# **Original Article**

# Spirulina platensis extract in alloxan-induced diabetic rabbits: a combined experimental study of antidiabetic activity in vivo and anticancer ability in vitro

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## Abstract

The present study was designed to study the efficacy of Spirulina platensis algal extract on hepatocellular carcinoma cell lines (HAMC Cell line), as well as its antidiabetic effect on alloxan-induced diabetic male rabbits. The study included the treatment of 24 male rabbits for four periods (7, 14, 21, 28) of days. The experiment included four groups of rabbits: the negative group, the positive group, the first treatment group and the second treatment group. Each group contained six rabbits. The negative (intact) and positive (diabetic induced), control groups were injected with 1ml of the normal saline and the first treatment group (diabetic induced) was injected with a concentration of 50 mg/kg of the algal extract. In contrast, the second treatment group (diabetic induced) was injected with a concentration of 100 mg/kg of the algal extract. The treatment results with algal extract showed a significant decrease in blood glucose levels in the first and second treatment groups at a level of probability (P<0.05) compared with the positive control group for all periods. The results also showed that the algal extract had efficacy against cancer, as the concentration of 17.2 µg caused the highest HAMC cell line inhibition rate at 62.23%. The current study concluded that Spirulina platensis had therapeutic effects in alloxan-induced diabetic rabbits as well as high potential inhibition of cancer cells.

Keywords: Spirulina platensis, rabbits, diabetes, alloxan, HAMC cell line.

# Introduction

Diabetes is a chronic metabolic disease characterized by a rise in the glucose level, i.e., more than the normal level, resulting from a complete or relative deficiency of the hormone insulin or because of factors against insulin action [1]. It is a chronic condition caused by various factors, including hereditary, viral, environmental, or functional [2]. Global estimations indicate that the proportion of adults with diabetes will increase to 69% in 2030 [3]. Many health problems result from taking the chemical drugs used to treat diabetes, and their prolonged use may cause side effects that negatively affect the body's organ functions [4]. In this regard, many studies were conducted to find natural treatment sources. Researchers have turned their attention to alternative treatments from natural products such as plants, herbs and algae, which can be more effective in decreasing glucose levels and reducing side effects compared with chemical drugs [5]. Algae, including cyanobacteria, have gained wide attention in recent years in this field due to the biologically active compounds that they contain [6].

Spirulina algae is one of the most important algae belonging to the cyanophytes/cyanobacteria group from the eukaryotic algae. S. platensis contains a high content of carotenoids, especially  $\beta$ -carotene and vitamins, including vitamins B3, B9, B12, B1, B2, B6, C, D, E and Folic acid [7]. Algae produce many bioactive compounds within their secondary metabolism. Moreover,



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secondary bioactive compounds in algae have an effective role as anticancer, antioxidants, and antihyperlipidemic and enhance the body's immunity [8–10]. Spirulina fusiformis significantly reduces high blood glucose levels at a dose of 400 mg/kg and has an antidiabetic effect in rats-induced diabetes [11]. The treatment of laboratory rats with an alcoholic extract of *Caulerpa* lentillifera showed a high ability to regulate hyperglycemia metabolism [12].

The study of Ghwenm et al. (2020) confirmed the efficiency of Chlorella vulgaris in reducing the glucose level in diabetic-induced mice after injecting them with concentrations of 40 and 80 mg/kg of the algal ethanolic extract for a month [13]. Sun et al. (2010) found the effective role of Chlorella protothecoides and Nitzschia laevis in reducing blood glucose among 20 selected green algae [14]. Roy et al. (2007) mentioned the possibility of inhibiting liver cancer in laboratory rats through the use of the substance phycocyanin that was extracted from S. platensis, as well as the polysaccharides which were extracted from the algae [15], also considered an active agent against stomach cancer [16]. Laungsuwon & Chulalaksananukul's study (2013) showed that the algal extracts of Cladophora glomerata and Microspora floccose were high efficiency against cancer diseases due to their containing a high percentage of chemically active compounds in killing cancer cells [17]. Many sulfated polysaccharides were extracted and diagnosed from Bryopsis sp. which was used as an antibiotic in lung cancer treatment [18].

Finally, in the light of the medicinal importance of algae and their widespread usage in medicinal and pharmaceutical products, and their contribution to the studies which deal with the effect of some species of algae as anticancer and hyperglycemia, the current project aimed to know the effect of S. platensis algal extract on blood glucose level in diabetic-induced male rabbits and its inhibitory effect on HAMC cell lines.

# **Material and methods**

## **Ethics declaration**

The College of Education for Pure Science, University of Basrah ethics committee reviewed and approved this study (No. 2020/4402). Animals were cared for in accordance with the Guide to the Care and Use of Experimental Animals by Canadian Council on Animal Care, Vol. 1, (2<sup>nd</sup> Ed.), (1993) and its revision version (2020).

## **Experiment animals**

A total number of 24 male rabbits (1250–1400 g) were used in the animal house of the Dep. Biology/Col. Education for pure science/Uni. Basrah and the animal house were maintained at a temperature of 20°C and humidity of 40% and provided on a 14:00-10:00-hour light: dark cycle. Rabbits were fed with food and water ad libitum. The animals were left fasting for at least 18:00 hours before the experiments were carried out. Male laboratory rabbits were randomly divided into four treatment categories and each category consisted of six rabbits. The negative control group included intact animals that were not induced by diabetes and were orally administered with 1ml of normal saline. The positive control group represented alloxan-induced diabetic animals and were orally administered with 1ml of normal saline. Whereas the first and second treatment groups contained rabbits induced by diabetes and were orally administered at doses of 50 and 100 mg/kg of algae extract, respectively.

## Induction of diabetes in laboratory animals

Diabetes was induced in laboratory rabbits after the food was withheld for 12 hours and then injected with the Alloxan (150 mg/kg for b.w) in the intraperitoneal cavity (IP) and the volume (0.1 ml) for each animal for three days. Drinking water was replaced with w/v (20%) of glucose to reduce hypoglycemia's shock due to alloxan treatment. After the first week of injection, the glucose level in the serum was measured to confirm the occurrence of diabetes [19].

## **Preparing doses**

Two concentrations of S. platensis extract were prepared. The highest dose represented 1/50 LD50, and it measured 100 mg/kg b.w., while the lowest dose represented 1/100 LD50 and it measured 50 mg/kg b.w., depending on the LD50 value of 5,000 mg/kg [20]. Moreover, laboratory rabbits were orally treated for 28 days, within one dose per day.

#### **Collection blood samples**

Blood samples were drawn weekly for a month directly from the heart using a 3-ml medical syringe. Blood samples were placed in container tubes on Heparin Lithium/Gel and left for 10–15 minutes to coagulate, then were placed in a centrifuge at 3,500 rpm for ten minutes

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Table 1: Impact of S. platensis extract treatment on serum glucose of alloxan-diabetic rabbits for 7 and 14 days (mean±standard deviation).

7 days od Glucose mg/dl	14 days Glucose mg/dl
110.65°±14.16	115.78°±10.56
414.45 <sup>a</sup> ±34.86	404.58 <sup>a</sup> ±45.72
211.53 <sup>b</sup> ±22.35	176.34 <sup>b</sup> ±29.24
195.69 <sup>b</sup> ±17.60	161.88 <sup>b</sup> ±21.02
i	7 days        Glucose mg/dl        110.65 <sup>c</sup> ±14.16        414.45 <sup>a</sup> ±34.86        211.53 <sup>b</sup> ±22.35        195.69 <sup>b</sup> ±17.60

Note: <sup>a, b, c</sup> – Different letters indicate the significance differences at (P $\leq$ 0.05).

Table 2: Impact of S. platensis extract treatment on serum glucose of alloxan-diabetic rabbits for 21 and 28 days (mean±standard deviation).

Treatments	Period	21 days Glucose mg/dl	28 days Glucose mg/dl
Negative control (normal saline	e)	108.25°±8.38	$112.56^{b} \pm 8.87$
Positive control group (alloxan)	)	393.86ª±12.50	378.88 <sup>a</sup> ±19.94
The first treatment group (50 m	g/kg)	$134.02^{b} \pm 14.09$	119.03 <sup>b</sup> ±16.84
The first treatment group (100 n	ng/kg)	140.32 <sup>b</sup> ±16.75	124.51 <sup>b</sup> ±14.57

Note: <sup>a, b, c</sup> – Different letters indicate the significance differences at ( $P \le 0.05$ ).

to obtain the serum, then pulled the serum separated from the coagulated blood and saved in special tubes at -10°C until serum glucose tests were performed [21].

## **Determination of serum glucose in rabbits**

The serum glucose concentration was estimated according to the enzymatic method Tietz, (1999) by the kit supplied by the British company Randox [22]. The absorbance of the sample is examined at a wavelength of 500 nm.

## **Estimating cancer cell inhibition**

All solutions, buffers and tissue culture media were prepared according to Freshney (2010) [23] by using the hepatocellular carcinoma cell lines (HAMC), which were obtained from the Iraqi Center for Cancer Research and Medical Genetics in Baghdad (ICCMGR).

## **Statistical analysis**

Data were analyzed using SPSS V.20 program. Differences between groups were calculated for statistical significance using One-way ANOVA. The data were analyzed at a probability level ( $P \le 0.05$ ).

# Results

The treatment of rabbits with two doses of algal extract (50&100 mg/kg) for 4 weeks caused a significant reduction in serum glucose level at ( $P \le 0.05$ ) when compared with the control group. Also, the glucose level in both groups treated with two doses of the algal extract was not significantly affected compared with the negative control group for a period of 28 days (Table 1 and Table 2).

The current results showed the inhibitory efficacy of the algal extract in the hepatocellular carcinoma cell lines in all concentrations, and the concentration of  $17.2 \,\mu$ g/ml was the most effective in killing cancer cells among the used concentrations in measuring efficacy against cancer Table (3).

Table 3: Effectiveness of the S. platensis extract on HAMC cell line.

Concentrations of algal extract (µg/ml)	Viability %	Inhibition %
6.4 µg/ml	44.56	55.43
10 µg/ml	45.27	54.72
17.2 µg/ml	37.76	62.23

# Discussion

The induction of diabetes by using the alloxan and at a dose of 150 mg/kg in male rabbits had a distinct role in raising the level of blood glucose by causing damage to the pancreas function, as the high level can infer in the blood serum to 400 mg/100 dl. This finding is consistent with the results of the researcher [24].

Treatment of male rabbits with the S. platensis ethanolic extract caused a significant decrease in the glucose level in the first and second treatment groups compared to the positive control group. This finding was consistent with a study by Mridha et al. (2010) [25]. The algal extract possesses many effective chemical compounds that have the ability to dissolve in water, such as saponins, glycosides, carbohydrates and phenols. These compounds have a significant effect in reducing the level of glucose in the blood, especially in animals with hyperglycemia, because these compounds possess the ability to stimulate the body tissues to uptake glucose from the blood, so the tissue's consumption of glucose increases [26].

In addition, terpenoids are considered antioxidants, as they prevent the oxidation of the hormone insulin via free radicals and prevent the formation of lipid peroxides that disturb the hormone insulin [27]. Moreover, they diminish glucose absorption in the intestine while stimulating the pancreas secretion of the hormone insulin and facilitating the entry of glucose into the cells of surrounding tissues and muscles [27]. As a result, these active compounds depend on a combination of antidiabetic mechanisms and measures, which include preventing diabetic nephropathy, stimulating insulin secretion from beta cells, inhibiting insulin catabolism, preventing oxidative processes and increasing glycogenesis and glycolysis [28–32].

The active compounds of Spirulina platensis, such as ascorbic acid, 9, 12-octadecadienoic acid – gamolenic acid – and hexadecanoic acid, have an important protective role against free radicals generation. These free radicals cause damage to the DNA, lipids and proteins, so these active compounds reduce the effect of free radicals from oxidation of the insulin hormone [10, 33]. The decrease in glucose may also be due to the presence of carotenoids and phycocyanin pigments that have a similar function to the action of ascorbic acid. They also have an antioxidant effect in removing free radicals and thus prevent pancreatic disorders by inhibiting apoptosis (programmed cell death) and suppressing free radicals production to alleviate Oxidative stress. In turn, these pigments reduce damage in the pancreas cells caused by alloxan treatment. In addition to their ability to reduce cell resistance to insulin and increase insulin sensitivity, they improve the function of beta cells and renew their construction, and more importantly, their ability to activate insulin receptors [7, 33–35]. Epidemiological and clinical algae studies showed an inverse correlation between antioxidant properties and the incidence of chronic diseases such as cardiovascular diseases associated with hyperlipidemia and diabetes [36].

Moreover, the composition of the algal ethanolic extract contains turbines and steroids that stimulate  $\beta$  cells in the pancreas to secrete insulin hormone, and these effective chemical compounds have pharmacological effects, as they are used as antithyroid, anticancer and antihypercholesterolemic. Interestingly, these compounds do not cause damage to the heart and liver functions [37]. In this context, the compound 9, 12, 15-Octadecatrienoic acid (Z, Z, Z) identified within the algal extract may increase the sensitivity of cells to the insulin hormone [10, 38].

The results of the current study showed the ability of the algal extract to inhibit the hepatic cell line (HAMC), as the highest rate of killing the cancer cells was 62.21% at a concentration of 17.2  $\mu$ g/ml. Perhaps the reason for this is that algal extract possesses many active compounds that act as antioxidants and play a role in suppressing cancer development in its last stages. The most active water-soluble antioxidants isolated from algae include phenols, vitamins and carotenoids [39]. In fact, oxidative stress and free radicals generation promote the process of carcinogenesis, while antioxidants, including Linolenic acid, cause scavenging reactive oxygen species (ROS), thereby preventing the emergence of cancer cells [20–40].

Some studies have confirmed the algal extract's protective role by inhibiting proliferation and stimulating apoptosis in treating cancer patients [41]. Cancer cells operate under oxidative stress, and their vitality can be impaired by scavenging free radicals via various antioxidants; therefore, antioxidants have a high potential to destroy tumors. In particular, the pathway of the nuclear factor erythroid-2-related factor 2 (Nrf2) controls the genetic expression of important cellular antioxidants and detoxification enzymes. It was found that active compounds isolated from algae appear to suppress the Nrf2 pathway. As a result, inhibition of this pathway in cancer cells is most effective in promoting their death [42].

Konickova et al. (2014) observed that S. platensis and algae-derived tetrapyrroles were highly potent as

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the anti-cell division in pancreatic cancer, and this result was similar to the results of the current study [8]. Interestingly, the bioactive substance tetrapyrroles can potentially improve the glutathione redox status, increasing tumor inhibitory efficacy [43]. In fact, the phycocyanobilin compound inhibits ROS production in mitochondria and is a strong inhibitor of the enzyme NADPH oxidase [8, 44]. The mitochondrial NADPH oxidase has induced superoxide, which represents a carcinogenesis factor affecting the cell cycle and several metabolic pathways [45, 46].

# Conclusion

The algae Spirulina platensis is rich in nutrients and active components that act as antioxidants. It has therapeutic effects due to its antidiabetic and anticancer activity in alloxan-induced diabetic male rabbits and it could play a significant role in suppressing cancer development in its last stages.

# **Conflict of interest**

The authors declare no conflict of interest.

# **Ethical approval**

The College of Education for Pure Science, University of Basrah ethics committee reviewed and approved this study (No. 2020/4402).

# Reference

- Saikat, D., Sekahar, B., Ranabir, S. & Subhash, M. (2008) Antidiabetic effect of matured fruits of Diospyros peregrine in alloxan induced diabetic rats. International journal of green pharmacy. 2(2): 95-99.
- 2. Chauhan, N.S. & Dixit, V.K. (2007) Antihyperglycemic activity of the ethanolic extract of Curculigo orchiides Gaertn. Phatmacognosy Magazine, 3(12):236-239.
- Maria-Luisa, L. & Cristina, F. (2013). Oxidative stress in diabetes mellitus and the role of vitamins with antioxidant actions. Agricultural and Biological Sciences. Chapter 9. 209-232. doi: 10.5772/51788
- Stein, S.A., Lamos, E.M. & Davis, S.N. (2012). A Review of the efficacy and safety of oral antidiabetic drugs. National Institutes of Health Public Access. 12(2): 153-175
- 5. Souza, P.M., Sales, P.M., Simeoni, L.A., Silva, E.C., Silveira, D. et al. (2012). Inhibitory activity of a-amilase and a-glukosidase

by plant extracts from the Brazilian Cerrado. Planta Medica. 78(4): 393–399.

- Dahms, H.U., Xu, Y. & Pfeiffer, C. (2006). Antifouling potential of cyanobacteria: a mini-review. Biofoul. 22(5):317–327.
- Chu, W., Lim, Y., Radhakrishnan, A.K. & Lim, P. (2010) Protective effect of aqueous extract from Spirulina platensis against cell death induced by free radicals. BMC Complement Altern. Med. 10, 53.
- Konickova, R., Vankova, K., Vanikova, J., Muchova, L., Subhanova, I. et al. (2014). Anticancer effects of blue-green alga Spirulina platensis, a natural source of bilirubin-like tetrapyrrolic compounds. Annals of hepatology,13(2):273-283.
- 9. Ngo-Matip, M.E., Pieme, C.A., Azabji-Kenfack, M., Moukette,.B, Korosky, E. et al. (2015).Impact of daily supplementation of Spirulina platensis on the immune system of naïve HIV-1 patients in Cameroon: a 12-months single blind, randomized, multicenter trial. Nutr. J. 14:70.
- Kata, F.S., Athbi, A.M., Manwar, E.Q., Al-Ashoor, A., Abdel-Daim, M.M. & Aleya, L. (2018) Therapeutic effect of the alkaloid extract of the cyanobacterium Spirulina platensis on the lipid profile of hypercholesterolemic male rabbits. Environmental Science and Pollution Research, 25(20):19635-19642.
- Simon, J.P., Baskaran, U.L., Shallauddin, K.B., Ramalingam, G. & Evan, P.S. (2018) Evidence of antidiabetic activity of Spirulina Fusiformis against streptozotocin-induced diabetic wistar albino rats. 3 Biotech.8(2):129.
- Sharma, B.R. & Rhyu, D.Y. (2014). Antidiabetic effects of Caulerpa lentillifera: stimulation of insulin secretion in pancreatic β-cells and enhancement of glucose uptake in adipocytes. Asian Pacific journal of tropical biomedicine, 4(7): 575-580.
- Ghwenm, S.S., Kata, F.S. and Athbi, A.M. (2020). Hypoglycemic and antioxidant effect of the ethanol extract of Chlorella vulgaris in alloxan-induced diabetes mice. Biochem. Cell. Arch. 20(2):3535-3542.
- Sun, Z., Peng, X., Liu, J., Fan, K., Wang, M. et al. (2010). Inhibitory effect of microalgal extraction on the formation of advanced glycation endoproducts (AGEs). J. Food Chem. 120(1):261-267.
- Roy, K.R., Arunasree, K.M., Reddy, N.P., Dheeraj, B., Reddy, G.V. et al. (2007). Alteration of mitochondrial membrane potential by Spirulina platensis C-phycocyanin induces apoptosis in the doxorubicin resistant human hepatocellular-carcinoma cell line HepG2. Biotechnol. Appl. Biochem., 47(3):159-167.
- Oh, S.H., Ahn, J., Kang, D.H. & Lee., H.Y.(2011). The Effect of ultrasonificated extracts of Spirulina maxima on the anticancer activity. Mar. Biotechnol. (NY)., 13(2):205-214.
- Laungsuwon, R. & Chulalaksananukul, W. (2013). Antioxidant and anticancer activities of freshwater green algae Cladophora glomerata and Microspora floccose, from Nan River in northern Thailand. Maejo Int. J.Sci. Technol., 7(2):181-188.
- Smit, A.J. (2004) Medicinal and pharmaceutical uses of seaweed natural products : a review. Journal of applied phycology, 16(4):245-262.
- Misra, M. & Aiman, U. (2012) Alloxan: An unpredictable drug for diabetes induction? Indian J Pharmacol. 44(4): 538–539.
- 20. Altug, T. (2003). Introduction to toxicology and food. CRC Press, Boca Raton, FL.pp.256.
- Tiedge, M., Richter, T. & Lenzen, S. (2000) Importance of cysteine residues for the stability and catalytic activity of human pancreatic beta cell glucokinase. Arch Biochem Biophys. 375(2):251-60.

- Tietz, N.W. (1995). Clinical Guide to Laboratory tests. 3<sup>rd</sup> ed. Philadelphia. WB. Saunders, 268-273.
- 23. Freshney, R.I.(2010). Database of misidentified cell lines. International journal of cancer, 126(1):302-304.
- 24. Yakubu, O.E., Otitoju, O. & Okwara, D.(2017).Comparative hepatoprotective and haematological effect of the ethanol extracts of the leaves of vitex doniana and Senna occidentalis in alloxan- induced diabetic in male wister rats. J Anal Pharm Res 4(2):00099.
- Mridha, M.O.F., Jahan, M.A.A., Akhtar, N., Munshi, J.L. & Essa,
  Z. (2010) Study on Hypoglycaemic Effect of Spirulina platensis on Long -Evans Rats. Bangladesh J.Sci.Ind.Res.45(2), 163-168.
- Mizoguchi, T., Takehara, I., Masuzawa, T., Saito, T. & Naoki, Y. (2008) Nutrigenomic studies of effects of Chlorella on subjects with high-risk factors for lifestyle-related disease. Journal of medicinal food, 11(3):395-404.
- 27. Hui, H., Tang, G., Liang, V.& Go, W. (2009). Hypoglycemic herbs and their action mechanisms. Chinese Medicine, 4:11.
- Miura, T., Itoh, C., Iwamoto, N., Aato, M., Kawai, M. et al. (2001). Hypoglycemic activity of the fruit of the Momordica charantia in Type 2 diabetic mice. J. Nutr Sci Vitaminol, 47 (5): 340-344.
- Eddouks, M., Magharani, M., Lemhadri, A., Ouahidi, M.L. & Jouad, H. (2002). Ethnopharmacolgical survey of medicinal plants used for the treatment of diabetes mellitus, hypertension and cardiac disease in the south – east region of morocco (Tafilalet). J. Ethnopharmacol; 82 :97-103.
- Esmaeili, M. A.& Yazdanparast, R. (2004). Hypoglycemic effect of teucrium polium: Studies with rats pancreatic islets. J Ethnopharmacol; 95:27-30.
- Mukherjee, P.K., Maiti, k., Mukherjee, k. & Houghton, P.J. (2006). Leads from Indian medicinal plants with hypoglycemic potentials. J. Ethnopharmacol,106(1):1-28.
- Bartosikova, L., Necas, J., Such, S.V.& Stanadov, V. (2003). Monitoring of antioxidation effect of mornine in alloxan- induced Diabetes mellitus in laboratory rat. Acta vet Brno. 72: 191- 200.
- Takaichi, S. (2011) Carotenoids in algae: distributions, biosynthesis and functions. Mar Drugs 9(6):1101–1118.
- Hozayen, W.G., Mahmoud, A.M., Soliman, H.A. & Mostafa, S.R. (2016). Spirulina versicolor improves insulin sensitivity and attenuates hyperglycemia-mediated oxidative stress in fructose-fed rats. J Intercult Ethnopharmacol. 5(1): 57–64.
- 35. Rani, Y., Gondo, H. & Indahsari, N.K. (2019) The Effect of spirulina on Apoptosis (Stored Biology Materials) To Pregnant Rat Wistar in the Second Trimester Which is Induced By IL-6. IOP Conference Series Earth and Environmental Science 217(1):012042.

- Belay, A. (2002). The Potential Application of Spirulina (Arthrospira) as a nutritional and therapeutic supplement in health management. Journal of the American Nutraceutical Association, 5(2):26–48.
- Saeidnia, S., Manay, A., Gohari, A. & Abdollah, M. (2014). The story of Beta –sitosterol – A Review. European Journal of Medicinal plants 4(5): 590-609.
- Rajaram, S. (2014).Health benefits of plant- derived α-linolenic acid, Am. J.Chin Nutr,100(1): 443S-448S.
- Plaza, M., Santoyob, S., Jaimeb, L., Garcia-Blairsy, R.G., Herrerob, M. et al. (2010). Screening for bioactive compounds from algae. J. of Pharm. And Biomed. Analysis, 51(2):450-455.
- Kohen, R. & Nyska, A. (2002). Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. Toxicology Pathology, 30(6):620-50.
- Azamai, E. S., Sulaiman, S., Habib, S. H., Looi, M. L., Das, S. et al. (2009). Chlorella vulgaris triggers apoptosis in hepatocarcinogenesis-induced rats. Journal of Zhejiang University Science B., 10(1): 14-21.
- Sznarkowska, A., Kostecka, A., Meller, K., & Bielawski, K.P. (2017). Inhibition of cancer antioxidant defense by natural compounds. Oncotarget, 8(9):15996-16016.
- Perchellet, J.P., Perchellet, E.M., Orten, D.K. & Schneider, B.A. (1989) Decrease ratio of reduced/oxidized glutathione in mouse epidermal cells treated with tumer promoters. Carcinogenesis, 7:503-6.
- 44. Zheg, J., Inoguchi, T., Sasaki, S., Maeda, Y., McCarty, M.F. et al. (2013) Phycocyanin and Phycocyanobilin from Spirulina platensis protect against diabetic nephropathy by inhibiting oxidative stress. Am J Physiol Regul Integr Comp Physiol. 304(2):R110-R120.
- Antio, A.V., Elguero, M.E., Poderoso, J.J.& Carreras, M.C. (2012) Mitochondrial regulation of cell cycle and proliferation. Antioxid Redox Signal.16:1150-80.
- Nazarewicz, R.R., Dikalova, A., Bikineyeva, A., Ivanov, S., Kirilyuk, I.A. et al. (2013). Does scavenging of mitochondrial superoxide attenuate cancer prosurvival signaling pathway ? Antioxid Redox Signal.19(4):344-9.