



The effect of some plant extracts on the infection of okra plant *Abelmoschus esculentus* with root rot disease

Tsyar Mohammad Khudair and layla Abdul Raheem benyan
Plant Protection Department, College of Agriculture, University of Basrah, Iraq

Abstract

The study aimed to know the best treatment to control the fungi that cause okra root rot by using aqueous extract of the leaves of eucalyptus, Bamber (*Cordia myxa*) and oleander plants, with finding the best concentrations to inhibit the growth of pathogenic fungi. The results showed the presence of three pathogenic fungi *R. solani*, *F. oxysporum* and *M. phaseolina*, and the most prominent of them were *R. solani* in the areas from which samples were collected in Basrah province. The results showed that the fungus *R. solani* had a clear effect in reducing the percentage of germination of Cabbage seeds, which amounted to 32.7%, with a significant difference from the treatment of *F. oxysporum* and *M. phaseolina*, which amounted to 55.0 and 57.7%, respectively, while it reached 63.93% in the control treatment. The results of the pot experiment showed that the fungus *R. solani* reduced the germination rate of okra seeds, reaching 57.7%, while it was 58.2 and 64.4% in the two fungi *F. oxysporum* and *M. phaseolina* treatments, with a significant difference from the 90% control treatment. The fungi also caused the death of seedlings, where the percentage reached 28.6% for *R. solani* and *F. oxysporum* and 19.0% in *M. phaseolina* treatment with a significant difference from 0% control treatment. The aqueous extracts affected the percentage of pathogenic fungi growth in the laboratory. The aqueous extract of the leaves of the eucalyptus plant gave the highest percentage of inhibition of the growth of the fungus *R. solani*, the percentage was 43.11%, with a significant difference from the extracts of the leaves of the plant Bamber (*Cordia myxa*) and oleander, which amounted to 39.62 and 29.68%, respectively. The effect of the extracts increases with the increase of the concentration, where the aqueous extract of eucalyptus at a concentration of 40% gave the highest percentage of inhibition which amounted to 73.89%, with a significant difference from the extracts of Bamber (*Cordia myxa*) and oleander.

3340

Keywords: plant extracts, okra plant, *Abelmoschus esculentus*, root rot disease

DOI Number: 10.14704/nq.2022.20.5.NQ22635

NeuroQuantology 2022; 20(5):3340-3347

Introduction

Okra (*Abelmoschus esculentus*) is a summer vegetable crop belonging to the family Malvaceae. It is native to Africa and has spread to the eastern Mediterranean, India, the Arabian Peninsula,

America, Europe and other countries. It is one of the most desired vegetables in Iraq that needs a warm growing season to grow (Wanted et al, 1989). It is considered one of the plants of high nutritional value, where its fresh fruits contain a high



percentage of water and many important nutrients such as carbohydrates, calcium, phosphorous, iron, sodium and magnesium (Gemede et al., 2015). The okra plant is infected with many agricultural pests such as insects such as aphids, cutworms, and various fungal diseases that have increased with protected agriculture, and the most important diseases are powdery mildew, wilt and root rot, as well as snake worms, Root rot disease is considered one of the common diseases that cause great losses caused by many fungi in the soil, including *Rhizoctonia solani*, and one of the symptoms of this disease is the rotting of the roots and the bases of the stems that are close to the surface of the soil and thus leads to weak plants and thus wilt and plant death. The plant, which caused losses in the crop (Jassim 2012). The fight against these causes is one of the difficulties facing workers in this field due to its ability to remain in the form of static structures for a long time and its ability to restore dead organic matter when the host is not available (Agrios, 2007). Several methods have been used to combat soil fungi, including chemical control, which is the most common method because it is easy to use and has a quick effect on pathogens, but its negative effects are due to repeated use and the emergence of resistance in addition to its danger to humans and animals and environmental pollution (Taylor et al., 2002). Many strategies have been used to control diseases caused by fungi such as *Rhizoctonia solani* and *Fusarium oxysporum*, such as soil sterilization, planting resistant varieties, and following crop rotations, but they did not achieve effective control of these pathogens (El-mohamedy et al., 2011). Recently, plant extracts have been used as an important part in the biological control of plant pathogens, and several attempts have been made to use them in the control of plant pathogenic fungi such as *Macrophomina phaseolina* and *Fusarium oxysporum* and *R. solani* (Al-Rubaie, 2008). This is due to its abundance and being of low toxicity compared to chemical pesticides, as well as being easy to decompose (Harish et al., 2004). The leaves, flowers, and roots of many plants contain chemical compounds that have a toxic effect on agricultural pests (insect, bacterial and fungi), such as the inhibitory effect of the ethanolic extract of

eISSN 1303-5150

Nerium oleander on four types of important plant pathogenic fungi *R. solani*, *F. oxysporum* and *A. alternate*. The extract has a highly effective effect on inhibiting *F. solani* and *F. solani* species. *oxysporum* (Hadizadeh et al., 2009.). The eucalyptus plant is also one of the fast-growing trees and is used for medicinal and therapeutic purposes (Mubark, 2015), as its leaves contain phenolic compounds such as sterols, saponins, tannins, flavonoids and triterpenoids (Shayoub et al., 2015). Al-Khair and Al-Obaidi (2020) also referred to the study of the effect of two types of alcoholic and aqueous extracts of eucalyptus leaves on the growth of isolates of *M. phaseolina*, *R. solani* and *F. solani*, as the alcoholic extract was excelled in inhibiting the growth of isolated fungi except for *R. solani*, which was only significantly inhibited by the aqueous extract of eucalyptus leaves. The Bamber (*Cordia myxa*) plant is one of the plants grown in central and southern Iraq, native to Burma and used to combat pathogens in soil (Chacravarty, (1976). Where between Sarhan and Al-Shibli, 2003)) in a study of the effect of three concentrations of 0.5, 0.10 and 0.15% for each extract of Bamber (*Cordia myxa*) (cold aqueous extract, hot aqueous extract and alcoholic extract) on radial growth, dry weight and spore germination percentage of a number of plant pathogenic fungi *Fusarium solani* And *Alternaria alternate*, *Rhizoctonia solani*, *Helminthosporium sativum* and *Rhizopus stolonifer*, where the hot aqueous extract recorded a remarkable effect, followed by the cold aqueous extract, while the alcoholic extract did not have any significant effects. In view of the importance of the okra plant for being one of the vegetables with high nutritional and marketing value, the study included isolating and diagnosing the fungal pathogens and finding the best treatment to combat the pathogenic fungus by using aqueous extract of the leaves of eucalyptus, Bamber and oleander plants with finding the best concentrations to inhibit the growth of pathogenic fungi.

Materials and methods:-

Isolation and identification of pathogens

The pathogenic fungi *F. oxysporum*, *M. phaseolina* and *solani*.R were isolated from the roots of the okra plant. Samples of the roots of the okra plant

3341



that showed symptoms of yellowing and wilting were collected from different regions of Basrah province, including Abu Al-Khasib, Al-Bahadria, Al-Sahel, Al-Qurna District, Al-Sharsh and Shatt Al-Arab region Al-Hawtah was placed in marked plastic bags and brought to the laboratory. The roots were washed with running water for half an hour to get rid of the mud stuck to them, then they were cut to a length of (1 cm) the cuts were superficially sterilized with a solution of 10% sodium hypochlorite (Naocl) solution at a concentration of 6% of the commercial preparation for 2-3 minutes, then It was washed with sterile distilled water to remove the remnants of the sterilization solution for 2 minutes and left on filter paper for the purpose of drying. They were cultured in Petri dishes with a diameter of 9 cm containing sterile PDA medium supplemented with the antibiotic Chloramphenicol in 3 replicates at 250 mg/L. The dishes were incubated in the incubator at 25±2°C for 4 days. Then the isolates of fungi were purified by transferring parts of the tip of the fungal hyphae of the colony of pathogenic fungi using a sterile needle to Petri dishes containing sterile PDA media, and the plates were incubated in the incubator under the same temperature for 4 days for the fungi isolated after adding lactophenol dye by Prof. Dr. Yahya Ashour Saleh according to the taxonomic keys contained in (Parameter and Whitney, 1970), the fungal isolates were kept on the PDA medium at a tilt in the refrigerator until completion of the experiments.

Testing the effect of the pathogenic fungi *F. oxysporum*, *M. phaseolina* and *R. solani* using Cabbage seeds on the aquatic aquarium culture media:-

The germination rate of the seeds of Cabbage was tested treated with isolates of the pathogenic fungi *R. solani*, *F. oxysporum* and *M. phaseolina* according to the method of Bolkan and Butler, 1974) The nutrient medium was prepared Water Agar after sterilization with the Autoclave steam device and the antibiotic was added to it Chloramphenicol and shaken well and then poured into Petri dish with a diameter of 9 cm was then left to harden and then the center of each dish was inoculated with a disk of fungi(0.5) cm diameter from a colony of fungi

grown on PDA medium at the age of 7 days, then the dishes were incubated at a temperature of 25±2°C for a period of 3 days. After this period, the broccoli seeds were superficially sterilized with a solution of sodium hypochlorite at a concentration of 10% of the commercial solution 6% and in a circular motion near the edge of the dish and at a rate of three replications and at a rate of 10 seeds for each dish, leaving a control treatment containing broccoli seeds and without any pathogenic fungi. The dishes were incubated at a temperature of 25±2°C, and the results were recorded after (7 days) of planting by calculating the percentage of germination as in the following equation:

germination percentage = $\frac{\text{(number of germinated seeds)}}{\text{(total number of seeds)}} \times 100$

Pathogenicity test of the isolated fungi *R. solani*, *F. oxysporum* and *M. phaseolina* on potted okra plants:-

The pathogenicity of isolates of the pathogenic fungi *R. solani*, *F. oxysporum* and *M. phaseolina* was tested, using agricultural soil sterilized with formalin solution (1 part formalin: 50 parts water),The solution was used at a ratio of 3 L/m³ (Poultry, 1975). The soil was placed in plastic bags and closed well for 3 days. Then it was exposed to the air under the sun for 3 days in order to volatilize formalin. A quantity of peat moss was sterilized in the same way as in sterilizing the soil, mixing the peat moss with the soil in a ratio of 1:3 (soil: peat moss).Then the fungal pollen grown on millet seeds was added to it at a ratio of 1% w/w and the fungal pollen was mixed well with the sterilized soil using cellophane bags to ensure the homogeneity of the vaccine with the sterilized soil. By 5 seeds per pot and an average of three replicates, leaving three replicates as a comparison treatment.After the appearance of the symptoms of wilting and yellowing, the percentage of germination and the percentage of seedling death were calculated after 10 and 20 days of planting.

Germination% = $\frac{\text{(number of germinated seeds)}}{\text{(total number of seeds)}} \times 100$

seedling death% = $\frac{\text{Number of dead seedling}}{\text{Total number of seedlings}} \times 100$



Preparation of the aqueous extract of the leaves of plants used in the experiment

Metspula et al. (2001) used the method of preparing the aqueous extracts of the plants used in the study, which are the leaves of eucalyptus, Bamber (*Cordia myxa*) and oleander. 20 gm of dry plant powder was taken. Placed in glass flasks of 500 ml, each flask containing 200 ml of distilled water, then mixed the dry plant material in a Panasonic electric mixer for 15 minutes, then left for 30 minutes, then filtered with a dull cloth to separate the large plankton, Then centrifugation was conducted at 3000 cycles for 30 minutes, then the filtrate was taken and filtered through Whatman No.1 filter paper and sterilized using Millipore microfilters of 0.45 µm diameter. An appropriate amount of each plant extract was taken and the filtrate was used as a base solution. It was stored in the refrigerator at 4°C until use.

The effect of the aqueous extract of the tested plants on the growth of the pathogenic fungus *R. solani*

The medium of the PDA culture medium was prepared and sterilized by an Autoclave device at a temperature of 121 °C and a pressure of 15 pounds / inch for 20 minutes. After decreasing the temperature, the aqueous extract was added to the glass beaker containing the food medium at concentrations of 10, 20, 30 and 40% for each extract, then the beaker was shaken for the purpose of homogenization of the extract with The culture

medium, then poured into Petri dish 9 cm in diameter, The plate was inoculated upon solidification of the nutrient medium with a 0.5 cm diameter disc from the growing medium containing the pathogenic *R. solani* fungus and with three replications for each concentration of the extract. Then the plates were incubated at a temperature of 25 ± 2 °C for a period of 7 days. According to the growth rate, the average of two perpendicular diameters was taken for the growth of the fungal colony from the bottom plate, and the percentage of inhibition was calculated according to the following equation:

$$\text{Percentage of inhibition} = \frac{(\text{The growth rate of the fungus in the treatment} - \text{the growth rate of } (\text{The growth rate of the fungus in the treatment})}{\text{The growth rate of the fungus in the treatment}} \times 100$$

Results and Discussion

Isolation and diagnosis

The results of isolation and diagnosis Table (1) for okra plants infected with root rot and plant death showed the presence of three pathogenic fungi, *F.oxysporum*, *R.solani* and *M.phaseolina*, and the most prominent ones were in the areas from which samples were collected in Basra Governorate, namely, Abu Al-Khasib, Hamdan, Al-Sahel and Al-Houta Shatt areas. Arabs, Bahadriya and Qurna. Its phenotypic and microscopic characteristics were consistent with what was mentioned by (Watnabe and Shiyome, 1975) and (Sinclair, 1982, Leslie and Summurel, 2006).

3343

Table (1) shows the pathogenic fungi isolated from the roots of okra plants grown in different areas of Basrah province.

No.	Region	pathogenic fungi
1	Abi Al-Khasib (Hamdan)	<i>M. phaseolina</i> <i>R. solani</i>
2	alsaahil region	<i>R. solani</i> <i>M. phaseolina</i> <i>F.oxysporum</i>



3	Al-Houta	<i>R. solani</i> <i>M. phaseolina</i>
4	Al-Qurna / Shark region	<i>R. solani</i> <i>M. phaseolina</i> <i>F.oxysporum</i>
5	Al-Bahadria	<i>R. solani</i> <i>F.oxysporum</i>

Pathogenicity of the pathogenic fungi *F.oxysporum*, *R.solani* and *M.phaseolina* in germination percentage:-

Effect of pathogenic fungi on germination rate on Cabbage seeds:

The results of this experiment showed Table (2) that the fungi isolated *R.solani*, *M.phaseolina* and *F.oxysporum* caused a reduction in the germination rate of Cabbage seeds, but the effect of the pathogenic fungus *R.solani* in reducing the percentage of seed germination was greater than the effect of other fungi. The germination rate reached 32.7% compared to the fungi *F.oxysporum* and *M.phaseolina*, which amounted to 55.0% and 57.5%, respectively, compared to the control treatment, which had a germination rate of 63.93%. This is due to the ability of the pathogenic fungus *R. solani* degrading enzymes such as Polygalacturonase, Cellulases and Pectinases, as these enzymes work on decomposing plant cell

walls, as well as destroying the middle lamina (Ravjit et al., 1999). (Weinhold and Sinsclair, 1996),

Or, the reason may be due to the fungus' invasion of the seed coat and growth between the starch granules and the secretion of enzymes Amylases that lead to killing cells and turning the seeds into a dark brown color (Mahmoud et al., 2007). The reason is attributed to the mycotoxins that the fungus produces that inhibit seed germination and its ability to secrete kinase-degrading enzymes in the host cell wall, and this is consistent with what was found by Gordon et al, 1989). As for the fungus *M.phaseolina*, the results agreed with (Hassan et al., 2015), as the pathogenic fungus attacks the seeds of many plant families and leads to preventing them from germination and rotting because it secretes toxic compounds that lead to the killing of embryos, as the fungus secretes many enzymes that degrade cellulose and protein, which in turn It causes seed rot and infection.

Table (2) Effect of pathogenic fungi on the percentage of germination of Cabbage seeds

germination%	Tretments
32.7	<i>R.solani</i>
55.0	<i>F. oxysporum</i>
57.7	<i>M.phaseolina</i>
63.93	Control
27.35	(0.01)L.S.D**

*Each number represents an average of three replicates

Testing the pathogenicity of the pathogenic fungi *R.solani*, *M.phaseolina* and *F.oxysporum* in the percentage of germination and seedling death of okra plants in pots:

The results of the study showed in Table (3) that the pathogenic fungi *R.solani*, *F.oxysporum* and *phaseolina.M* had a clear effect in reducing the percentage of germination of okra seeds, where the

germination percentage in the treatment of the fungus *R.solani* was 57.7%, while it was 58.2% and 64.4% In the two treatments of *F.oxysporum* and *M.phaseolina*, respectively, with a significant difference from the control treatment, which amounted to 90.0%, The results also showed the ability of these pathogenic fungi to infect seedlings after germination, where the percentage of seedling



death in the treatment of the pathogenic fungi *R.solani* and *F.oxysporum* at an average of 28.6%, while it reached 19.0% in the treatment of *M.phaseolina* with a significant difference from the control treatment. 0%, These results agreed with previous studies regarding the ability of the fungus *R.solani* to reduce the percentage of germination of seeds of many crops such as okra, sesame, cotton and tomato (Fayyad and Al-Atabi, 2018), because the fungi *R.solani* and *F.oxysporum* cause a decrease in the germination rate of plant seeds. Al-Haidari okra, (2007) because the fungus *R.solani* secretes enzymes that degrade plant cell walls: Proteinate, Cellulase and Pectinases and secrete

toxins that kill the protoplasm of plant cells. (Kawchuk, 2002). The reason for the appearance of symptoms of infection on seedlings infected with *F.oxysporum* fungus, where the plant wilt with yellowing leaves was observed. The reason may be due to the fungus entering the plant tissue by penetrating the cell walls due to its ability to produce enzymes degrading chitinase, cellulose and pectin that break down cell walls (Booth, 1971). Also, *M.phaseolina* has the ability to produce vasodilating toxic substances that break down cellulose and pectin, causing rotting of seeds and death of seedlings (Mahato et al., 1987).

Table (3) Pathogenicity of the pathogenic fungi *R.solani*, *M.phaseolina* and *F.oxysporum* on germination of okra seedlings and seedling death

fungi	seedling death%	germination%
<i>R.solani</i>	28.6	57.7
<i>F.oxysporum</i>	28.6	58.2
<i>M.phaseolina</i>	19.0	64.4
Control	0	90.0
L.S.D*	22.6	15.11

3345

Effect of aqueous extracts on the growth of pathogenic *R. solani* . fungus

The results of this experiment showed Table (4) that the type of extract had a significant effect on inhibiting the growth of pathogenic fungi, where the eucalyptus extract gave the highest percentage of inhibiting the growth of the pathogenic fungus *R. solani* with a rate of 43.11%, with a significant difference from the aqueous extracts of Bamber (*Cordia myxa*) and oleander , with a percentage of 39.62 and 29.68%, respectively. The results also showed that the effect of the aqueous extracts of all studied plants increases with the increase in the concentration of the extract, as the percentage rate of inhibition reached 19.75, 25.68, 43.08 and 61.06% for the 10, 20, 30 and 40% concentrations,

respectively. Also, the aqueous extract of eucalyptus at a concentration of 40% gave the highest percentage of inhibiting the growth of the pathogenic fungus *R. solani*, which was 73.89%, with a significant difference from the extracts of Bamber (*Cordia myxa*) and oleander . Previous studies indicated that the aqueous extract of eucalyptus is effective in reducing the growth rate of pathogenic fungi *R.solani* and *F.solani* (Al-Hamiri, 2016). He also due (Al-Khair and Al-Obaidi, 2020) the superiority of eucalyptus leaf extract and its effect on root rot fungi of pine seedlings, because eucalyptus leaves contain Triterpeoids, sterols and flavonoids, which have an effective role in inhibiting the growth of pathogenic fungi.

Table (4) Effect of aqueous extracts of some plants on the growth of the pathogenic fungus *Rhizoctonia solani* in the laboratory

Effect average of extract type	fungus growth inhibition%				extract
	Concentrations used				
	%40	%30	%20	% 10	



39.62	54.99	54.44	29.63	19.44	<i>Cordia myxa</i>
43.11	73.89	41.48	32.03	24.07	<i>Eucalyptus</i>
29.68	54.29	33.33	15.37	15.74	<i>Nerium oleander</i>
	61.06	43.08	25.68	19.75	Average effect of concentrations
The least significant difference (L.S.D) test at the level of significance 0.01					
Interaction		concentrations		extract	
8.68		5.01		4.34	

References

Al-Humairi, Yasser Nasser Hussein (2016). Integrated control of root rot pathogens and the bases of potato stems in central Iraq. *Karbala Journal of Agricultural Sciences*. Volume 3 Issue 4.

Al-Haidari, Ali Ajel Jassim (2007). Isolation and identification of some fungi that cause seed rot and death of okra plants and their resistance by different techniques with the fungus *Trichoderma harzianum* Rifai. Master's thesis - College of Agriculture - University of Kufa, 122 pages.

Al-Khair, Anwar Nouri Muhammad and Muhannad Hamid Younis Al-Obaidi (2020). Effect of aqueous and alcoholic extracts of leaves of eucalyptus *camaldulensis* trees Dehn *Eucalyptus* on the growth of root rot fungi of *Ten brutia* Pinus seedlings in vitro. *Journal of Education and Science*, 29(1): 75-92.

Al-Rubaie, Afrah Abd Ali. (2008). Manufacture of a biocidal from *Bacillus circulans* vaccine to combat some pathogens of wheat root rot. Master Thesis. College of Technology / Al-Musayyib, Technical Education Authority. 111 pages.

Hassan, Abdullah Abdul Karim and Walid Khalid Ahmed (2015). Evaluation of the efficiency of fermented organic fertilizer and biological factors in controlling charcoal rot disease caused by the fungus *Macrophomina phaseolina* (Tassi) Codi. on yellow corn. *Tikrit University Journal of Agricultural Sciences*, Volume (15), Issue (3).

Sarhan, Abd al-Ridha Taha and Majid Kazem About al-Shibli (2003). Comparative study of the effect of amber extract and fungicides on some plant pathogenic fungi. *Iraqi Journal of Science*. Volume 44, Issue (1), pg. 68-83.

Tawajen, Ahmad Muhammad Musa (1975). *Greenhouse Environment*, Basra University Press. 571 - 573.pg.

Fayadh, Muhammad Amer and Saja Sabih Khudair Al-Atbi (2018). Evaluation of the effectiveness of extracts of some plants from the southern Iraqi marshes in controlling the death of okra seedlings caused by the fungus *Rhizoctonia solani* and the semi-fungus *Pythium* sp. . Master Thesis . faculty of Agriculture . Albasrah university . 104 pages. matlub

, Adnan Nasrouzuddin Sultan Muhammad and Karim Salih Abdoul (1989). *Vegetable Production, Part II, Revised Version*. Mosul University, Mosul Higher Education Press, second edition. Iraq. 337 pages.

Jassim, Naji Salem (2012). Evaluation of the efficiency of salicylic acid (SA) and the fungus *Trichoderma harzianum* in resisting root rot disease in okra caused by the fungus *Rhizoctonia solani*.

Agrios.G.N.(2007).Plant pathology .4 Ed..Academic Prees 606 pp, New York.U.S.A.

Bolkan , H.H. and Butler , E.E. (1974). Studies on Heterokaryosis virulence of *Rhizoctonia solani*. *Phytopathology*. 64 : 513-522.



Booth, C. (1971) .The Genus *Fusarium* common wealt. Institute, Kew, Surrey, England.

Chakravarty, H.L. (1976). Plant wealth of Iraq. Ministry of Agriculture & Agrarian Reform, Baghdad, 304.

El-Mohamedy R.S.R.; Abd El-Samad, E.H.A.; Habib , H.A.M.; and El-Bab, T.S.F.(2011).Effect of using biocontrol agents on growth, yield head quality and root rot control in broccoli plants intern.J.of acad. Res.3(2):71-81.

Gemedede, H. F.; Ratta, N.; Haki, G.; Woldegiorgis, A. Z. and Beyene, F. (2015). Nutritionnal quality and health benefits of okra (*Abelmoschus esculentus*):A review.J.Food P|rocess Technol 66PP.

Gordon, T.R., Okamoto, D., and Jacobson, D.J.(1989). Colonization of Muskmelon and Nonsusceptible Crops by *Fusarium oxysporum f. sp. melonis* and Other Species of *Fusarium*. Phytopathology 79:1095-1100.

Hadizadeh I., B. Peivastegan, M. Kolahi (2009). Antifungal activity of nettle (*Urtica dioica* L.), colocynth (*Citrullus colocynthis* L. Schrad), oleander (*Nerium oleander* L.) and konar (*Ziziphus spinachristi* L.) extracts on plants pathogenic fungi. Pakistan J Biol Sci. 1;12:(1):58-63.

Harish, S.; Saravanan, T. and Radjacomare, R.(2004). Mycotoxic Effect of Seed Extracts against *Helminthosporium oryzae* the Incitant of Rice Brown Spot. J. Boil. Sci., 4:366-369.

Leslie , J.F. and Summerell, B.A. (2006) . The *Fusarium* Laboratory Manual photographs by Suzanne Bullock.388 pp.

Linglis, G.D.; and Kawchuk, L.M.(2002). Comparative degradation of oomycetes, ascomycetes, and basidiomycetes cell walls by mycoparasitic and biocontrol fungi . Can.J.Microbiol. 48: 60-70.

Mahmoud , Y. G.; Gaafar , R. M. and Mubarak , H. M. (2007). Genetic Diversity among Nile delta isolates of *Rhizoctonia solani* Kuhn based on Pathogenicity, Compatibility, Isozyme Analysis and total protein pattern. Journal Botany. 31 :19-29.

Mahato, S. B., Siddigui, K. A. I., Bhattacharya, G., Ghosel, T., Migahara, K., Sholichin, M. T. and Kawasaki, T. (1987). Structure and stereo. Chemistry of phaseolinic acid: A new acid from eISSN 1303-5150

Macrophomina Phaseolina. J. Nat. Prd. 50: 245 – 247.

Metspalu L. ; Hiiesaar K .; Joudu, J .and Kuusik, A .(2001) .The effects of certain toxic plant extracts on the larvae of Colorado Potato beetle *Leptinotarsa decemlineata* (Say) . Institute of plant protection ,Estonian Agriculture University p .93-100.

Mubark, E.E.; Ali ,L.Z.; Ahamed ,I.F.A.; Ali AB.(2015). Int J Agr Biol. 17(2): 320-326.

Ravjit, K. K., M.J. Barbetti and M.W. Sweetingham. (1999). Characterization and pathogenicity of *Rhizoctonia* species on canola. Plant Disease. 83 (8): 714-727.

Sinclair, J. B. (1982). Compendium of soybean disease 2nd ed. American Phytopathological Soc. St. Paul. MN. 104pp.

Shayoub , M.H.; Dawoud, A.D.H; Abdelmageed ,M.A.M.; Ehasan, A. M. (2015). Phytochemical analysis of leaves extract of *Eucalyptus camaldulensis* Dehnh . 2(1).

Taylor, R.J.; Salas, B.; Secor, G.A.; Rivera, V. and Gudmestad, N.C. (2002). Sensitivity of north american isolates of *Phytophthora erythroseptica* and *Pythium ultimum* to mefenoxam (metalaxyl). Plant Disease. 86 : 797-802pp.

Parmeter, J. R. and Whitney, H. S. (1970). Taxonomy and nomenclature of the imperfect stage In: *Rhizoctonia solani* Biology and Pathology. (ed.) J. R. Parmeter. University of California Barkely. Los Angeles. 7–19 pp.

Watanabe, T. and Shiyomi, M . (1975) . Hyphal morphology of *Rhizoctonia solani* Kuhn and related fungi isolated from sugarcane in Taiwan. Trans . Mycol . Soci.Japan.16:253-263pp.

Weinhold, A. S., Sinsclair, J. (1996). *Rhizoctonia solani* : penetration, colonization and host response. P : 163 – 174 (C. F. Sneh, et al., 1996).

