Laboratory diagnosis of veterinary important agents from genera Brucella and Campylobacter

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Brucella in animals

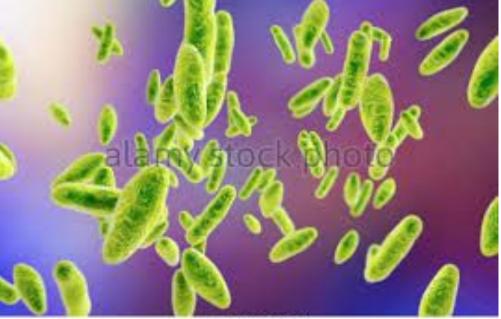
- Brucellosis is a zoonotic disease caused by Gram-negative coccobacilli of the genus *Brucella*.
- In livestock, the disease results in significant economic losses due to reproductive impairment caused by abortion, stillbirth or weak calves and neonatal mortality, infertility.
- The genus *Brucella* is currently classified into 12 known species, according to basic differences in pathogenicity and host preference.
- Each one may infect different host species, but each *Brucella* species has a preference for its host species.

• Epidemiological features of Brucella species.

Species	Natural host	Prevalent region	Reported human cases			
B. melitensis	Sheep, goats	Mediterranean littoral, Arabian Peninsula, Latin America	Several cases			
B. abortus	Cattle	Asian countries, Europe	Several cases			
B. suis	Pigs	Latin America, Southern China, Southeast Asia, Europe	Several cases (biovar 1) Rare cases			
B. canis	Dogs	Argentina, Brazil, China, Czech Republic, Germany, Japan, Madagascar, Mexico, Papua New Guinca, Peru, Philippines				
B. ovis	Sheep	Argentina, Chile, France, Germany, South Africa, USA, Spain, countries of the former Soviet Union	No human cases			
B. neotomae	Rodents (desert wood rats)	United States	Two human cases			
B. microti	Wild voles	North Europe	No human cases			
B. ceti	Marine mammals	Mainly Northern Hemisphere	One laboratory infection			
B. pinnipedialis	Marine mammals	Mainly Northern Hemisphere	No human cases			
B. inopinata	Unknown		Prosthetic breast implant infection (one human case)			
B. papionis	Baboons	141	No human cases			
B. vulpis	Red foxes	Austria	No human cases			

Laboratory diagnosis

• Extreme care must be applied when working with *brucellae* because humans are highly susceptible to brucellosis and laboratory infections could happen.



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Laboratory diagnosis

- Phenotyping identification, through biochemical profiling, is not conclusive for all *Brucella* species due to the high similarity between them.
- There is little antigenic variation among *Brucella* spp., therefore the differentiation of species and strains is based on approximately 25 biological and physiological characteristics (phenotype)
- Therefore a combination of the following techniques are required:
 - 1. <u>Classical microbiological identification</u>
 - 2. <u>Serology</u>
 - 3. <u>Molecular techniques</u>.

Samples for *Brucella* spp. For isolation, serology, and molecular techniques:

- Fetal membranes
- fetal organs such as the lungs, bronchial, lymph nodes, spleen and liver, as well as fetal gastric contents.
- Milk, vaginal secretions and semen.
- Blood and serum

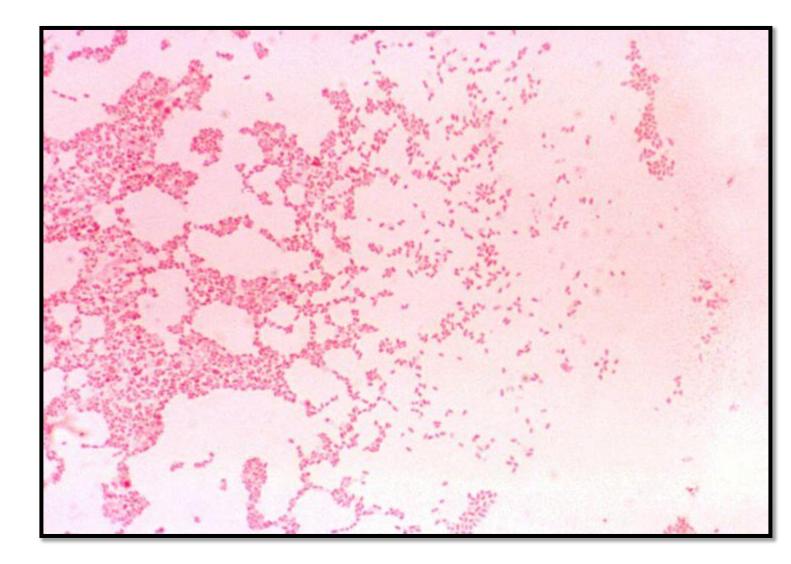
1- Classical microbiological identification

General microbial features for *Brucellae* :

- 1. Aerobic
- 2. Gram-negative cocci, coccobacilli or short rods of 0.5-0.7 by 0.6-1.5 μm in size
- 3. Usually individual and less frequently are found in pairs, short chains, or small groups.
- 4. They are non-motile and do not produce flagella.
- 5. Multiplication is slow at the optimum temperature of 37°C but the organism can grow under temperatures ranging from 20 oC to 40 oC

- Brucella spp. are fastidious bacteria that need rich culture medium to support adequate growth. The growth occurs on Brucella agar, MacConkey Agar, Trypticase Soy agar, Sheep Blood agar and Standard Nutrient agar at 25–42°C.
- 6. Colonies on media are transparent, convex and have an entire edge. They are usually small (0.5–1.0 mm after 2–3 days of incubation of a fresh inoculum).
- 7. Brucella strains are catalase positive and superoxide dismutase positive, most of them are also oxidase positive.
- 8. Many strains require supplementary CO2 for growth.

• <u>Brucella melitensis</u>, gram, negative, coccobacillus,



• Brucella abortus,

• Grown on blood agar



Example : Phenotypic features of *Brucella* species and biotypes, and the tests for their identification and differentiation

		Biotype	: CO,	H,S		NA - 2017-0		iochemic		10 -51 						Lysis by phage		
	Species				Urease	Oxidase	Catalase	Citrate Nitr	Nitrate	Growth on dyes **		Agglutination in serat			(RTD)Tb#	Motility	Acriflavin	
	Species	Biotype								Thioni a	n *** b	Basic fuchsin	Anti -A	Anti -M	Anti -R	Tb 10 000 x RTD	Methily	test
lot for memorization	B. melitensis	L	-	2	+	+	+	-	+	-	+	+	-	+	-		-	-
		2	2	2	+	+	+	-	+	-	+	+	+	5	<i>©</i>	-		2
		3	-	23	+	+	+	2	+	-	+	+	+	+	22	14	-	2
	R. abortus	1	(+)	+	+	+	+	-	+	-	-	+	+		-3	+	-	-
		2	(+)	+	+	+	+	-	+	-	-		+		*	+	-	-
		3	(+)	+	+	+	+	-	+	+	+	+	+	•		+		•
		4	(+)	+	+	+	+		+	•		(+)		+	7	+	-	-
		5	-	•	+	+	+	-	+	•	+	+	-	+	-	+	-	-
		6	-	-or+	+	+	+	-	+		+	+	+	-		+	-	-
		7	2	-or+	+	+	+	-	+		+	+	+	+	-	+	-	-
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		3	-	7	+	+	+	73	+	+	+	+	+	-	-	+	-	-
		4	-	·	+	+	+	-	+	+	+	(-)	+	+	-	+	-	-
		5	2		+	+	+	-	+	+	+		-	+	2	+	-	2
	B. neotomae		-	+	+	-	+	-	+	-	-	-	+	-	-	(+)	-	2
	B. ovis		+	-	-	-	-	-	-	+	+	(-)	-	-	+	1	-	+
	B. canis		-		+	+	+	-	+	+	+	(-)	-	-	+	-	-	+
	B. ceti		-		+	+	+	-	+	+	+	+	+	+or-	-	-	-	-
	B. pinnipedialis		-	-	+	+	+	-	+	+	+	+	+	+or-	5	-		-
	B. microti		-	-	+	+	+	-	+	+	+	+	-2	+	51	+		-
	B. inopinata		-	+	+	+	+	-	+	+	+	+	-	-	+	+	-	

+ positive; - negative; (+) usually positive; (-) usually negative; ** Species differentiation is obtained on Trypticase-soy agar or tryptose agar with the concentrations of dyes: (a)1:25 000, (b)1:50 000; ***B. abortus. 1, 2 and 4 do not grow in 1:100 000 thionine; † Monospecific antiserum: A-abortus; M-melitensis; R-rough; d-doubtful; # Tbilisi; RTD-routine test dilution
* Adapted from Alton et al. 1975; Foster et al., 2007; Jacques et al. 2007; Poester et al., 2010.

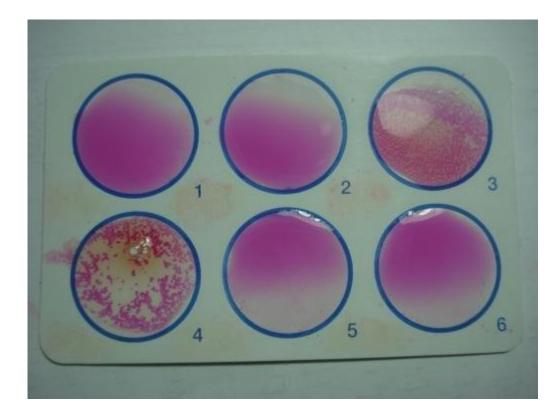
2-Serological tests

- Serological tests are crucial for laboratorial diagnosis of brucellosis
- Inactivated whole bacteria or purified fractions (i.e. lipopolysaccharide or membrane proteins) are used as antigens for detecting antibodies generated by the host during the infection.
- Problem = cross reaction

Several serological methods are available. They can be classified as :

- Screening tests (e.g. buffered antigen plate agglutination BPAT),
- Monitoring or epidemiological surveillance tests (e.g. milk ring test),
- Complementary or confirmatory tests (e.g. 2-mercaptoethanol, complement fixation, ELISAs, and fluorescence polarization assay).
- Example: Rose Bengal test is a traditional methods that has been extensively used for diagnosis of brucellosis in cattle, pigs, and goats

 Rose Bengal Plate Test; Positive test showing agglutination no.3 and 4.



3-Molecular diagnosis

- Molecular techniques are important tools for diagnosis and epidemiologic studies.
- They provide information for identification of species and biotypes of *Brucella* spp.
- Molecular detection of *Brucella* sp. can be done directly on clinical samples without previous isolation of the organism.
- These techniques can be used to complement results obtained from phenotypic tests.
- Most of the molecular diagnostic methods for brucellosis have sensitivity ranging from 50% to 100% and specificity between 60% and 98%. This is because the DNA extraction protocol, type of clinical sample, and detection limits of each protocol, are factors that can influence the efficiency of the technique.
- Polymerase Chain Reaction (PCR): amplification of specific genomic sequences of the genus or virulence genes to determine pathogenicity.
- Real time PCR qPCR is much more sensitive and can be used even on low number of bacteria in the samples.

Campylobacter

- Campylobacter spp. belong to the family *Campylobacteriaceae*.
- *Campylobacter* constitute a diverse group of organisms, some of which are well-known causes of clinical illness in animals and humans, whereas many other members of the genus appear to be commensals in the intestinal tract.
- They are Gram-negative rods.
- *Campylobacter* cells are S/seagull-shaped, spirally curved rods 0.2–0.8 µm wide and 0.5–6.0 µm long, but cells may transform to spherical or coccoid forms in response to stress conditions.
- non-spore-forming
- Most are motile, exhibiting a characteristic corkscrew-like or darting motility motion. Motility is conferred by flagella, usually in the form of a single flagellum at one or both ends of the cell, although non motile species or species with multiple flagella have been described
- Gram-negative bacteria and

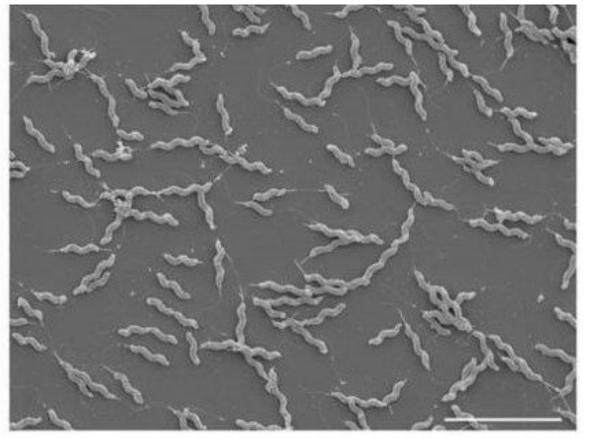


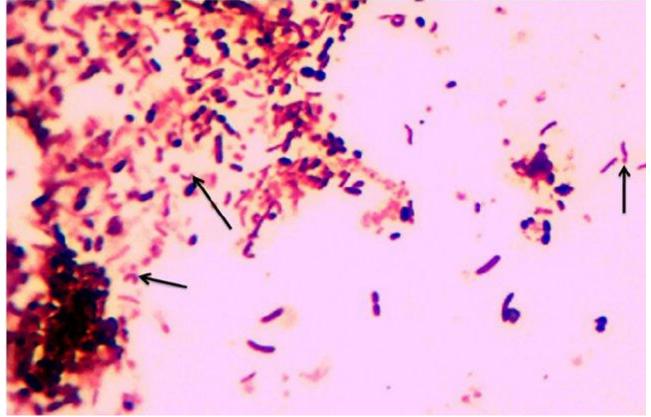
- Optimal growth occurs at **37–42**°C for thermophilic members (*Campylobacter jejuni* and *Campylobacter coli*), whereas *C. fetus* grows well at **25**°C and 37°C and usually does not grow at **42**°C.
- *Campylobacter* spp. are typically unable to ferment or oxidize carbohydrates, and thus energy is derived from the degradation of amino acids or tricarboxylic acid cycle intermediates.
- They are slow-growing, fastidious organisms and require a microaerobic atmosphere (containing **5%** O2, 10% CO2, and **85%** N2) for optimal growth.
- In general, *Campylobacter* is sensitive to oxygen, desiccation, osmotic stress, low pH, and high temperatures.

- Campylobacter species (primarily C. jejuni and C. coli) are essentially commensals in birds and frequently colonize the intestine in high numbers.
- •*Campylobacter* contamination of poultry meat is a significant concern for food safety.

Scanning electron micrograph of the spiral shape and flagella of Campylobacter jejuni

The C, S or gull wing shape of Campylobacter species (arrows) stained by 1% Carbol fuchsin.





https://www.researchgate.net/publication/271214444_Evaluation_of_detection_methods_for_Campyl obacter_infections_among_under-fives_in_Mwanza_City_Tanzania/figures?lo=1

https://www.nature.com/articles/srep38303#f



- Culture in conjunction with histopathological findings are used for definitive diagnosis of *Campylobacter* in ruminant abortions.
- The organism could be isolated from the placenta and fetal stomach contents, or from fetal lung, and occasionally from fetal liver.
- To isolate *Campylobacter*, a variety of selective culture media can be used such as chrome agar.
- Dark field examination of wet mounts of fetal stomach contents or placenta for bacteria of typical morphology and characteristic of *Campylobacter such as* darting movement of corkscrew-like organisms (especially with *C. fetus* subsp. *fetus*) represents a rapid test to establish a presumptive diagnosis
- Direct staining with dilute carbol fuchsin or fluorescent antibody staining of smears of fetal abomasal contents can also be used to demonstrate *Campylobacter*.

- Following the culture, colonies resembling *Campylobacter* need to be confirmed for definitive diagnosis, which could be done using a variety of assays, including phenotypic tests, polymerase chain reaction (PCR),
- In sheep, a standard PCR is used in detection of both *C. fetus* subsp. *fetus* and *C. jejuni* DNA directly from aborted fetal tissues and placentas

• Colonies of Campylobacter on Blood Agar plate



CHROM agarTM Campylobacter: Typical Appearance of microorganisms Campylobacter jejuni, coli, lari \rightarrow red

