



The Role of Nanotechnology in the Control of Bacterial Contamination in Plant Tissue Culture: A Review

Esraa A. H. Al Samir¹, Intisar Albandar², Muntaha J. Kadhim³ and Shaimaa A. Alsamir^{4*}

^{1, 2, 4}Department of Biology, College of Science, University of Basrah, Basrah, Iraq

³Date Palm Research Center- University of Basrah, Basrah, Iraq

*Corresponding Author: Shaimaa A. Alsamir

ORCID ID: 0000-0002-6528-3676

Abstract

Successful “micro-propagation” requires avoiding microbial contamination of plant tissue cultures at every stage of growth. Bacterial contaminants (both Endophytic and epiphytic organisms) can lead to the loss of propagated plants at every stage of development. Plants infected with bacterial contamination will suffer from impaired rooting and multiplication, and may even die. Plant death results in the loss of plant tissue, which is particularly detrimental for the tissue culture of economically important plants such as potatoes and date palms. Early detection of bacterial contamination is difficult, as it is present within plant tissues, leading to the loss of plant tissue before the contamination is detected and treated. Bacterial contamination can result from various causes, including the use of improper sterilization techniques or the use of improperly sterilized equipment. To produce sterile plant cultures, various criteria must be known, such as the source of contamination and the type of pathogen. These criteria can be minimized, and sterilized plant cultures can be produced using antibiotics or other advanced techniques. It has been found that methods of preventing bacterial contamination using chemical compounds cause harm to both the transferred and growing plant tissues and to workers in plant tissue culture laboratories or fields. Therefore, nanotechnology using various types of nanoparticles has emerged as a promising new solution to this intractable and widespread problem in plant tissue cultures. Nanoparticles may serve as an alternative to antibiotics, particularly when bacteria develop resistance to existing antibiotics.

Keywords: *Plant tissue culture, Contamination, Bacteria, Nanoparticles, Nanotechnology*

Introduction

Sources of Bacterial contamination

It is difficult to recognize the sources of contaminated cultures [1]. However, it was evident that bacteria causing contamination in plant tissue cultures may originate from laboratory environments, explants, ineffective sterilization techniques, or operators [2]. Moreover, it was detected that the species of plant and the type of explant

can determine bacterial contamination [3]. One of the most important features of bacterial contamination is "decreased shoot proliferation, unstable growth or tissue necrosis" [4,5]. Figure 1 shows an image of a plant contaminated with bacteria that grew as creamy colonies on a nutrient agar plate. Figure 2 illustrates the colonies of three types of bacteria found in contaminated palm tree tissue.



Figure 1. The potato culture is contaminated with endogenous bacterial growth at the base of the explants and the red colonies of contaminated bacteria [6]



Figure 2. Figure 2. Bacterial colonies isolated from contaminated plant tissue culture, showing different bacterial species [7]

In plants, there are two kinds of bacteria, which are “Epiphytes or Endophytes” [8]. Various types of disinfectants can effectively remove epiphytic bacteria, as they are commonly found on the surface of plants [9, 10]. [11] found that “Endophytic bacteria” didn’t cause any symptoms of disease during a part of the life cycle or the entire life cycle. In contrast, bacteria living inside healthy plants can be hazardous, leading to decreased growth and rooting rates, and can also show resistance to surface disinfectants [3].

Prevention of bacterial contamination

To prevent contamination, it must take into account the importance of each step in plant tissue culture (choosing explant, preparation of media, sub-culturing, plant incubation) [12]. *In vitro* plant cultures, most bacterial contaminations are present in the “culture vessels, laboratory media, and inefficient sterilization of explants, which happens during transmission by thrips and mites or during handling the plant material [13].

The plant contamination can be reduced by maintaining laboratory cleanliness during the subculturing of plant cultures and ensuring the entire plant tissue culture process is conducted aseptically. [12] pointed out that “bacterial contamination” can be

decreased by adjusting the air source and laboratory room cleanliness during subculturing of plant cultures. Furthermore, the populations of epiphytic organisms (stock plants) applied for plant tissue cultures should be grown under non-endangered environments (greenhouses and growth chambers) [12, 14]. [2] emphasized using the watering stock plants with filtered water to reduce bacterial contamination. Moreover, [15] suggested that bacterial characterization is necessary to identify the sources of contamination, determine contamination level from each source, and develop strategies to eradicate or stop contaminants.

Antibiotics as a traditional solution for plant tissue bacterial contamination

In plant tissue culture, bacterial contamination, which is caused by “Endophytic bacteria,” can not be removed by any superficial sterilization methods; therefore, it is a substantial issue that demands antibiotic therapy [6]. The antibiotics used in micropropagation should be non-resistance-inducing, stable, unaffected by pH, soluble, inexpensive, dissolved in the media, bactericidal, highly active, suitable for combination use, and nontoxic to human health [16].

The studies revealed that antibiotics are used to eliminate bacteria when a contaminant is indicated, or they can be used in combination as prophylactics in tissue culture medium [16]. Additionally, the studies revealed that bacterial resistance develops when a single antibiotic is used repeatedly [2, 7]. Antibiotics are classified into four main categories: inhibitors of bacterial cell wall synthesis, those acting by mode of action, DNA replication blockers, and inhibitors of bacterial protein synthesis [16]. Antibiotics can also be grouped by chemical structure into: aminoglycosides, β -lactams, glycopeptides, quinolones, macrolides, lacosamides, polymyxins [17].

Knowing the kinds of bacteria present—whether they are Gram-positive or Gram-negative—is necessary in order to choose an appropriate antibiotic. Therefore, some simple biochemical tests and initial characterization with Gram staining are necessary [18], and no single antibiotic is effective against all bacteria. [17] showed that many kinds of antibiotics are used to treat plant tissue cultures, such as carbenicillin, gentamicin, cephalothin, polymyxin,

streptomycin, rifampicin, and timentin have been used, and sometimes combinations of antibiotics may be used, which have positive effects against bacterial contamination.

Figure 3 shows different types of bacterial strains that are isolated from the contaminated plant tissues and tested for sensitivity using five types of antibiotics. The figure shows that different bacterial isolates showed different patterns of sensitivity against these antibiotics, which reflect their ability to affect the plant tissue.

On the proper media, different bacterial strains were isolated from contaminated plant tissues. After purifying the isolates, one colony was cultivated on the nutrient broth medium. Following an overnight incubation period at 37°C, the culture was spread out on Muller-Hinton agar plates, and using sterile forceps, the five different kinds of antibiotic discs were placed on the agar plate's surface. To determine how sensitive the bacterial isolates were to the antibiotics, the plates were then incubated at 37 °C. The diameter of the inhibition zones surrounding the antibiotic discs was then determined.



Figure 3. Antibiotic sensitivity test for bacteria isolated from contaminated plant tissue cultures [17].

Identification and characterization

To avoid bacterial plant tissue contamination, it's necessary to identify bacterial colonies, which provide important information about the sources of contamination [17]. There are numerous methods for isolating and cultivating bacteria. We can purify non-tamable bacteria using standard bacteriological procedures and characterize them through biochemical tests, such as motility, gelatinase, Gram stain, and oxidase [19]. A reliable systematic bacteriology source, which is crucial for characterizing genera and species to identify bacteria, is used in this procedure [20]. These traditional detection techniques need time, and in some cases, bacterial growth can delay micropropagation while

Molecular recognition based on direct deoxyribonucleic acid investigation is subtle enough for most bacterial sets, rendering these time-consuming methods redundant. At the genus level, the most common way for

bacterial identification is the 16S ribosomal RNA (rRNA) [21]. The detection of bacteria in plant tissue cultures has been done using this technique for the *Prunus avium* [22], *Cynara scolymus* L (Artichoke) [23], *Vitis vinifera* [24], *Echinacea purpurea* (Echinacea) [25], and in many propagated ornamental plants, *Dendranthema*, *Rosa*, *Spathiphyllum*, *Pelargonium*, *Ficus*, and *Dahlia* [21]. [26, 27] recommend using multiple tests for accurate identification of bacteria in micropropagation.

Nanotechnology as an advanced strategy for plant tissue bacterial contamination

Nanotechnology is essential for improving several facets of plant tissue culture [28]. Recently, nanoparticle applications have auspiciously directed the removal of bacterial contaminants from plant tissue cultures and revealed the useful role in callus proliferation, carnal embryogenesis,

organic tissue development, genetic alteration, and production of secondary metabolites [29-31]. Additionally, nanoparticles function as enhancers to

promote the large-scale production of bioactive chemicals, rapid shoot development, and root protraction [32] (Figure 4).

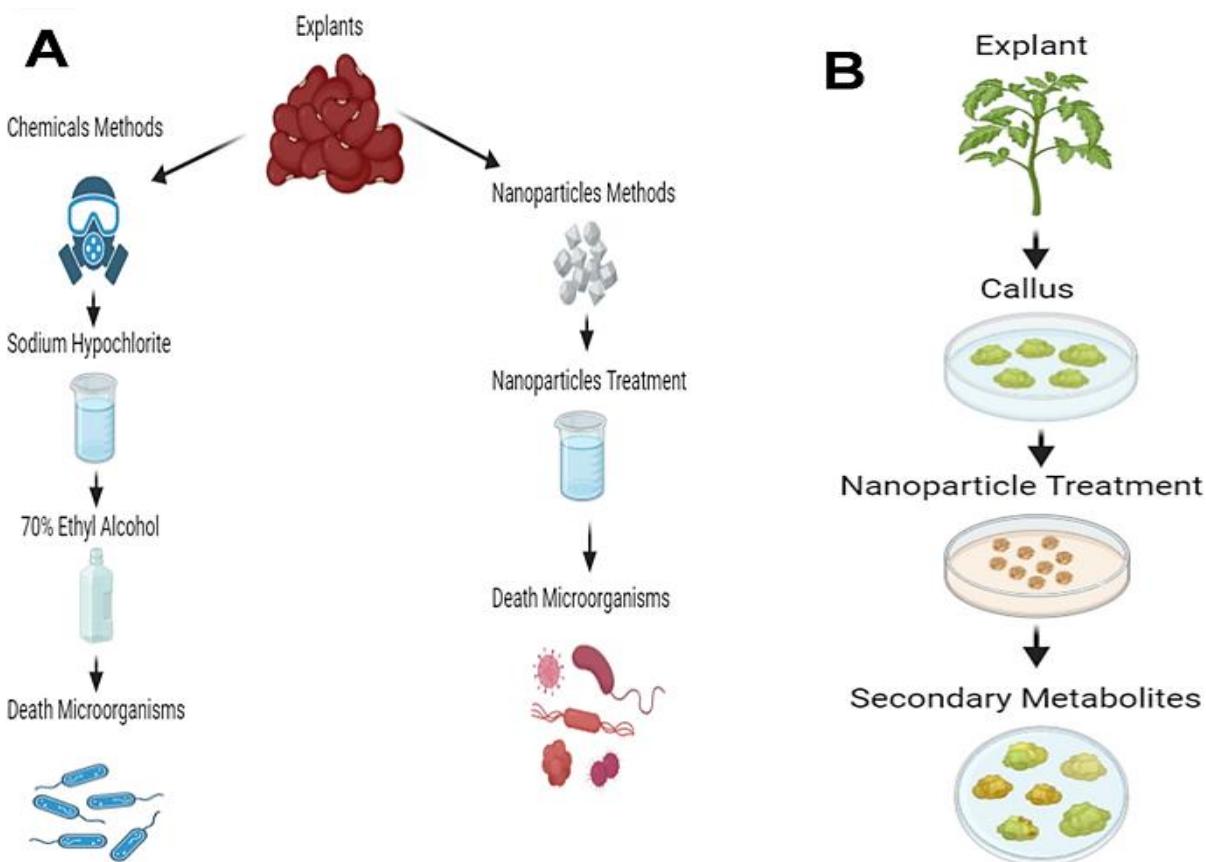


Figure 4. Treatment of explant with different types of nanoparticles. A comparison of two methods of treating bacteria isolated from contaminated plant tissue: chemical treatment and nanoparticle therapy. Figure A illustrates that chemical treatment requires wearing a face mask and employing hazardous chemicals. While nanoparticles, particularly green-manufactured ones, are safe to use and do not require chemicals, B: Applying nanoparticles to the medium with the callus. The use of nanoparticles improved callus development and efficiently generated secondary metabolites [32]

Furthermore, nanoparticles are involved in reducing the oxidative stress in plant cells, thus encouraging the development of more

calluses and higher levels of secondary metabolites [33]. The nanoparticle's potential to promote more effective, long-lasting, and

improved technological settlement in in vitro culture is highlighted by its combined presence in plant tissue culture [34]. The connotations of nanoparticles include reducing dependence on natural resources and minimizing environmental impacts, with nanotechnology considered a transformative strategy for sustainable plant tissue biotechnology [35].

Nanoparticles with an average size of 1–100 nm, have had a significant impact on the field of plant tissue culture due to their distinct physical and chemical characteristics. Different types of nanoparticles, such as silver (AgNPs), Gold (AuNPs), Magnesium (MgNPs), Copper (CuNPs), Zinc/ Zinc oxide (ZnNPs/ ZnONPs), and Titanium dioxide (TiO₂NPs), have appeared as innovative critical tools in the development of traditional plant tissue culture [36-38]. These nanoparticles are available in various forms, such as small

particles, powders, clusters, and tubes, and come in different sizes. The most recognized property of nanoparticles (NPs) is their ability to kill various types of bacteria and microorganisms [39]. They act as antibacterial agents by contributing numerous functionalities, depending on their distinctive physicochemical characteristics [40], and also inhibit browning [41]. Figure 5 illustrates the applications of nanoparticles in plant tissue culture [42]. The figure illustrates how nanoparticles can be used in plant tissue for a variety of purposes, including preventing contamination by either adding nanoparticles to the prepared media of plant tissue before explant transfer or to the cultured explant after it has grown. Additionally, by preserving nutrients, improving nutrient absorption, maintaining moisture in media, and enhancing secondary metabolites, nanoparticles can promote the growth of plant tissue.

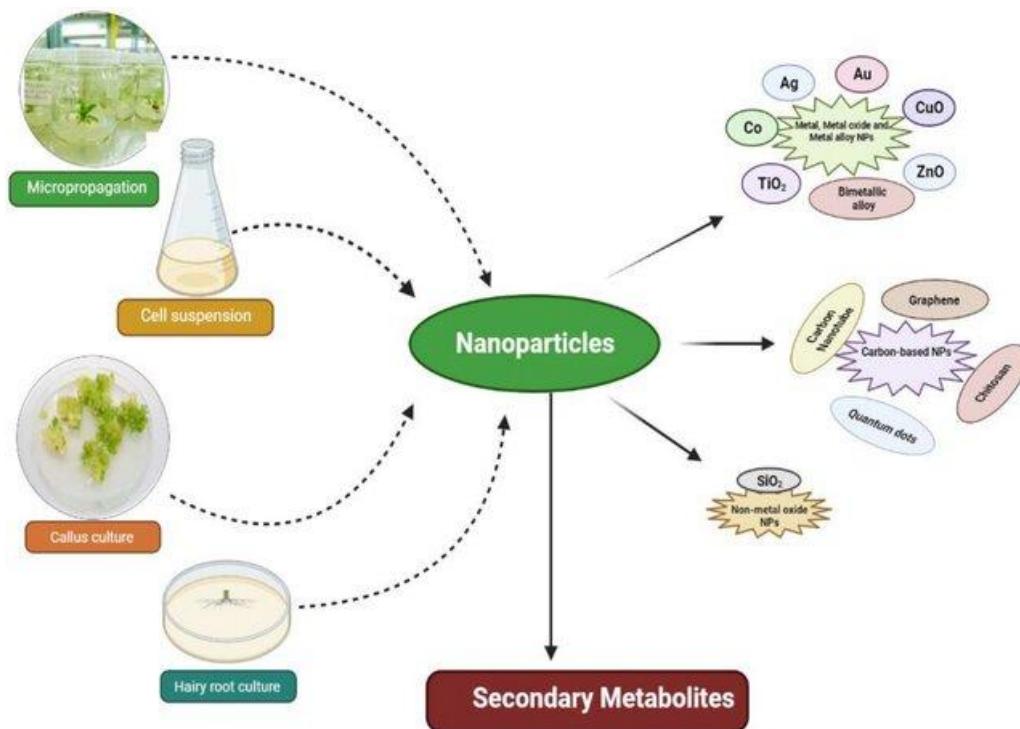


Fig. 5Different applications of Nanoparticles' in plant tissue culture [42]

Silver nanoparticles are commonly used in various fields, including medical and environmental applications [43], and primarily in plant tissue agriculture [44]. AgNPs can be used to disinfect the surface of explants and have been shown to reduce microbial contamination in various plants. Silver NPs' antibacterial properties appear to have been thoroughly investigated [45]. Although AgNPs have a strong capacity to eradicate microorganisms without impairing plants, numerous studies have examined their impact on plant disinfection, revealing both beneficial and detrimental effects on plants [46]. Since plants are a fundamental component of the world and a source of diet,

it was suggested that more research must be done on the effects of NPs on explants removed from various plant species to establish an appropriate technique for applying AgNPs in plant disinfection [47]. Therefore, it is crucial to understand how AgNPs affect plants, particularly since their widespread use raises the possibility that they will be discharged into the ecosystem [48-50]. Numerous studies have documented the use of gold nanorods (GNRs) and gold nanostars (GNSs) in tests to eradicate bacteria [51]. Furthermore, ZnO and Cu nanoparticles showed high effectiveness against microorganisms [52, 53]. It was discovered that the nanoparticles kill bacteria

by generating reactive oxygen species (ROS) [54, 55].

The introduced review focused on demonstrating the effect of microorganisms on plant tissue culture and the most commonly used materials to mitigate this effect. Moreover, the review indicated the role of nanotechnology, using different types of nanoparticles, to reduce or eliminate microbes due to their unique properties resulting from a high surface-to-volume ratio. Furthermore, nanoparticles could be used to enhance the survival rate of explants and remove contamination.

Conclusions

Bacterial contamination, in micropropagation, “plant tissue culture”, is still considered a continuing threat; however, there are many methods for reducing this contamination. First, Laboratories must ensure that all processes are performed in a sterile environment to determine or avoid bacterial contamination. Second, during the culture cycle and at the initiation stage, indexing culture is important because it reduces the number of contaminants that escape detection; furthermore, it uses antibiotics to determine bacterial contamination. However, nanotechnology has advanced to improve this sector of

biotechnology by enhancing plant growth, increasing its resistance to insects, and eliminating bacterial contamination that continues to threaten plant tissue culture.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

References

1. Kane, M." Bacterial and fungal indexing of tissue cultures." *Journal of Allergy and Immunology*.2003; 94, pp.393-400.
2. Leifert C, Waits, WM . "Bacterial growth in plant tissue culture media". *J Appl Microbiol*.1992; 72: 460-466.
3. Cassells, A. C. "Pathogen and biological contamination management in plant tissue culture: phytopathogens, vitro pathogens, and vitro pests. In: Plant cell culture protocols". Totowa, NJ: Humana Press.2012; 57–80.
4. Leifert, C. and Cassells, A. C." Microbial hazards in plant tissue and cell cultures". *In Vitro Cellular & Developmental Biology-Plant*.2001; 37(2): 133–138.
5. Klayraung, S., Niamsup, P., Poonnoy, P. and Topoonyanont, N." Diversity and control of bacterial contamination of plants propagated in temporary immersion bioreactor system". *Acta Horiticulturae*.2017; 1155: 439–446
6. Szewczyk-Taranek, B., Jaglarz, A., Pałka, P., Supel, P., Kaszycki, P., Mazur, J. and Pawłowska, B. "Identification and control of endophytic bacteria during in vitro cultures of *Staphylea pinnata L*". *Folia Horiticulturae*.2020; 32(1), pp.47-55.
7. Abd Burgal, A.H., Aboobmukhaifi, E., Alkanany, K.M. and AL-Tamimi, W.H."

Biological control of microbial contamination associated with plant tissue culture of date palm (*Phoenix dactylifera*)". *Pollution Res.*2021; 40(4), 1481-1488.

8. Gunson. H. E. and Spencer-Phillips. P. T.N." Latent bacterial infections: Epiphytes and endophytes as contaminants of micro propagated plants". *Physiology, Growth and Development of Plants in Culture.* J. R. Nicholas, ed. KJ uwer Academic Publishers, Dordrechr, Netherlands.1994; 379-396.

9. Tomaszewska-Sowa, M. and Figas, A." Optimization of the processes of sterilization and micropropagation of cup plant (*Silphium perfoliatum* L.) from apical explants of seedlings in in vitro cultures". *Acta Agrobotanica.*2011; 64(4): 3–10.

10. Bhadrawale, D., Mishra, J. P. and Mishra, Y." An improvised in vitro vegetative propagation technique for *Bambusa tulda*: influence of season, sterilization and hormones". *Journal of Forestry Research.*2018; 29(4): 1069–1074

11. Barzanti, R., Ozino, F., Bazzicalupo, M., Gabbielli, R., Galardi, F., Gonnelli, C. and MENGONI, A. "Isolation and characterization of endophytic bacteria from the nickel hyperaccumulator plant *Alyssum bertolonii*". *Microbial Ecology.*2007; 53(2): 306–316.

12. Leifen, C., Morris, C.E. and Waites. W.M. "Ecology of microbial saprophytes and pathogens in rissuc culrure and field-grown plants: Reasons for contamination problems in vajra". *Critical Reviews in Plant Science.*1994; 13: 139-183.

13. Moutia, M. and Dookun, A. "Evaluation of surface sterilization and hot water treatments on bacterial contaminants in bud culture of sugarcane". *Experimental Agriculture.*1999;35(3): 265–274.

14. Orlikowska, T., Nowak, K. and Reed, B." Bacteria in the plant tissue culture environment. *Plant Cell, Tissue and Organ Culture*".2017; 128(3): 487–508

15. Leifert, C., Ritchie, J. Y. and Waites, W. M. "Contaminants of plant-tissue and cell cultures". *World Journal of Microbiology and Biotechnology.*1991a; 7: 452–469.

16. Abass MH , Al-Utbi S.D and Al-Samir E.A.H. " Morphological and biochemical impact of different decontamination agents on date palm (*Phoenix dactylifera* L) pro callus".*Australasian journal of crap science.*2016;10(7):1022-1029.

17. Abass MH . "Microbial contaminants of date palm (*Phoenix dactylifera* L.) in Iraqi tissue culture laboratories. *Emir J of food and agricult.*2013a;25:875-882.

18. Al-Dosary NH, Al-Mussawi MA, Al-Taha HA ."Isolation and identification of bacterial types that cause contamination of date palm (*Phoenix dactylifera* L.) callus and studying the inhibition activities of some plant extracts and antibiotics". *Basra J Date Palm Res.*2011;10:68-81.

19. Leifert, C., Camotta, H., Wright, S. M., Waites, B., Cheyne, V. A. and WAITES, W. M. "Elimination of *Lactobacillus plantarum*, *Corynebacterium* spp., *Staphylococcus saprophyticus* and *Pseudomonas paucimobilis* from micropropagated *Hemerocallis*, *Choisya* and *Delphinium* cultures using antibiotics". *Journal of Applied Bacteriology.*1991b 71(4): 307–330.

20. Krieg, N.R. and Holt, J.G. (eds.). "Bergey's Manual of Systematic Bacteriology". Vol. I., Williams and Wilkens, Baltimore.1984.
21. Moreno-Vázquez, S., Larrañaga, N., Überhuaga, E. C., Braga, E. J. B. And Pérez-Ruiz, C. "Bacterial contamination of in vitro plant cultures: confounding effects on somaclonal variation and detection of contamination in plant tissues". *Plant Cell, Tissue and Organ Culture*.2014;119(3): 533–541.
22. Quambusch, M., Pirttilä, A. M., Tejesvi, M. V., Winkelmann, T. and BARTSCH, M. "Endophytic bacteria in plant tissue culture: differences between easy- and difficult-to-propagate *Prunus avium* genotypes". *Tree Physiology*.2014;34(5): 524–533
23. Navacchi, O., Zuccherelli, G. And Mazzucchi, U. "Development of ambivalent media to detect bacterial contamination of globe artichoke micropropagation media". *Acta Horticulturae*.2013; 983: 215–220.
24. Thomas, P. "Isolation of *Bacillus pumilus* from in vitro grapes as a long-term alcohol-surviving and rhizogenesis inducing covert endophyte". *Journal of Applied Microbiology*. 2004;97(1): 114–123.
25. Lata, H., Li, X. C., Silva, B., Moraes, R. M. And Halda-Alija, L. 2006." Identification of IAA-producing endophytic bacteria from micro propagated *Echinacea* plants using 16S rRNA sequencing". *Plant Cell, Tissue and Organ Culture*.2006; 85(3): 353–359.
26. Jones, J.B.. Chase, A.R. and Harris, G.K- 1993. Evaluation of the Biolog GN MicroPlare sysrem for idenrification of some plant-pathogenic bacteria. *Plant Dis*. 77: 553-558.
27. Vcmiere, c., Provost. O.. Civerolo, E.L., Garnbin. O., Jacquemoud-Collet. J.P. and Luisctti, J. "Evaluation of the biology substrate utilization system to identify and assess metabolic variation among strains of *Xanthomonas campestris* pv. *Citri*". *Applied and Environmental Microbiology*.1993; 59: 243-249.
28. Jan, S., Jan, N., Singh, S., Shah, M.A. and Bhat, I.A." Nanotechnology in Plant Tissue Culture: A Review". *Horticultural Plant Journal*.2025.
29. Gunasena, M.D., Alahakoon, A.M., Polwatthha, K.P., Galpaya, G.D., Priyanjani, H.A., Koswattage, K.R. and Senarath, W.T. "Transforming plant tissue culture with nanoparticles: a review of current applications". *Plant Nano Biology*. 2024;10, p.100102.
30. Alfarraj, N.S., Tarroum, M., Al-Qurainy, F., Nadeem, M., Khan, S., Salih, A.M., Shaikhhaldein, H.O., Al-Hashimi, A., Alansi, S. and Perveen, K. "Biosynthesis of silver nanoparticles and exploring their potential of reducing the contamination of the in vitro culture media and inducing the callus growth of *Rumex* nervous explants". *Molecules*.2023;28(9), p.3666.
31. Balamurugan, V., Abdi, G., Karthiksaran, C., Thillaigovindhan, N. and Arulbalachandran, D. "A review: improvement of plant tissue culture applications by using nanoparticles". *Journal of Nanoparticle Research*.2024; 26(8), p.188.
32. Khan, R., Sohail, A., Aslam, M.U., Khan, L., Ahmad, S. and Taimur, M. "Utilizing

nanoparticles as innovative elicitors to enhance bioactive compounds in plants". *Int. J. Res. Adv. Agric. Sci.*2023; 2, pp.24-34.

33. Sharma, S., Singh, V.K., Kumar, A. and Mallubhotla, S. "Effect of nanoparticles on oxidative damage and antioxidant defense system in plants". *Molecular plant abiotic stress: Biology and biotechnology*.2019; 315-333.

34. Mehbub, H., Akter, A., Akter, M.A., Mandal, M.S.H., Hoque, M.A., Tuleja, M. and Mehraj, H." Tissue culture in ornamentals: Cultivation factors, propagation techniques, and its application". *Plants*.2022; 11(23), p.3208.

35. Atanda, S.A., Shaibu, R.O. and Agunbiade, F.O. "Nanoparticles in agriculture: balancing food security and environmental sustainability". *Discover Agriculture*.2025; 3(1), p.26.

36. Voloshyna, I.M., Netiha, Y.M., Nechaiuk, Y.V., Khomenko, V.G. and Shkotova, L.V. "The influence of metal nanoparticles on plants". *Biopolymers & Cell*.2024; 40(2), p.83.

37. Sarmast, M.K. and Salehi, H. "Silver nanoparticles: an influential element in plant nanobiotechnology". *Molecular biotechnology*.2016; 58(7):441-449.

38. Thapliyal, D., Verros, G.D. and Arya, R.K. "Nanoparticle-doped antibacterial and antifungal coatings". *Polymers*.2025; 17(2), p.247.

39. Altammar, K.A. "A review on nanoparticles: characteristics, synthesis, applications, and challenges". *Frontiers in microbiology*.2023; 14, p.1155622.

40. Ozdal, M. and Gurkok, S. "Recent advances in nanoparticles as antibacterial agent". *ADMET and DMPK*.2022;10(2) :115-129.

41. Humbal, A. and Pathak, B. "Harnessing nanoparticle-mediated elicitation in plant tissue culture: a promising approach for secondary metabolite production". *Plant Cell, Tissue and Organ Culture (PCTOC)*.2023; 155(2):385-402.

42. Vadakan, K., Rumjit, N.P., Ngangbam, A.K., Vijayanand, S. and Nedumpillil, N.K. "Novel advancements in the sustainable green synthesis approach of silver nanoparticles (AgNPs) for antibacterial therapeutic applications". *Coordination Chemistry Reviews*.2024; 499: p.215528.

43 Albandar, I., Radhi, O.A., Al-Saadi, S.A. and Mahdi, M.A." Eco-Friendly Synthesis of Silver Nanoparticles Using *Prosopis farcta* Fruit Extract and Evaluate Their Biological Applications". *Pakistan Journal of Life & Social Sciences*.2024;22(1).

44. Alfosea-Simón, F.J., Burgos, L. and Alburquerque, N. "Silver nanoparticles help plants grow, alleviate stresses, and fight against pathogens". *Plants*.2025; 14(3): p.428.

45. Tung, H.T., Bao, H.G., Cuong, D.M., Ngan, H.T.M., Hien, V.T., Luan, V.Q., Vinh, B.V.T., Phuong, H.T.N., Nam, N.B., Trieu, L.N. and Truong, N.K." Silver nanoparticles as the sterilant in large-scale micropropagation of chrysanthemum". *In Vitro Cellular & Developmental Biology-Plant*.2021; 57(6): 897-906.

46. Courtois, P., Rorat, A., Lemiere, S., Guyoneaud, R., Attard, E., Levard, C. and Vandenbulcke, F. "Ecotoxicology of silver nanoparticles and their derivatives introduced in soil with or without

sewage sludge". A review of effects on microorganisms, plants and animals. Environmental pollution.2019; 253:578-598.

47. Khan, S., Zahoor, M., Khan, R.S., Ikram, M. and Islam, N.U." The impact of silver nanoparticles on the growth of plants". The agriculture applications. Heliyon.2023; 9(6).

48. Dang, F., Huang, Y., Wang, Y., Zhou, D. and Xing, B. "Transfer and toxicity of silver nanoparticles in the food chain". Environmental Science: Nano.2021 8(6): 1519-1535.

49. Kulkov, L., Arkhipov, R., Abramova, A., Vereshchagin, M., Voronkov, A., Khalilova, L., Kartashov, A., Tarakanov, I., Kreslavski, V., Kuznetsov, V. and Pashkovskiy, P." Long-term effects of silver nanoparticles and mineral nutrition components on the photosynthetic processes, chloroplast ultrastructure and productivity of *Solanum lycopersicum* plants". Journal of Photochemistry and Photobiology B: Biology. 2024;260:p.113038.

50. Tanisha, Jadhav, U. and Alim, H. "Genotoxicity of Silver Nanoparticles in Plants and Underlying Mechanism. In Plant Response to Silver Nanoparticles: Plant Growth, Development, Production, and Protection"2024: (pp. 59-78). Singapore: Springer Nature Singapore.

51. Rabiee, N., Ahmadi, S., Akhavan, O. and Luque, R. "Silver and gold nanoparticles for antimicrobial purposes against multi-drug resistance bacteria". Materials .2022;15(5): p.1799.

52. Yagoub, A.E.A., Al-Shammary, G.M., Al-Harbi, L.N., Subash-Babu, P., Elsayim, R., Mohammed, M.A., Yahya, M.A. and Fattiny, S.Z. "Antimicrobial properties of zinc oxide nanoparticles synthesized from *lavandula pubescens* shoot methanol extract". Applied Sciences.2022;12(22), p.11613

53. Albandar, I., Jabbar, S., Ibrahim, T.K., Radhi, O.A. and Mbalaha, Z.S." Advances of Nanotechnology in Eradication Bacterial Infectious Diseases".A Recent Review. Kirkuk Journal of Science.2024; 19(2).

54. Venditti, I. "Engineered gold-based nanomaterials: Morphologies and functionalities in biomedical applications". a mini review. Bioengineering.2019; 6(2), p.53.

55. Okkeh, M., Bloise, N., Restivo, E., De Vita, L., Pallavicini, P. and Visai, L. "Gold Nanoparticles: Can They Be the Next Magic Bullet for Multidrug-Resistant Bacteria? "Nanomaterials.2021; 11, 312.

دور تقنية النانو في السيطرة على التلوث البكتيري في زراعة الأنسجة النباتية: مراجعة

اسراء عبد الرزاق حميد السامر¹، انتصار البندر² ، منتهي جواد كاظم³ ، شيماء عبد الرزاق السامر⁴

^{1,4,2,3}قسم علوم الحياة، كلية العلوم، جامعة البصرة، البصرة، العراق

³مركز ابحاث النخيل ، جامعة البصرة، البصرة، العراق

الباحث المراسل: شيماء عبد الرزاق السامر

الباحث المراسل ORCID ID : 0000-0002-6528-3676

الخلاصة

تتطلب الزراعة النسيجية الناجحة منع التلوث الميكروبي في مزارع أنسجة النبات في كل مرحلة من مراحل النمو. يمكن أن تؤدي الملوثات البكتيرية (الداخلية والخارجية) إلى فقدان النباتات المتکاثرة، حيث ان النباتات المصابة بالتلود البكتيري ستعاني من ضعف بعملية التجذير وعملية التضاعف ويمكن ان تموت. ان موت النباتات يتسبب في خسارة النسيج النباتي والذي يعود بالضرر الكبير خاصة في حالة زراعة الانسجة النباتية لنباتات اقتصادية مثل البطاطا وتمر التخيل. وقد لوحظ انه من الصعب التعرف على "التلوث البكتيري" مبكراً لوجوده داخل أنسجة النبات وبالتالي فقدان الانسجة النباتية قبل اكتشاف التلوث ومعالجته. ان التلوث البكتيري قد ينبع عن اسباب كثيرة منها تقنيات التعقيم الريدية او استخدام المعدات غير المعقمة بشكل جيد. لإنتاج مزرعة نباتية معقمة، يجب معرفة معايير مختلفة، مثل مصدر التلوث ونوع مسببات الأمراض، لتقليل تلوث وإنتاج مزرعة نباتية معقمة باستخدام المضادات الحيوية او غيرها من التقنيات المتقدمة. وقد وجد ان طرق منع التلوث البكتيري باستخدام المركبات الكيميائية يتسبب بأضرار على كل من الانسجة النباتية المنقوله والنامية والعاملين في مختبرات او حقول زراعة الانسجة النباتية. لذلك ظهرت تقنية النانو باستخدام انواع مختلفة من الحسيمات النانوية كحل واعد جديد لهذه المشكلة المستعصية والمنتشرة في مزارع الانسجة النباتية. وقد تستخدم الحسيمات النانوية كبديل للمضادات الحيوية خاصة عندما تتطور البكتيريا المقاومة للمضادات الحيوية المتوفرة.