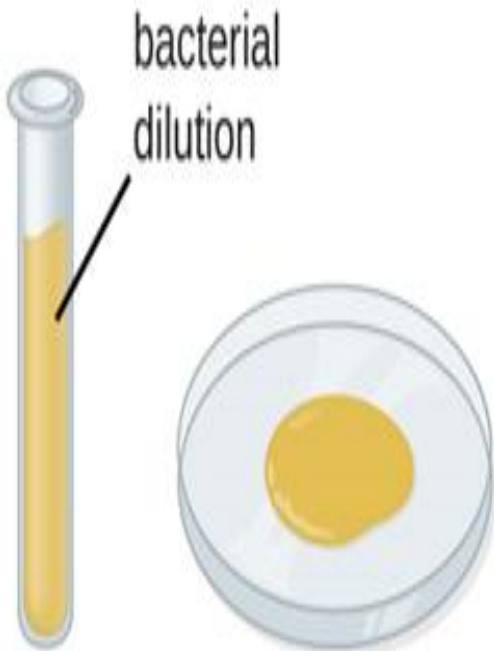
spread plate technique





Spread Plate Method

1 Sample (0.1 mL) poured onto solid medium



2 Spread sample evenly over the surface



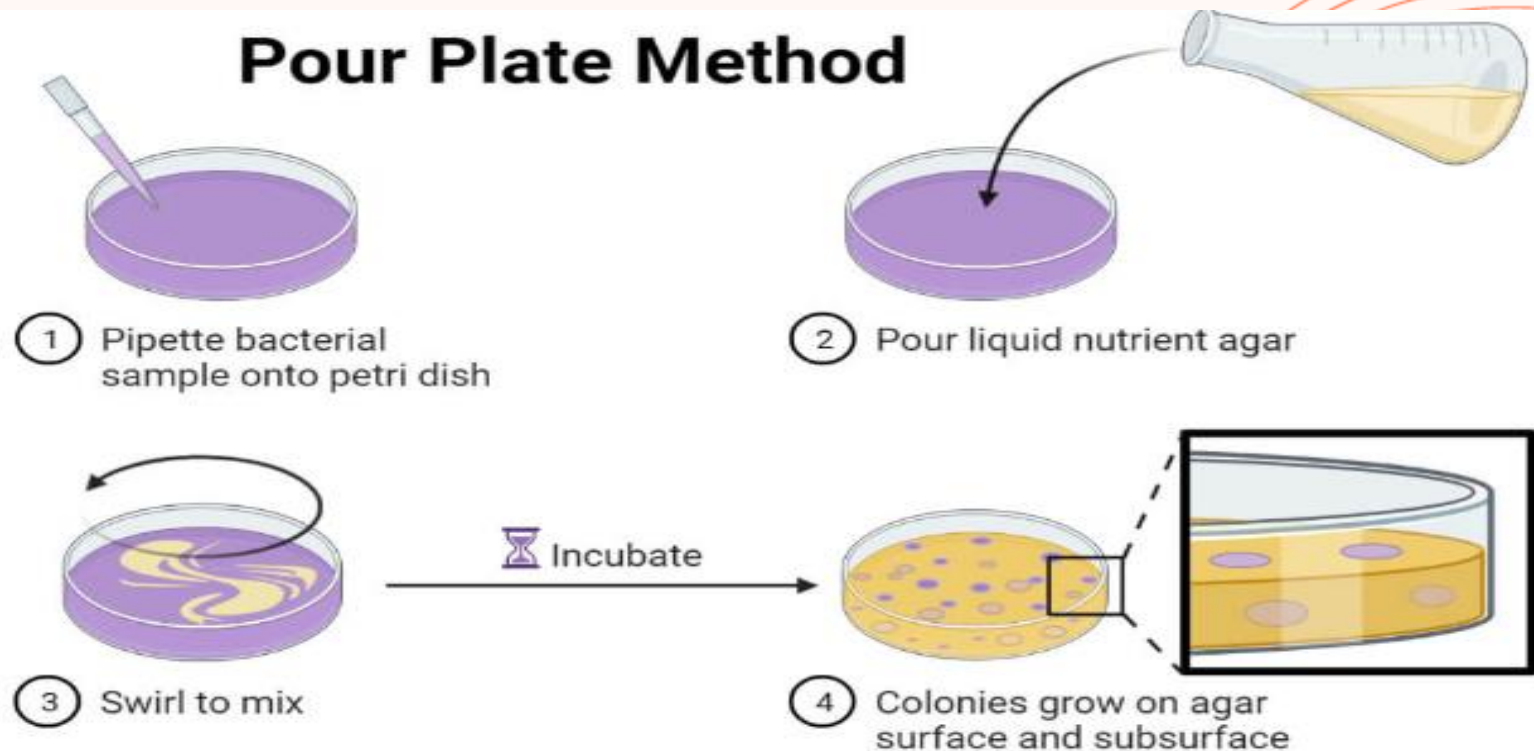
3 Plate incubated until bacterial colonies grow on the surface of the medium



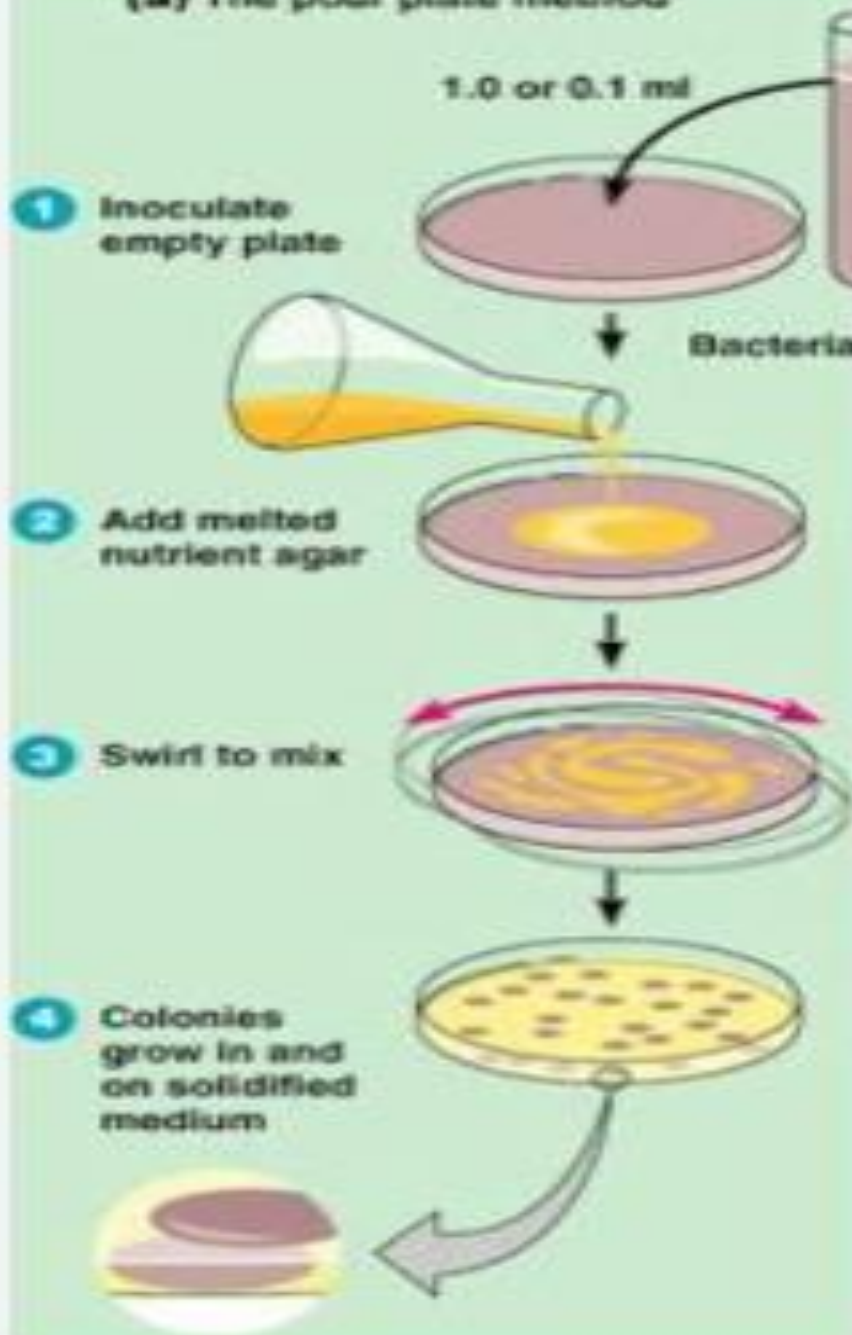


Pour Plate Method

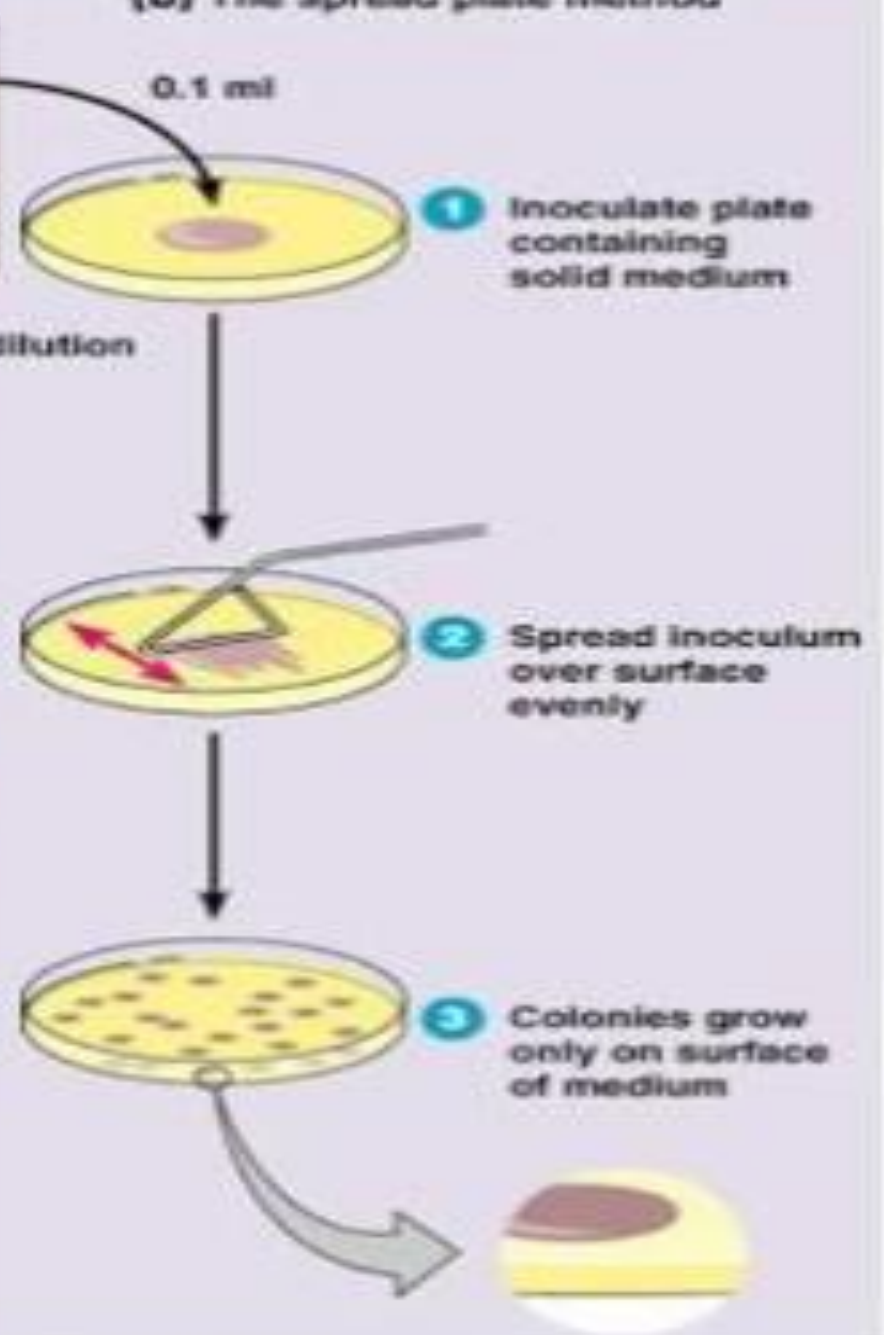
- ❖ Inoculum from a broth/sample placed in centre of sterile Petri dish using a sterile pipette
- ❖ Molten cooled agar then poured into the Petri dish containing the inoculum, mixed well and allowed to solidify
- ❖ After incubation, discrete bacterial colonies found growing both on the agar and in the agar



(a) The pour plate method



(b) The spread plate method



- **What are dilutions in microbiology?**
- **Dilution** is the process of making a solution weaker or less concentrated.
- In microbiology, serial dilutions (log dilutions) are used to decrease a bacterial concentration to a required concentration for a specific test method, or to a concentration which is easier to count when plated to an agar plate

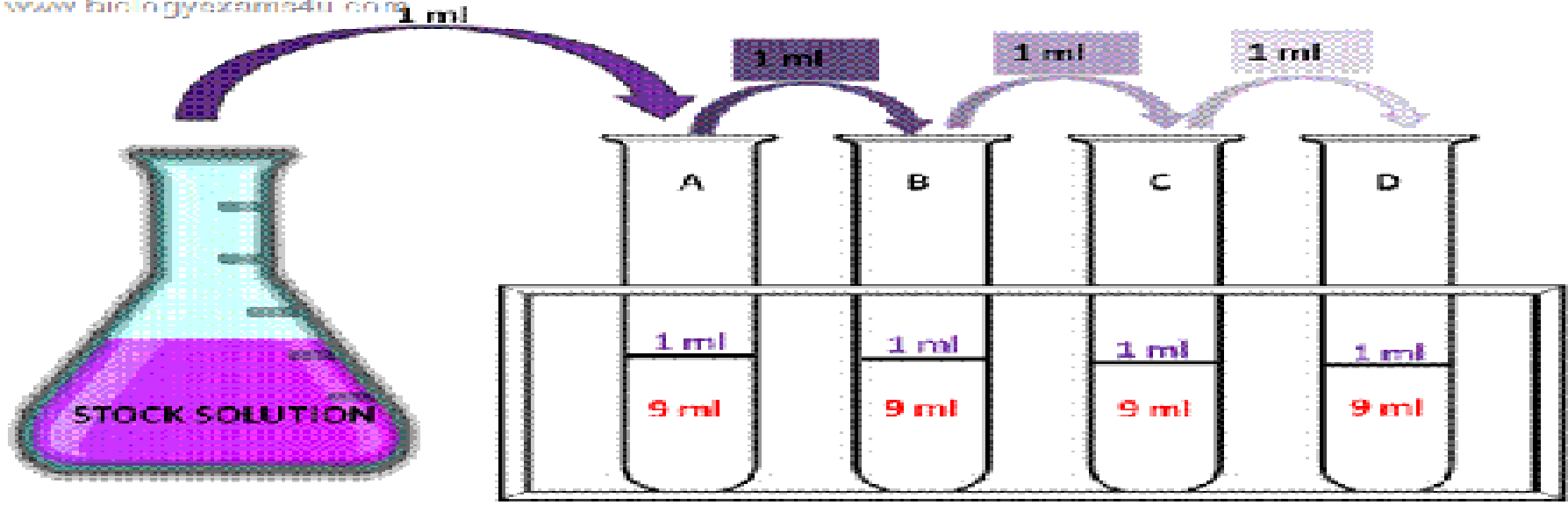
Serial Dilution Protocol

It is a method of diluting a stock solution where concentration decreases by the same quantity in each successive step.

Materials required: stock solution, test tubes, pipettes, beaker, and distilled water.

Procedure:

1. Take 4 test tubes label A, B, C and D
2. Pour 9 ml of distilled water to these test tubes
3. Transfer 1ml of stock solution to the test-tube labeled A and mix well
4. Transfer 1 ml of solution from test tube A to test tube B and mix well
5. Transfer 1 ml of solution from test tube B to test tube C and mix well
6. Transfer 1 ml of solution from test tube C to test tube D and mix well and continue up to the required dilution. Here we are making 10^{-4} dilution.



9 ml = distilled water
1 ml = stock solution

1/10th
dilution
or .1 or
10⁻¹
dilution

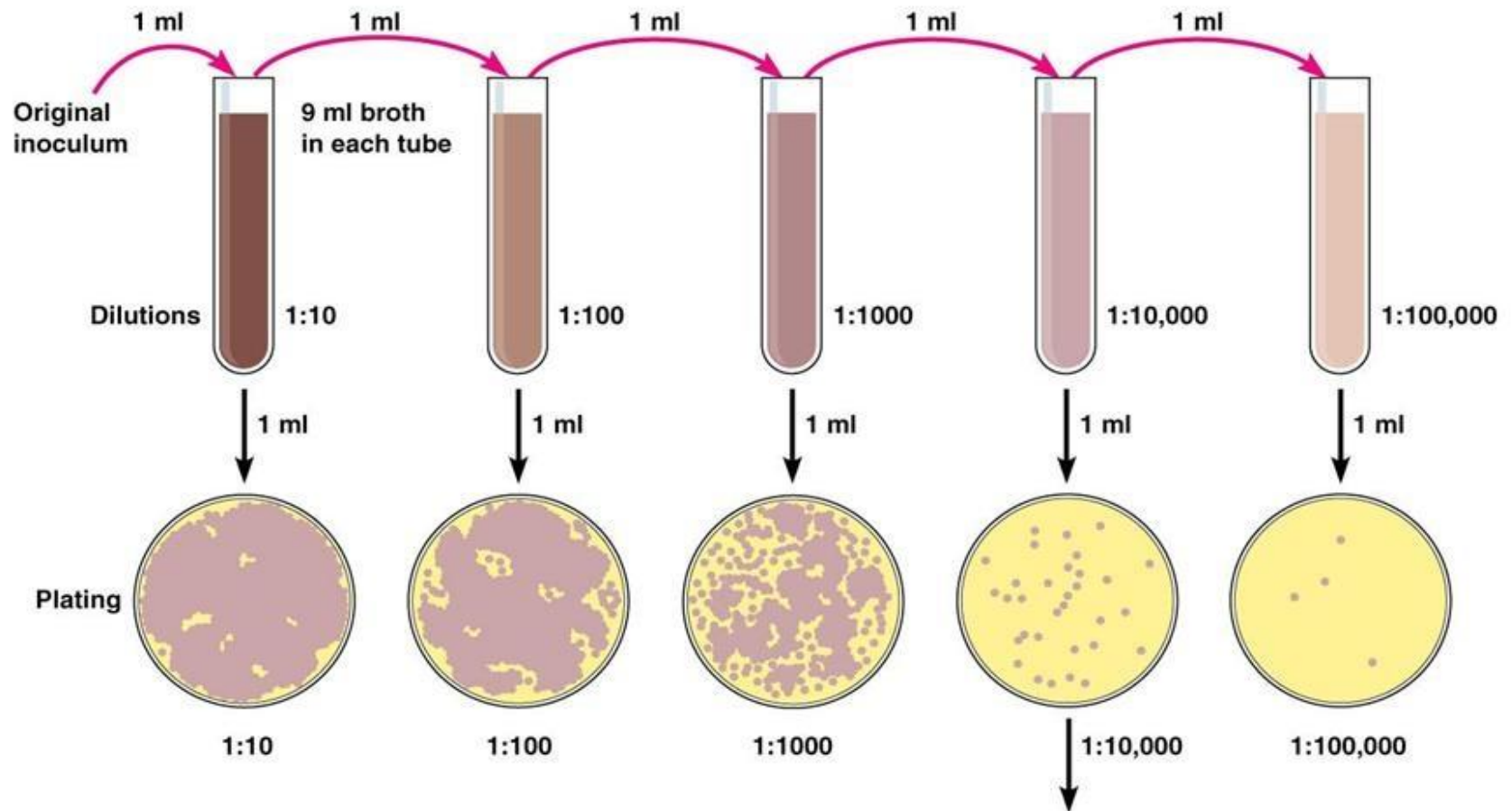
1/100th
dilution
or .01 or
10⁻²
dilution

1/1000th
dilution
or .001 or
10⁻³
dilution

1/10000th
dilution
or .0001
or 10⁻⁴
dilution

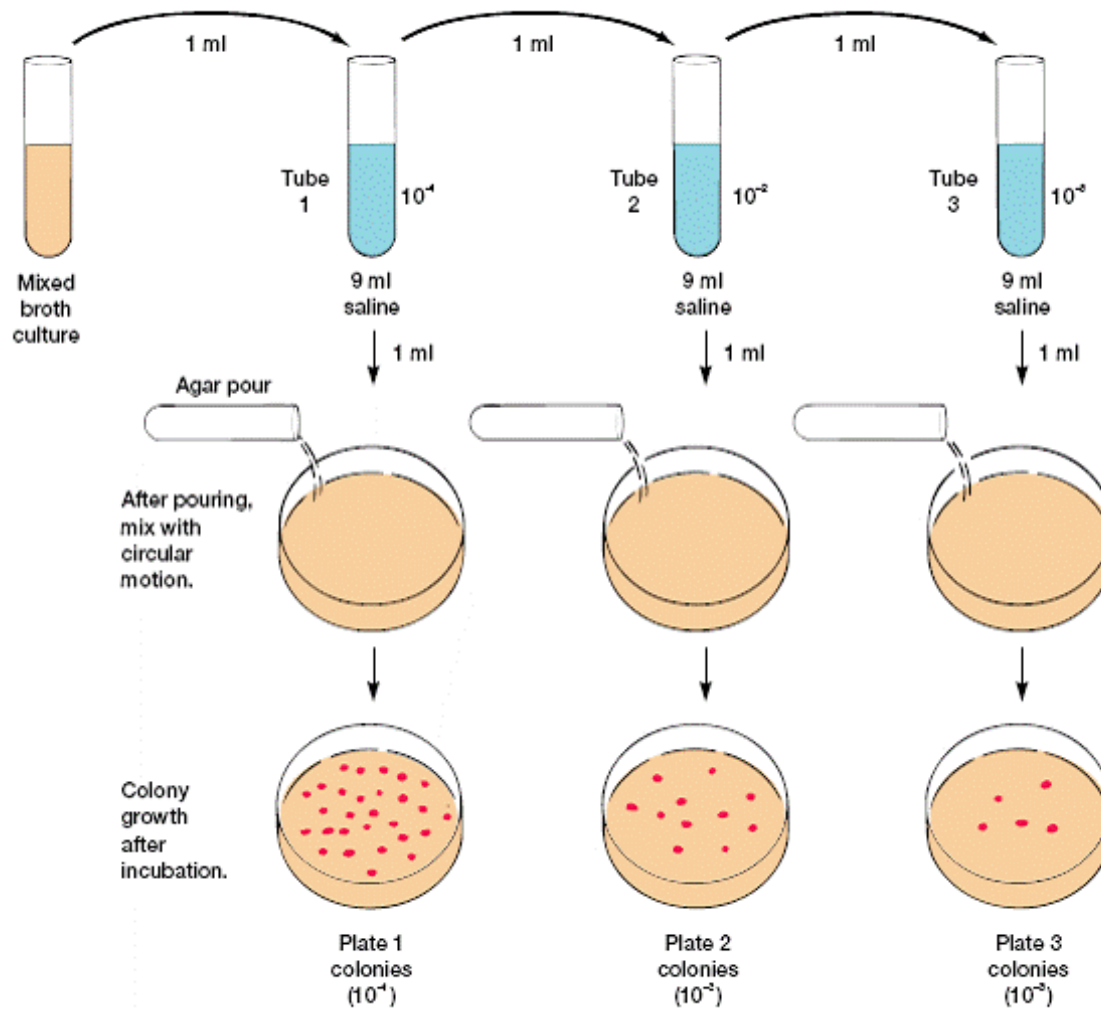
Inference:

- Test tube A has 10 times dilution or 1/10th or 10⁻¹ dilution of the stock solution.
- Test tube B has 100 times dilution or 1/100th or 10⁻² dilution
- Test tube C has 1000 times dilution or 1/1000th or 10⁻³ dilution
- Test tube D has 10000 times dilution or 1/10000th or 10⁻⁴ dilution.



Calculation: Number of colonies on plate \times reciprocal of dilution of sample = number of bacteria/ml
 (For example, if 32 colonies are on a plate of $1/10,000$ dilution, then the count is $32 \times 10,000 = 320,000$ bacteria/ml in sample.)

pour plate technique



A close-up photograph of a petri dish held by a hand wearing a blue nitrile glove. The petri dish contains a bacterial culture on a red agar medium, showing numerous small, circular, reddish-brown colonies. In the background, there are several glass bottles containing liquids of various colors: yellow, red, and green. The text "Thank you for listening" is overlaid in a black, cursive font on the petri dish.

*Thank you for
listening*