See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/343775076

Phylogenetic relationship among some species of Solanaceae in Iraq

Article in Eurasian Journal of Biosciences · August 2020

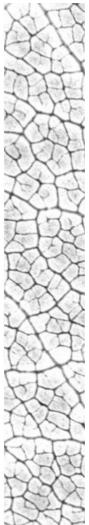
citations 0 reads 109

1 author:



Sadeq Sabeeh Kareem Al-Taie University of Misan 20 PUBLICATIONS 14 CITATIONS

SEE PROFILE





Phylogenetic relationship among some species of Solanaceae in Iraq

Sahar A. A. Malik Al-Saadi ¹, Alaa N. Al-wheeb ², Sadeq Sabeeh Kareem Al-Taie ^{3*}

¹ Department of Biology, College of Science, University of Basra, IRAQ

² Department of Biology, College of Science, University of Thi-Qar, IRAQ

³ Department of Biology, College of Science, University of Misan, IRAQ

*Corresponding author: sadeq_altaie@yahoo.com

Abstract

A phylogenetic analysis was conducted using sequences were studied. Sequences were obtained from six genera: (1) *Datura metel*; (2) *D. innoxia*; (3) *Solanum nigrum*; (4) *Hyosyamus reticula*; (5) *Capsicum annum*; (6) *Physalis alkakengy*; and (7) *Lycopersiconesculentum* and included seven species of the Solanaceae family. The taxonomy and phylogenetic relationships among these taxa were used to assess the genetic diversity of eight Solanaceae species using one nuclear internal transcribed space (ITS). Analysis of DNA sequence data from the three (ITS) regions yielded a high level of genetic variability (polymorphism) among the studied samples. Results indicate that six major clades within Solanaceae are supported by high bootstrap values. The sequences of *D.innoxia* and *D.metel* are almost 100% similar and have a close relationship with *S. nigrum*. Morphological characters such hairs, number of corolla and corolla color different between similar species. They were grouped into a different trichotomy in surface of leaves, stem, and flowers. Most species were different and occurred in separated clade.

Keywords: ITS, phylogenetic relationship, PCR, Solanaceae

Al-Saadi SAAM, Al-wheeb AN, Al-Taie SSK (2020) Phylogenetic relationship among some species of Solanaceae in Iraq. Eurasia J Biosci 14: 2391-2394.

© 2020 Al-Saadi et al.

This is an open-access article distributed under the terms of the Creative Commons Attribution License.

INTRODUCTION

The family Solanaceae is one of the largest and most important economic families of angiosperms, including important food, spice, and drug plants (D'Arcy, 1979). This family contains 83-90 genera and 2671 species, but the most recent estimate includes more than 3000– 4000 species (Gebhardt, 2016; Ganaie *et al.*, 2018). Approximately half of the species in the family are included in the widespread, morphologically diverse, and economically important genus *Solanum*. The family is nearly cosmopolitan in distribution and found throughout tropical and temperate regions, but with a concentration of diversity in Australia and Latin America. (Ganaie *et al.*, 2018; Ande et al, 2017).

Based on amplified fragment length polymorphism (AFLP) data, a new classification for the tribe Datureae was proposed, and the arborescent species was placed into a separate genus Brugmansia. Within *Datura* three sections have been recognized, namely Stramonium, Dutra and Ceratocaulis. In addition, *D. discolor*, previously placed in the section Dutra was reported to be an intermediate between sections Dutra and Stramonium (Mace *et al.*, 1999).Plastid sequences are less plagued by problems of among-site and lineage rates of heterogeneity, which have often compromised the accuracy of phylogenetic inference in animals.

Comparative studies of cpDNA are particularly relevant to Solanaceae because the chloroplast genome of Nicotiana tabacum is completely sequenced and could serve as a reference genome within the family. Many of the systematic advances based on extensive cpDNA studies have been reviewed by several researchers(Palmer 1988), phylogenetic et al., relationships of Solanum (Spooner et al., 1993), Nicotiana (Olmstead et al., 1997), Jaltomata (Mione et al., 1994), and Physalis and related genera (Mione et al., 1994; Wei et al., 2012; Marghali et al., 2015; Zhao et al., 2015; Zamora-Tavares et al., 2015). Based on the molecular phylogeny of chloroplast and mitochondrial genes, Kulcheski et al. (2006) separated the genus Petunia into two complexes while Livingstone et al. (1999) showed that Capsicum species differed by at least one reciprocal translocation. Rodriguez and Spooner (1997) and Bohs and Olmstead (1997) used a chloroplast DNA study for taxonomic analysis of Solanum. Mione et al., (1994) and Olmstead and Palmer (1997) explored systematic implications of chloroplast DNA variations in taxonomy of Jaltomata and Solanum,

> Received: June 2019 Accepted: April 2020 Printed: July 2020

 Table 1. The ITS of nuclear ribosomal DNA, including the 5.8S RNA gene were amplified by PCR using the forward primer

 AB101 (5)

	Primers		Sequences (5 - 3)	(Base)
ITS	ITS5	F	5 - GGAAGGAGAAGTCGTAACAAGG-3	22
	ITS4	R	5 - TCCTCCGCTTATTGATATGC-3	20

Temperature	Time (min)				
95	4				
No. of cycles = 32 cycle					
95	1				
56	1				
72	2				
72	10				
	95 b. of cycles = 32 cycle 95 56 72				

respectively. Melotto-Passarin (2008) studied phylogenetic relationships in Solanaceae and related species based on cpDNA sequences while using an internal transcribed sequence (ITS) as a DNA barcode for unique *Lycium barbarum* studied by Xin *et al.* (2013). The *Petunia* genome sequence is under construction and has been studied by Vandenbussche *et al.* (2016).

The aim of this study was to evaluate phylogenetic relationships with respect to genetic diversity among some species of Solanaceae using sequence data obtained from nuclear ITS.

MATERIALS AND METHODS

In the present study, we collected seven species of Solanaceae species from different places in Iraq. The samples were placed in perforated aluminum foil and sealed tightly and then placed inside polyethylene bags and closed with a little of silica gel for the purpose of absorbing moisture from the leaves.

DNA Isolation and PCR Amplification

DNA extraction was performed on young leaves using the Qiagen DNeasy (QIAGEN, Hilden, Germany)

Plant Genomic DNA Mini Kit according to the manufacturer's instructions with some modifications. Extracted DNA samples of 50 μ l were stored in the freezer at -20°C.

Polymerase chain reaction (PCR) involved amplification of the (ITS) region of nuclear ribosomal DNA. The ITS region of nuclear ribosomal DNA was composed of ITS 4 and 5 and was amplified using one primer shown in **Table 1**.

During electrophoresis, 9 μ L of each PCR product was mixed with 3 μ L of bromophenol blue, and the mixture was then spun briefly in a micro centrifuge before loading on a 1.5% agarose gel, which was previously stained with safe view. An aliquot (12.5 μ L) from the GoTaq Green Master Mix, 2X was taken and put in a PCR tube. Add Aliquots (1.5 μ L) from 10 μ M of upstream primer and 1.5 μ L of 10 μ M of downstream primer were taken. Five microliters of DNA template were taken, 4.5 μ L of nuclease-free water was added. The mixture was vortexed for 30 sec and placed into the PCR machine.

Programmer of PCR

The different stages of cycle sequencing were performed in a Gene Amp PCR System 9700 Thermocycler using the program shown in **Table 2**.

RESULTS AND DISCUSSION

Phylogenetic Analysis

The phylogeny of some Solanaceae species was estimated based on ITS multi-alignments (**Figs. 1** and **2**).

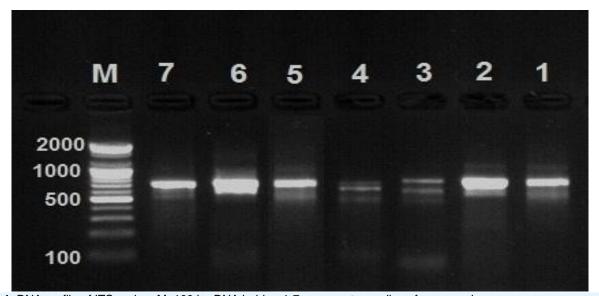
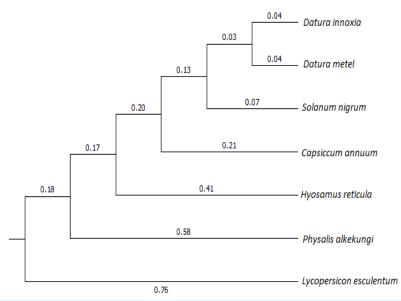
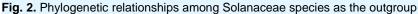


Fig. 1. DNA profile of ITS region, M: 100 bp DNA ladder, 1-7 represents amplicon from samples 1- Capsiccum annuum 2- Datura innoxia 3-Datura metel 4- Hyosamus reticula 5- Lycopersiconesculentum 6-Physalis alkekungi 7-Solanum nigrum





Analysis of DNA sequence data from the three ITS regions yielded a high level of genetic variability (polymorphism) among the studied samples. Results indicate that six major clades within Solanaceae are supported by high bootstrap values. The sequences of D.innoxia and D.metel were almost 100% similar, and they have close relationship with S.nigrum. Morphological characters such hairs, number of corolla and corolla color different between similar species. They were grouped into different trichotomies based on leaf, stem, and flower surfaces. Most species were different and occurred in separated clade.

The first analysis included seven species of Solanaceae. The analysis of this data set identified six main clades, one containing *Physalis*, two with *Capsicum*, the third clade was composed of *Solanum*, *Lycopersicon* species occupies fourth clade, and *Capsicum* formed the sixth clade. The sixth clade included two species of *Datura* (*D. metal* and *D. innoxia*), which were grouped into one clade as a sister group to *Solanum* (**Fig. 2**).

Three major clades were found in Solanaceae species: (1) the *Datura* clade with *Datura* species (0.04 % bootstrap support); (2) the *Solanuum* clade with C (0.07 %);(3), *Capsicum* 0.21 % clade; and (4), *Lycopersicon*, which was 0.75 % in agreement with

Borisjuk *et al.* (1994) who indicated that the new world *Solanum* species are more related to *Lycopersicon* than to other *Solanum* species.

One obvious example is the value of placing model organisms in the appropriate phylogenetic context in order to obtain a better understanding of both patterns and evolutionary processes, such as tomato (*Lycopersiconesculentum*) and other species dependent on an anther within the well-marked subclade (Olmstead *et al.* 1999, Spooner *et al.* 1993). This is an important observation for geneticists and molecular biologists.

CONCLUSION

Sequencing some of seven studied species of the Solanaceae family, yielded results in which chloroplast sequences are highly conserved among related species. However, several mutations, such as indels and base substitutions, were observed. This relative conservation of the genome allows for investigations of the phylogenetic relationships among divergent plants to be done. The non-coding ITS intergenic spacer thus showed nucleotide sequence polymorphism and variation in sequence length between Solanaceae and related species. It could be included in analysis with multiple chloroplast regions in order to improve the resolution of phylogenetic studies of the species studied.

REFERENCES

- Abdel Khalik K (2005) Morphological studies on trichomes of Brassicaceae in Egypt and taxonomic significance, Acta. Bot. Croat, 64 (1): 57–73.
- Ande OT, Huising J, Ojo AO, Azeez J, Are KS, et al. (2017) Status of integrated soil fertility management (ISFM) in southwestern Nigeria. International Journal of Sustainable Agricultural Research, 4(2): 28-44.
- Bohs L, Olmstead RG (1997) Phylogenetic relationships in *Solanum* (Solanaceae) based on ndhF sequences. Systematic Botany. 22: 5-17.

- Borisjuk N, Borisjuk L, Petjuch G, Hemleben V (1994) Comparison of nuclear ribosomal RNA genes among *Solanum* species and other Solanaceae. Genome 37: 271-279.
- D'Arcy WD (1979) The classification of the Solanaceae. In: JG Hawkes, RN Lester and AD Skelding (eds.). The biology and taxonomy of the Solanaceae. Academic Press: London: 3-47.
- Ganaie MM, Raja V, Reshi ZA, Verma V (2018) Family Solanaceae: Taxonomy and modern trends. Annals of plant science, 7(9): 2403-2414.
- Gebhardt C (2016) The historical role of species from the Solanaceae plant family in genetic research. Theor. Appl. Genet., 129(12): 2281-2294.
- Inamdar JA, Rao NV (1983) Light and scanning electron microscopic on trichomes of some Brassicaceae. Feddes Rep. 94: 183-190.
- Kelchner SA (2000) The evolution of noncoding chloroplast DNA and its application in plant systematics. Annual Missouri Botanical Garden 87: 482-498.
- Mace ES, Gebhardt CG, Lester RN (1999) AFLP analysis of genetic relationships in the tribe Datureae (Solanaceae). Theoretical and Applied Genetics: 634-641.
- Marghali S, Fadhlaoui I, Gharbi M, Zitouna N, Trifi-Farah N (2015) Utility of ITS2 sequence data of nuclear ribosomal DNA: molecular evolution and phylogenetic reconstruction of *Lathyrus* spp. *Sci. Hortic.* 194: 313-319.
- Melotto-Passarin DM, Berger IJ, Dressano K, Morell VF, et al. (2008) Phylogenetic relationships in Solanaceae and related species based on cpDNA sequence from plastid trnE-trnT region. Crop Breeding and Applied Biotechnology, 8: 85-95.
- Metcalfe CR, Chalk L (1950) Anatomy of Dicotyledon. Clarendon Press, Oxford, 2: pp. 724.
- Mione T, Olmstead RG, Jansen RK, Anderson GJ (1994) Systematic implications of chloroplast DNA variation in *Jaltomata* and selected physaloid genera (Solanaceae). American Journal of Botany. 81: 912-918.
- Olmstead RG, Sweere JA, Spangles RE, Bohs L, Palmer JD (1999) Phylogeny and Provisional Classification of the Solanaceae Based on Chloroplast DNA. In: Nee M., Symon D.E., Jessup J.P., Hawkes J.G. (eds.) Solanaceae IV, advances in biology and utilization. The Royal Botanical Gardens, Kew, Surrey.: 111-137.
- Orcan N, Binzet R (2003) The Anatomical and palynological properties of *Alyssum obtusifolium* steven ex DC. (Brassicaceae). Turk. J. Bot. 27: 63-68.
- Palmer JD, Jansen RK, Michaels HJ, Chase MW, Manhart JW (1988) Chloroplast DNA variation and plant phylogeny. Annals of Missouri Botanical Garden.75:1180-1206.
- Rodriguez JM, Berke T, Engle L, Nienhuis J (1999) Variation among and within Capsicum species revealed by RAPD markers. Theoretical and Applied Genetics. 99: 147-156.
- Spooner DS, Aanderson GJ, Jansen RK (1993) Chloroplast DNA evidences for the interrelationships of tomatoes, potatoes and pepinos (Solanaceae). American Journal of Botany 80: 676-688.
- Swofford DL (2002) PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods). Version 4. Sunderland: Sinauer Associates.
- Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positionspecific gap penalties and weight matrix choice. Nucleic Acids Research 22: 4673-4680.
- Vandenbussche M, Chambrier P, Rodrigues Bento S, Morel P (2016) *Petunia*, Your Next Supermodel?. Front. Plant Sci., ;7: 72.
- Wei JL, Hu XR, Yang JJ, Yang WC (2012) Identification of single-copy orthologous genes between physalis and *Solanum lycopersicum* and analysis of genetic diversity in physalis using molecular markers. *PLoS ONE* 7: 50164.
- Xin T, Yao H, Gao H, Zhou X, Ma X, et al. (2013) Super food *Lyciumbarbarum* (Solanaceae) traceability via an internal transcribed spacer 2 barcode. Food Research International, 54(2):1699-1704.
- Zamora-Tavares P, Vargas-Ponce O, Sanchez-Martinez J, Cabrera-Toledo D (2015) Diversity and genetic structure of the husk tomato (*Physalis philadelphica* Lam.) in Western Mexico. *Genet. Resour. Crop Evol.* 62: 141-153.
- Zhao S, Chen X, Song J, Pang X, Chen S (2015) Internal transcribed spacer 2 barcode: a good tool for identifying Acanthopanacis cortex. *Front. Plant Sci.* 6: 840.