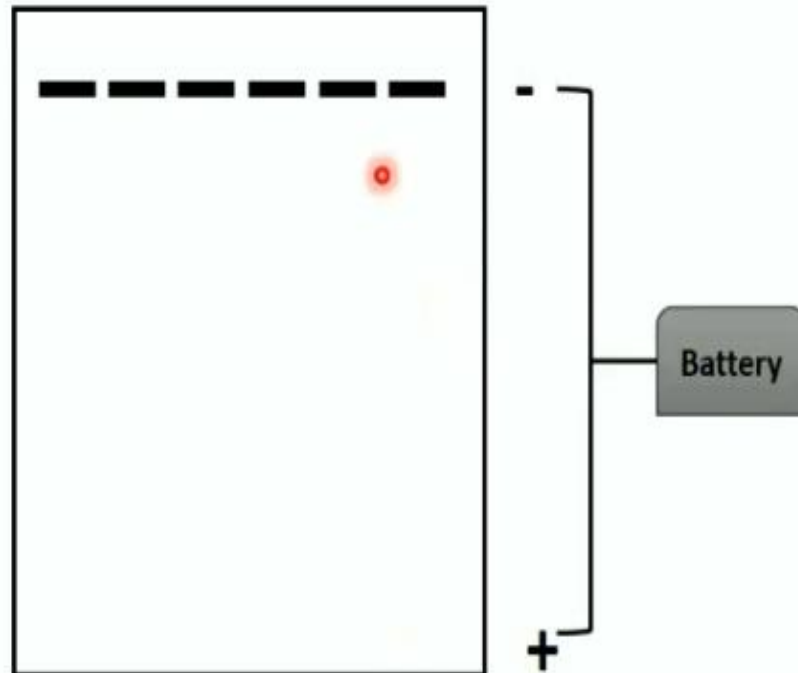


Pulsed Field Gel Electrophoresis(PFGE)

By

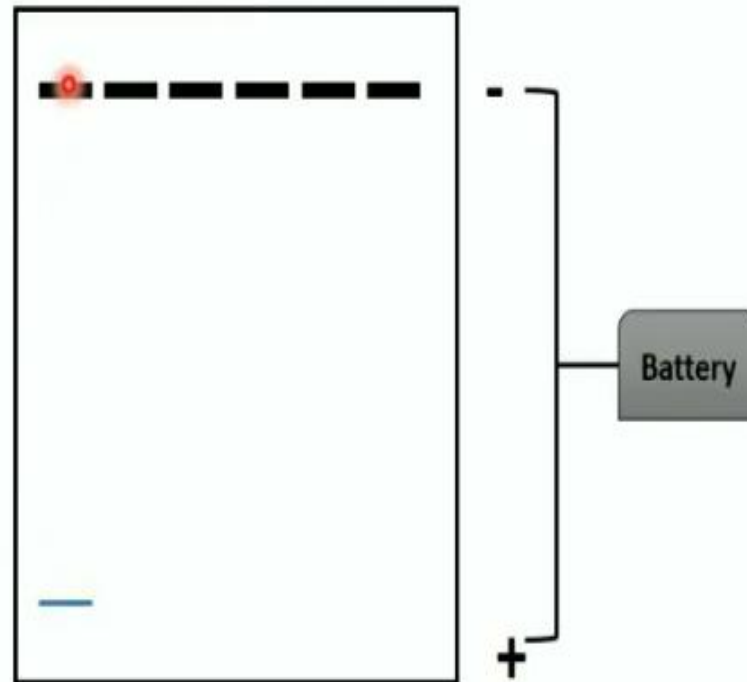
Assist. Prof. Dr. Ali Aldeewan

What is PFGE?



What is PFGE?

- Is a technique developed from agarose gel electrophoresis
- Used to separate large fragments of DNA molecules

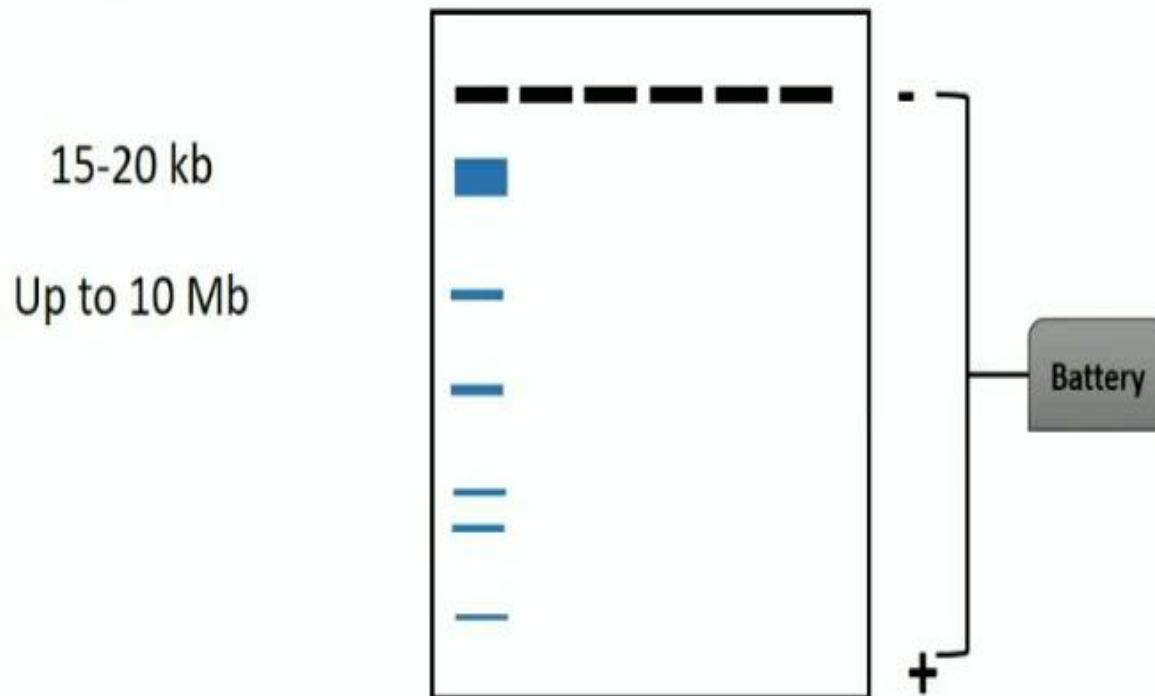


PFGE(Pulsed Field Gel Electrophoresis)

- Pulsed field gel electrophoresis is a technique used for the separation of large DNA molecules by applying to a gel matrix an electric field that periodically changes direction.
 - 1ST developed in 1980 by Schwartz and Cantor.
 - **Principles of PFGE technology**
 - It uses specially designed electrophoretic apparatus to separate large DNA fragments ranging from 40 kb to 2000 kb.
-

What is PFGE?

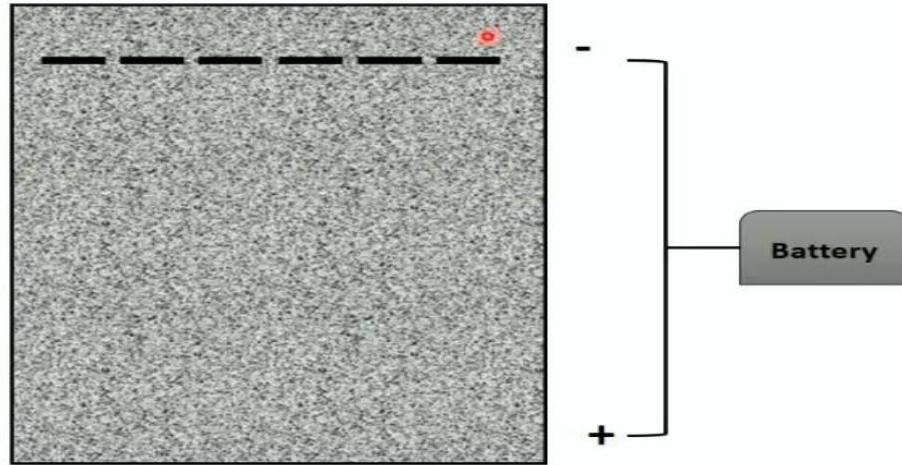
- Is a technique developed from agarose gel electrophoresis
- Used to separate large fragments of DNA molecules



What Modifications?

1- ~~the~~ concentration of the agarose solution

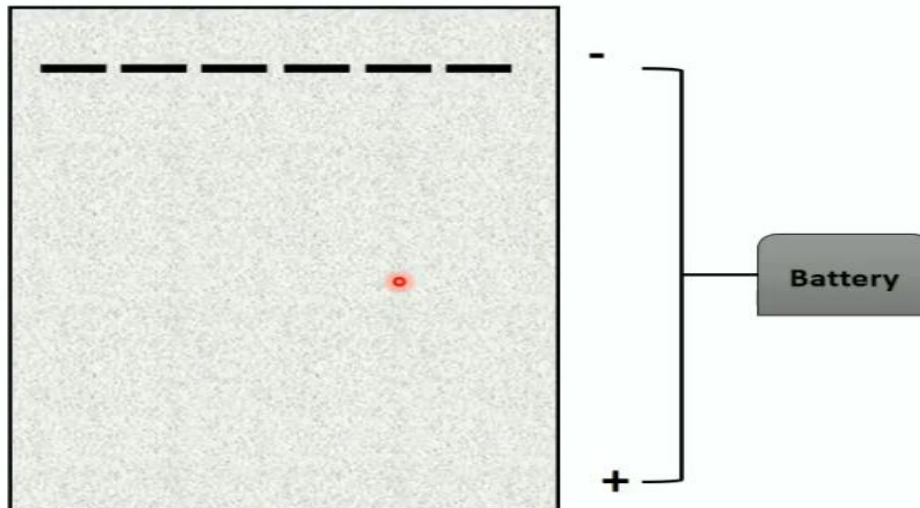
(2%)



What Modifications?

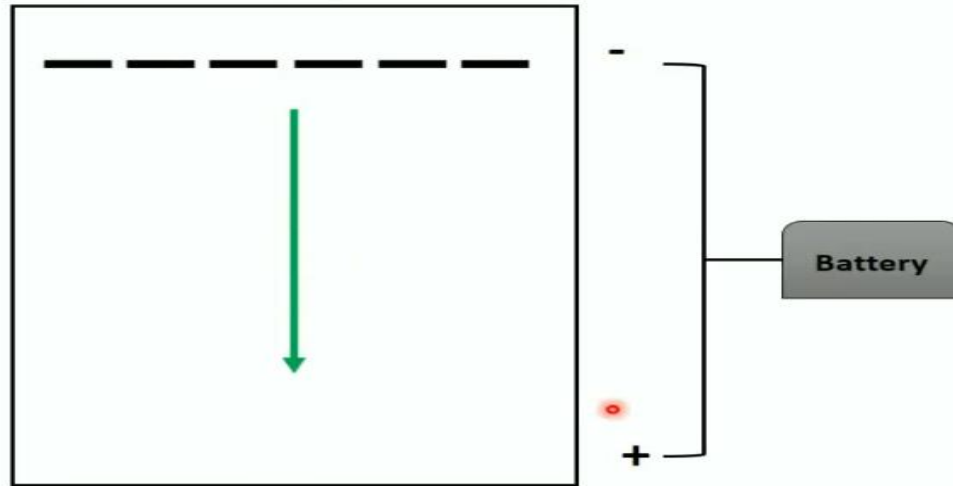
1- Low concentration of the agarose solution

(1%)



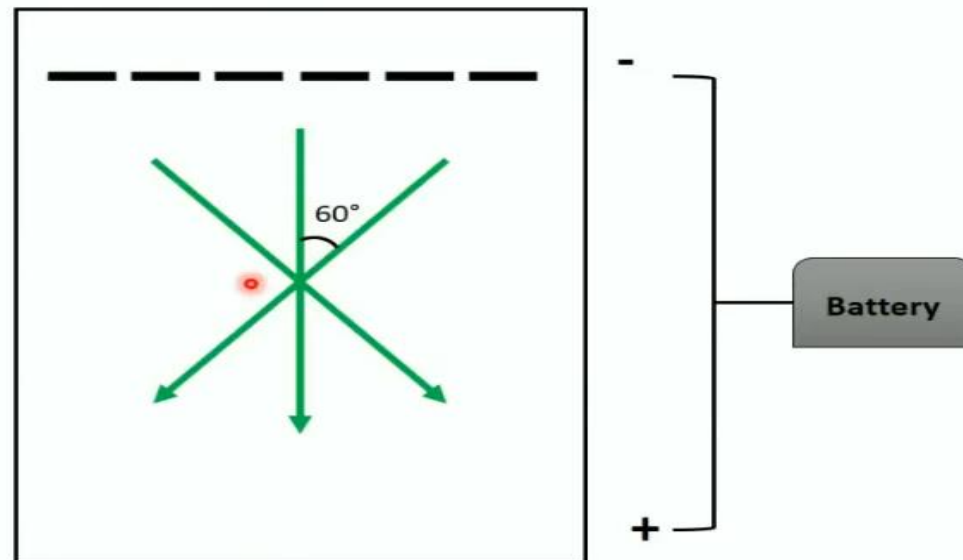
What Modifications?

2- Change the direction of the electrical field



What Modifications?

2- Change the direction of the electrical field



Uses

- strain characterisation
- Gene mapping in microbes and mammalian cells .
- Monitoring and evaluating different microorganisms in clinical samples and in soil and water.
- A reliable and standard method in vaccine preparation
- Epidemiological studies
- strain development
- separation of molecules having $>50\text{kb}$ molecular weight.
- produce DNA fingerprint for a bacterial isolate.

DEVELOPMENT OF PFGE

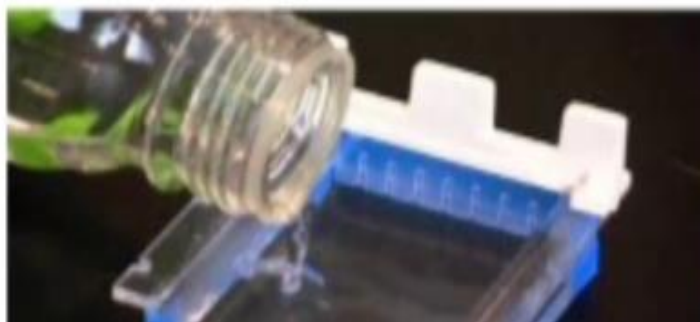
Conventional electrophoresis.....>PFGE

PFGE Procedure

- I. Cell lysis and release of intact chromosomal DNA
- II. Restriction Endonuclease digestion of chromosomal DNA
- III. Separation of large DNA fragments
- IV. Analysis of DNA fragment length polymorphism

1. Cell lysis and release of intact chromosomal DNA

- Overnight culture of the bacterial isolate (10^9 cells /ml)
- Add detergents and enzymes to the bacterial suspension
- Mix bacterial cells with warm agarose and pipette into plastic mold to form agarose plugs
- Wash with preheated water and TE buffer
- Agarose gel matrix keep chromosomal DNA and remove the other components.



2. Restriction Endonuclease digestion of chromosomal DNA

- R.E ENZYME(molecular scissors): to cleave ds DNA at restriction sites.
- The choice of RE enzyme depend on bacterial sp.
- Once the recognition site is located , the enzyme catalyses the digestion of DNA at defined position.....>produce restriction fragments
- Restriction Endonuclease Sma 1 recognises CCC/GGG sequence, that cleave DNA of most gram positive bacteria.
- After digestion, plugs are cut into appropriate size>loaded onto comb teeth.....>sealed.....>placed in electrophoresis chamber.

3. Separation of large DNA fragments

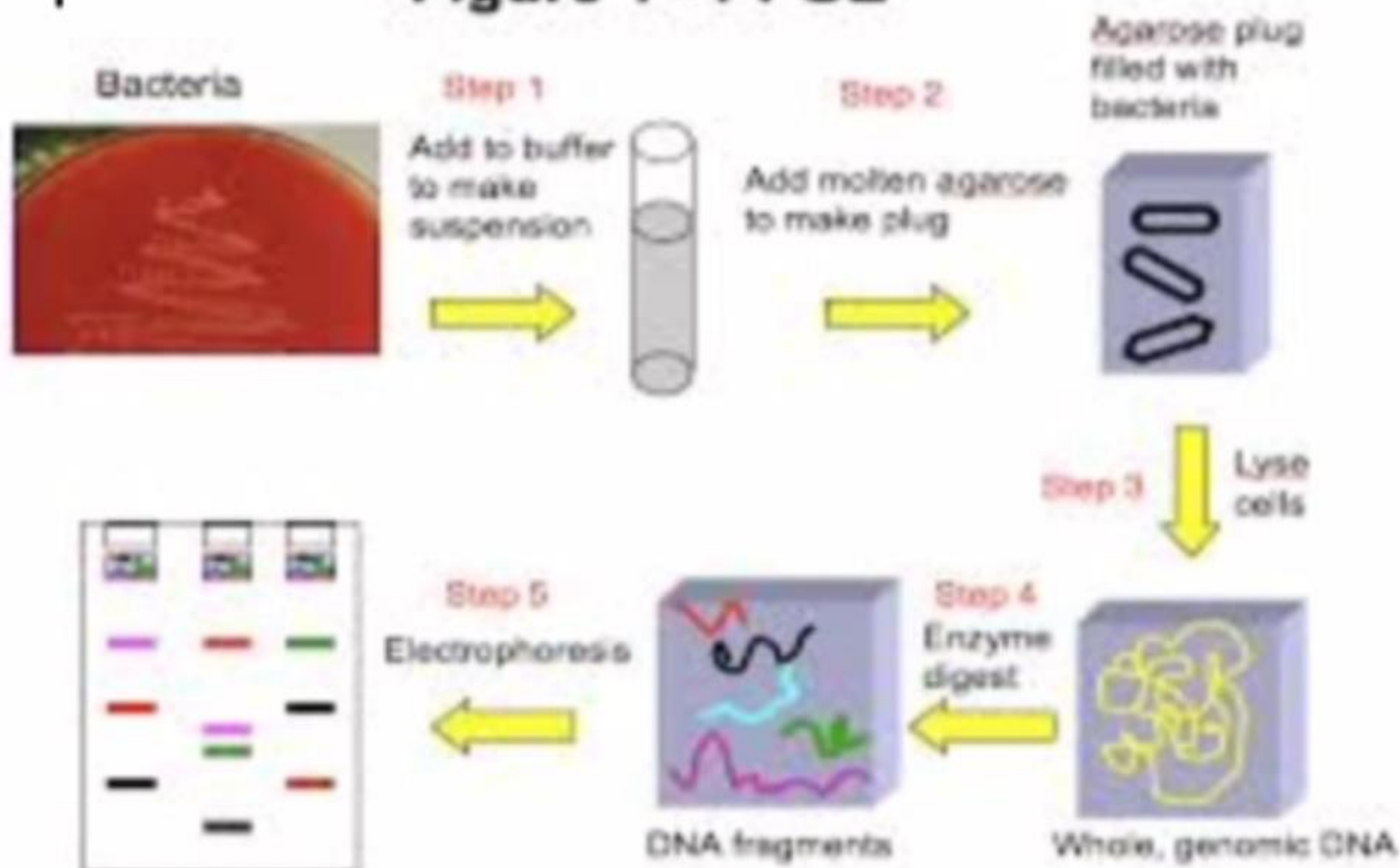
- **Resolution of DNA depend upon:**
 1. Concentration and composition of gel and buffer
 2. Temperature
 3. Pulsed field condition (electrophoresis duration, electric field strength, pulse angle, and switching time)
- **Migration rate vary in different buffer**
- **Agarose concentration: 0.8-1%**
- **Voltage 6 V /cm**

- **Problems associated with the detection of band:**
 1. DNA digestion in gel
 2. Incomplete digestion by Restriction endonuclease
 3. Incorrect electrophoresis condition

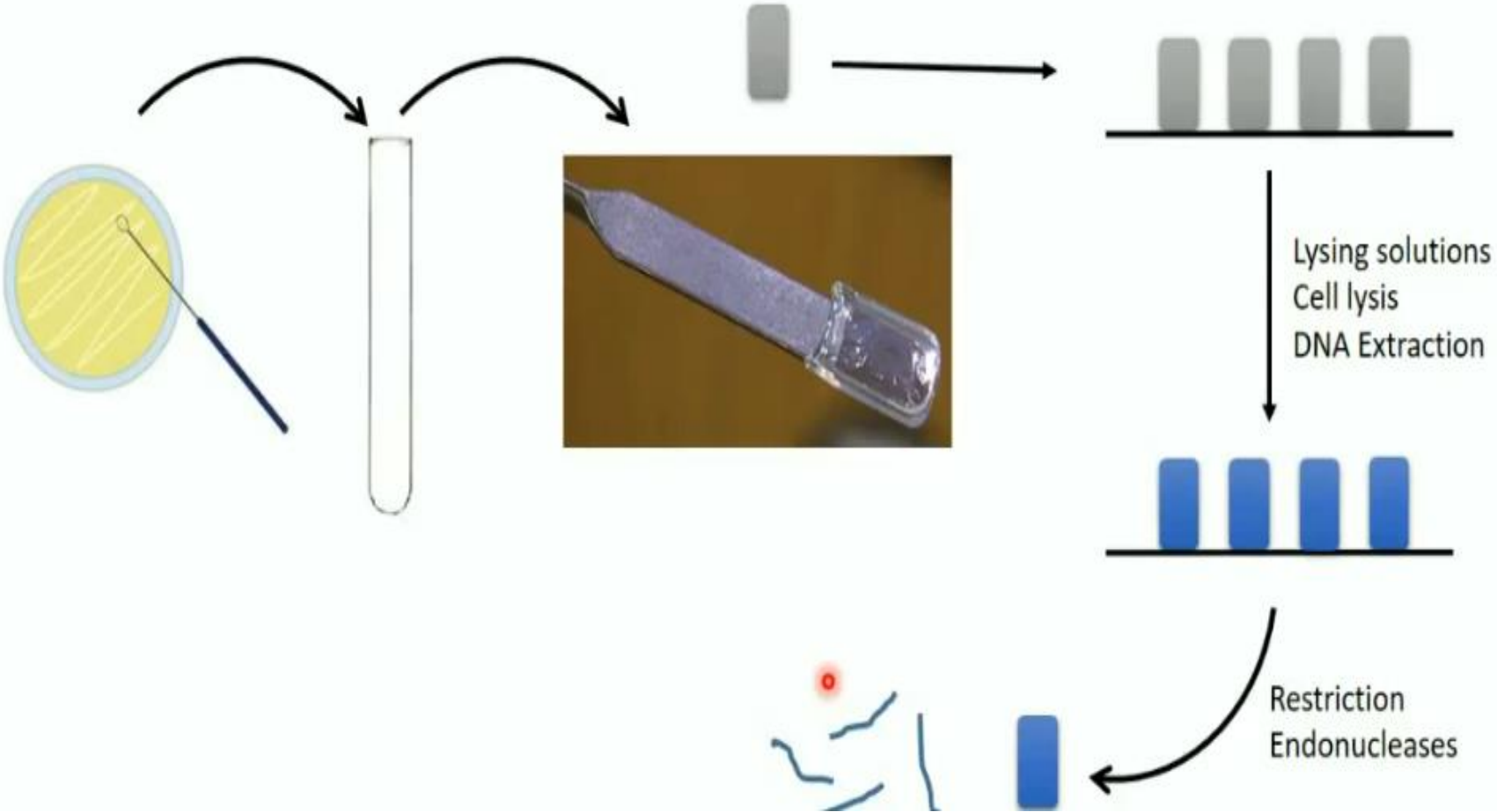
4. Analysis of DNA fragment length polymorphism

- Visualisation: staining with **ethidium bromide**
 - Each lane on gel represents the chromosomal pattern of one bacterial isolate.
 - Same band pattern: **indistinguishable**
 - 1-3 band difference: **closely related**
 - 4-6 band difference: **possibly related**
 - 6 or more difference: **unrelated**
 - Computerised gel scanning and data analysis:eg: **Dendrogram.**
-

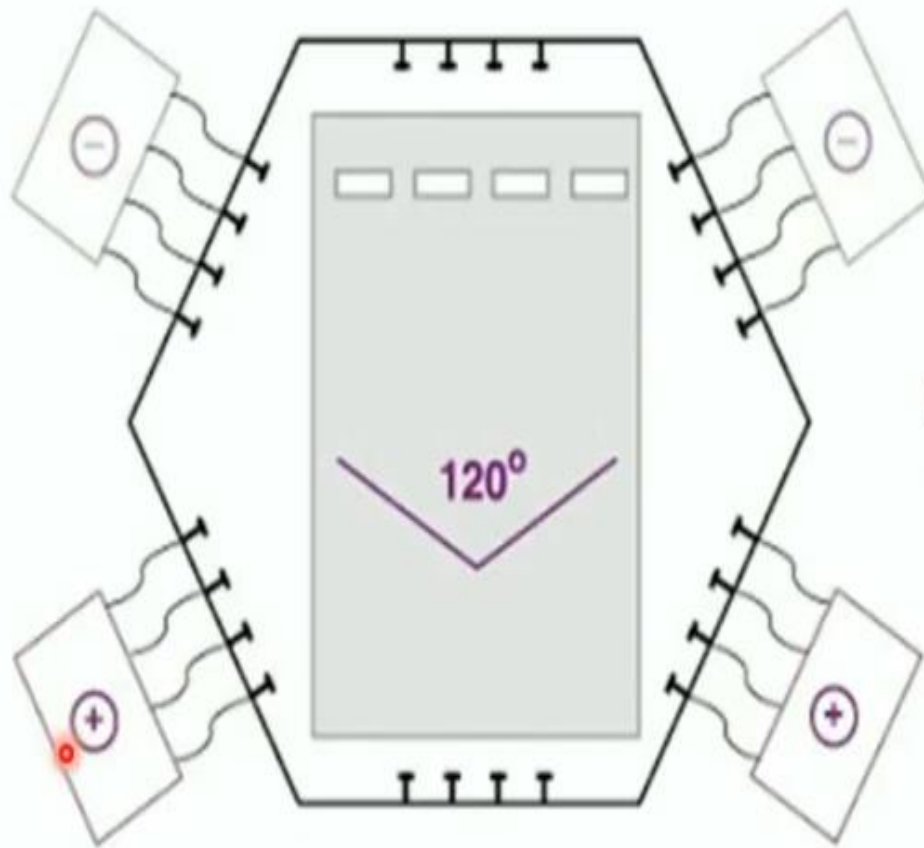
Figure 1 - PFGE



How To Perform It?

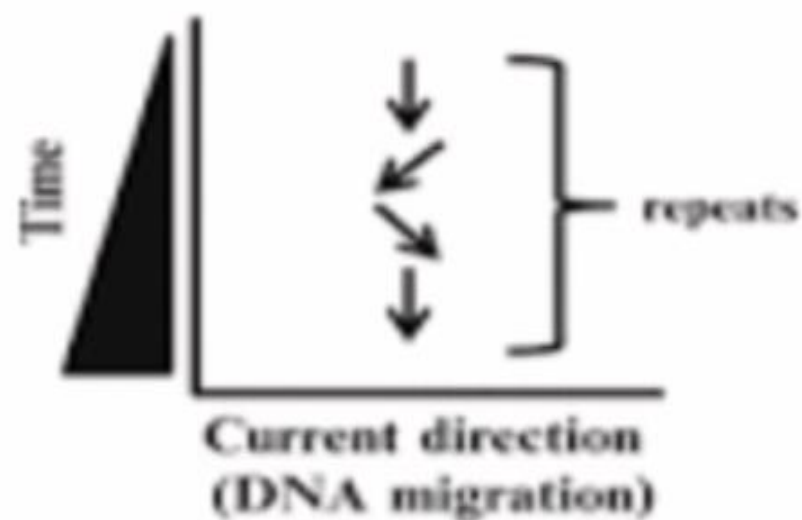
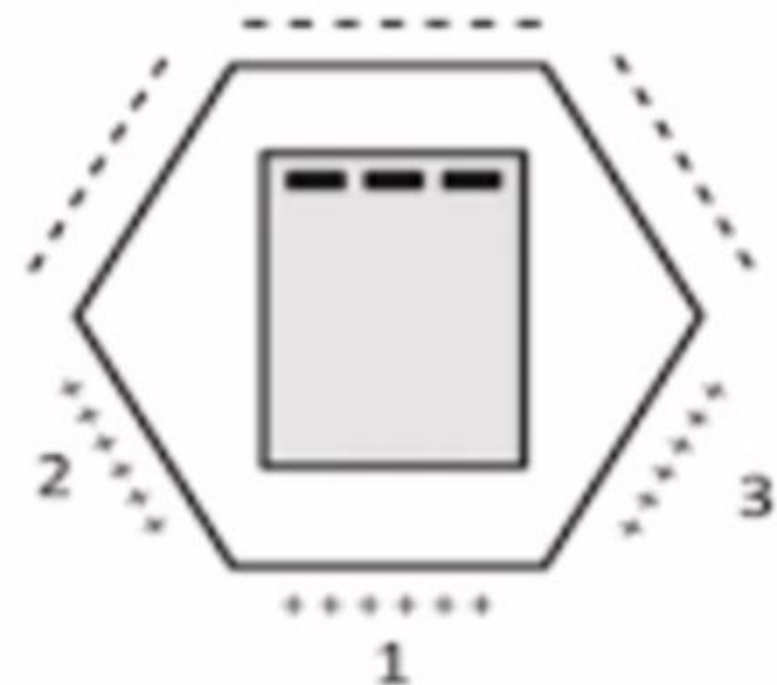


How Does It Work?



Obtuse reorientation angle

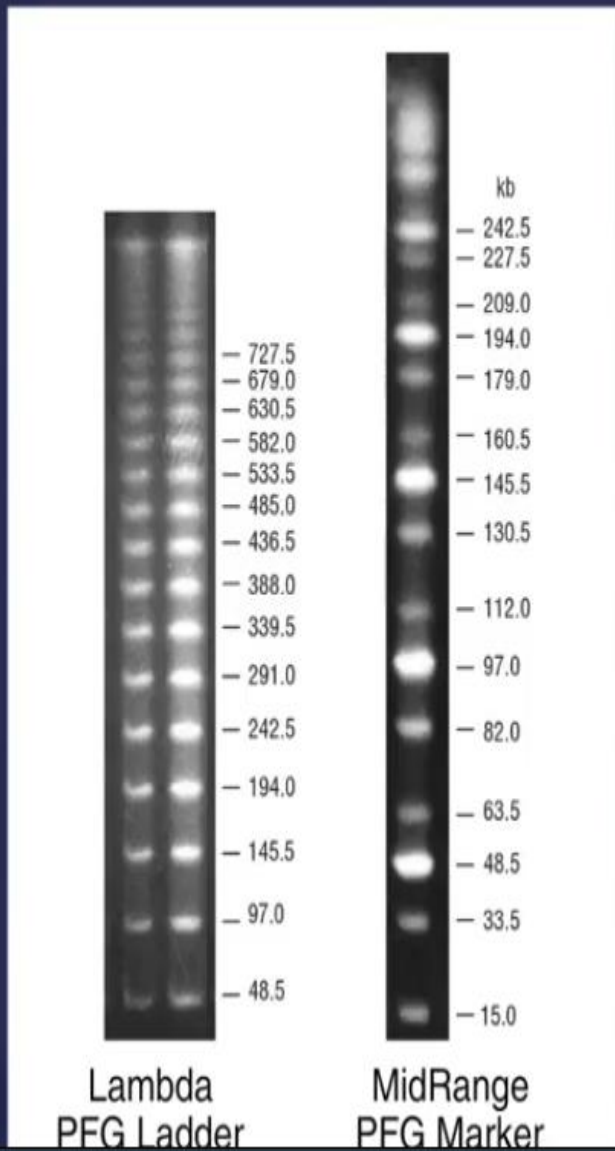
Pulse Field Gel Electrophoresis



Conventional electrophoresis



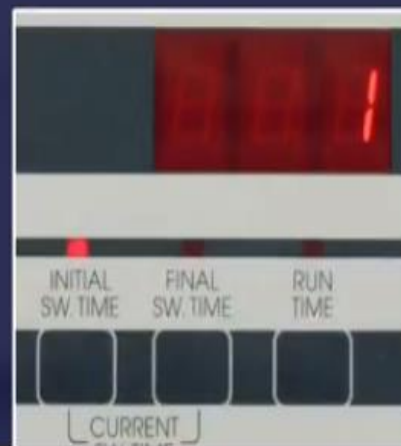
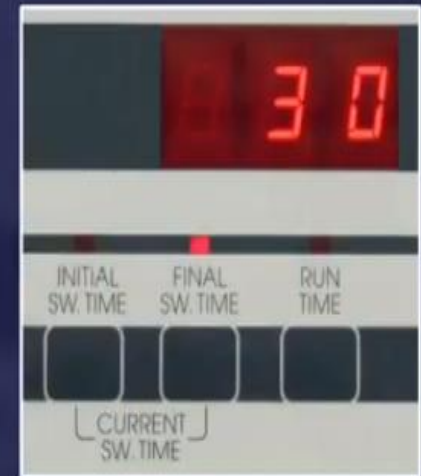
Switch Time Ramp Examples




Initial switch time



Final switch time



 Voltage

0.60

VOLTS/CM INCLUDED ANGLE ACTUAL CURRENT PAUSE START RUN

 Angle

120

/CM INCLUDED ANGLE ACTUAL CURRENT PAUSE START RUN



PFGE Separation Range

10 kb → 10 M



PFGE Separation Range

10 kb → 10 Mb

PFGE Separation Examples

Base Pairs

48,502 —
38,416 —
33,498 —
29,946 —
24,508 —
23,994 —

17,053 —
15,004 —

10,086 —

1,503 —



λ DNA-Mono Cut Mix
1.5 to 48.5 kb

Kilobases

1,900 + 1,640 —
1,120 + 1,100 —
945 —
915 —
815 —
785 —
745 —
680 —
610 —
555 —

450 —
375 —
295 —
225 —



Yeast Chromosomes
225 to 1,900 kb

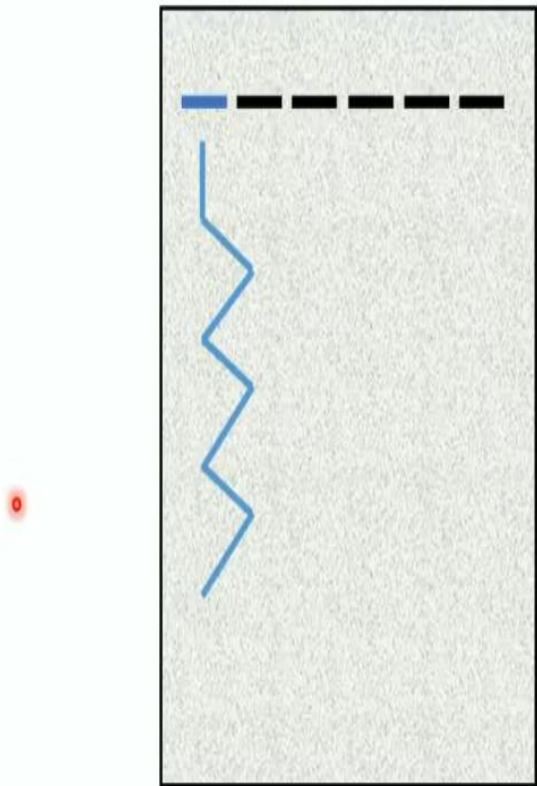
Principle of PFGE

DNA segments elongate in the presence of an electrical field

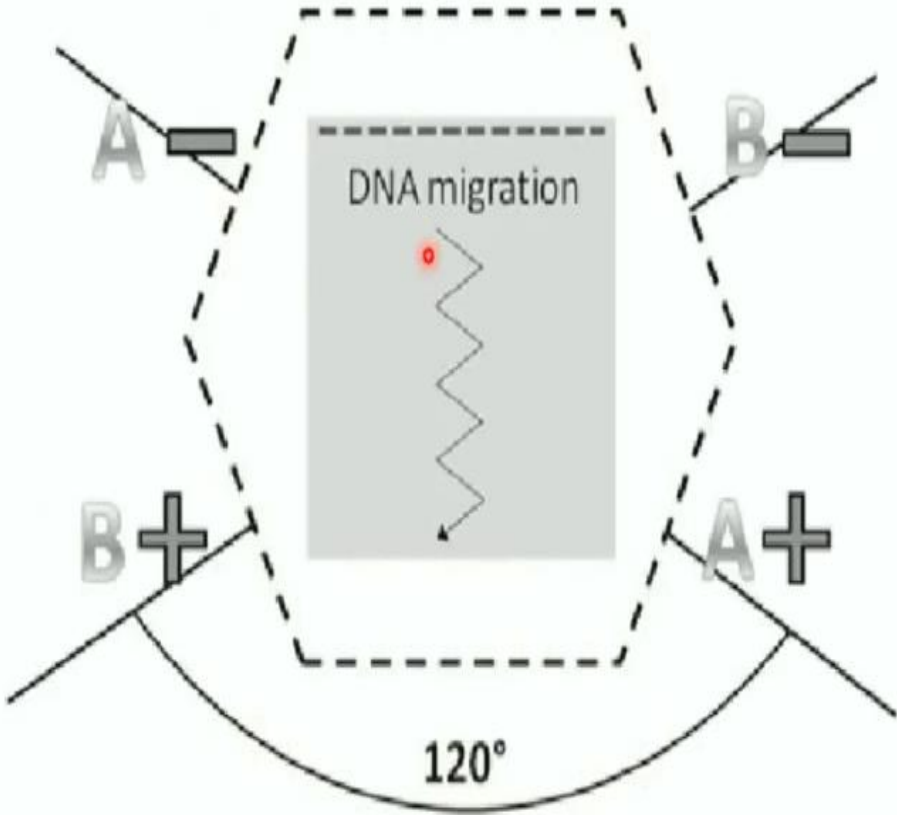
The relaxation rate depends on the size



Principle of PFGE



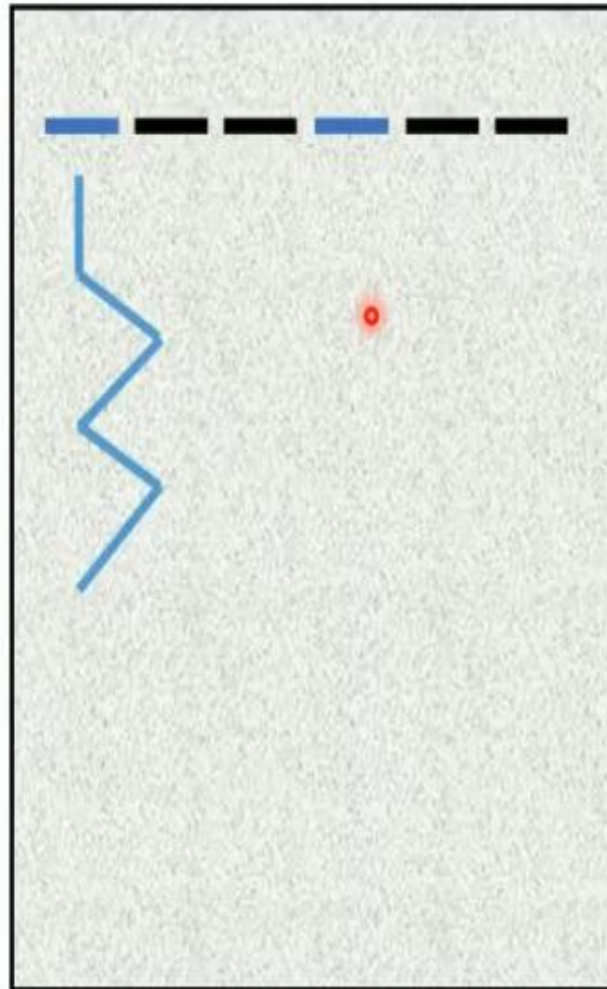
Principle of PFGE



What Important Parameters

1- Voltage (V/cm)

2- The angle:
Smaller angle (106°)
gives better resolution
with larger segments.



3- The switching time:
The longer the
switching time is, the
further the large
segments will migrate
through the gel.

4- Temperature:
DNA stays in the gel
overnight.
Pumping the running
buffer through a chiller
during the run
Switch the buffer.

PFGE performance characteristics

- Performance is measured by

1. Discriminatory power
2. Reproducibility
3. Stability
4. Typeability

- **Application to gram positive bacteria**

For molecular typing of nosocomial pathogens

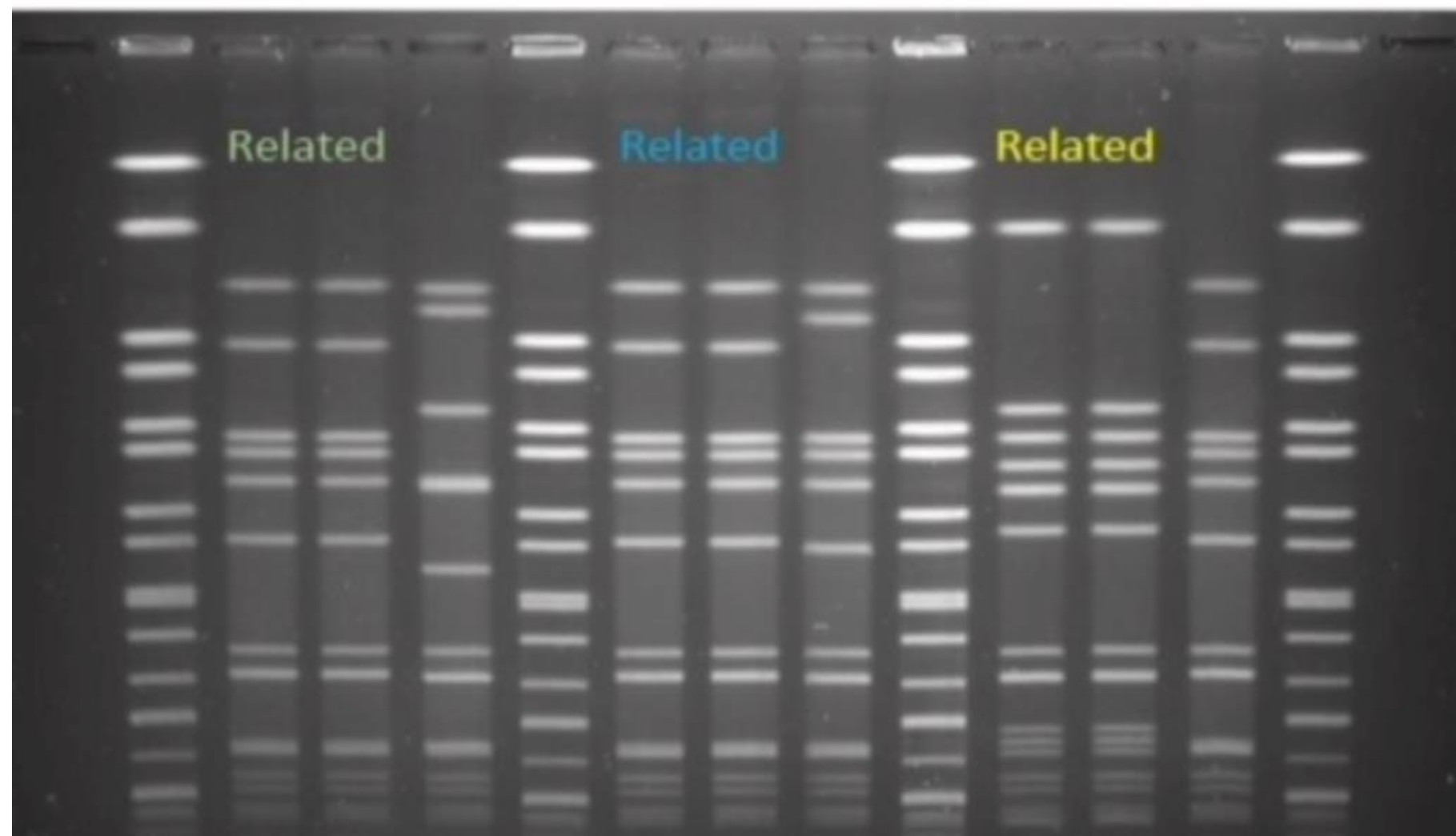
To examine genetic identity of MRSA

- **Application to gram negative bacteria**

To investigate epidemiological relatedness of strains of gram negative bacteria.

Epidemiological investigations

Thank you all.....





How to Prepare and Load a Pulse Field Gel Marker.mp4