



Cloning of oil degradation genes

A novel bacterium was engineered with an effective expression system to improve its ability to degrade oil in contaminated environments. Enrichment culture techniques were used with crude oil as the sole carbon source to isolate microorganisms. Seventeen bacterial isolates were identified using biochemical tests and 16S rRNA gene sequencing. Identified genera included *Acinetobacter*, *Pseudomonas*, *Aeromonas*, *Micrococcus*, *Achromobacter*, *Enterobacter*, *Klebsiella*, *Staphylococcus*, *Bacillus*, *Exiguobacterium*, *Brevibacillus*, and *Stenotrophomonas*. A specific primer was designed to amplify the *alkB* gene, confirming its presence in *Pseudomonas aeruginosa*. Plasmid curing confirmed the gene's presence on the chromosome. The *alkB* gene was cloned into a plasmid vector, transformed into *E. coli*, and confirmed via PCR. Expression of the gene was verified, and its contribution to biodegradation efficiency was tested using n-hexadecane as a substrate. Biodegradation efficiency increased from 32.63% to 77.42% after 72 hours, indicating a 44.8% improvement. Gene amplification and cloning strategies were simulated using SnapGene software. This study is the first of its kind in Iraq.

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Cloning of *alkB* gene from *Pseudomonas aeruginosa* associated with biodegradation of hydrocarbons



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