

DOI: <https://dx.doi.org/10.21123/bsj.2023.7590>

Impact of some environmental parameters on phytoplankton diversity in the eastern Al-Hammer marsh / southern Iraq

Ebtehal M. Jaffer^{1*} 

Nida J. Al-Mousawi² 

Imad J. M Al-Shawi³ 

¹Department of Ecology, College of Science, University of Basrah. Basrah, Iraq.

²Department of Biology, College of Science, University of Basrah. Basrah, Iraq.

³Department of Applied Marine Sciences College of Marine Sciences Basrah, Iraq.

*Corresponding author: ebtehal.jaffer@uobasrah.edu.iq

E-mail addresses: Nida.mohammed@uobasrah.edu.iq, imad.mohammed@uobasrah.edu.iq

Received 24/6/2022, Revised 12/9/2022, Accepted 14/9/2022, Published Online First 20/3/2023



This work is licensed under a [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/).

Abstract:

Biodiversity is one of the important biological factors in determining water quality and maintaining the ecological balance. In this study, there are 223 species of phytoplankton were identified, and they are as follows: 88 species of Bacillariophyta and were at 44%, 70 species of Chlorophyta and they were at 29 %, 39 species of Cyanophyta and they were at 16 %, 12 species of Euglenozoa and they were at 4 %, four species of Miozoa and they were at 3 %, and, Phylum Charophyta and Ochrophyta were only eight and two species, respectively and both of them were at 2%. The common phytoplankton recorded in the sites studied include *Nitzschia palea*, *Scenedesmus quadricauda*, *Oscillatoria princeps*, and *Peridinium bipes*. These species recorded a significant positive correlation with Ec, Sio₃, and WT. Phytoplankton including *Gomphosphaeria semen-Vitis*, *Dicloster acuatus*, *Tetrastrum heteracanthum*, and *Dictyocha fibula*, recorded a significant positive correlation with NO₃, PO₄, DO, and PH. Water temperature ranged between 14.200 -33.900 °c in Al-Mansoury and Al-Sada respectively. Electrical conductivity ranged between 2.790 - 11.900 ms/cm in Al-Sada and Al-Mansoury respectively. PH ranged between 7.750-8.600 in Al-Dawody and Al-Mansoury respectively. Dissolved Oxygen (DO) ranged between 5.950 -13.000 mg/l in Al-Dawody and Al-Mansoury respectively. WT recorded negative correlation with pH ($r = - 0.591$), NO₃⁻² ($r = - 0.463$) and DO ($r = - 0.603$). Nitrate ranged between 0.570-12.200 µg /l in Al- and Al-Sada respectively. Phosphate ranged between 0.003-0.154 µg/l, in Al-Dawody and Al-Mansoury respectively. Silicate ranged between 51.200-198.600 µg /l in Al-Baraka and Al-Dawody respectively. Shannon - Weiner index (H') ranged between 2.275-3.162 in Al-Dawody and Al-Mansoury respectively. Simpson index ranged between 0.856-0.950 in Al-Mansoury and Al-Sada respectively, while the Evenness index was 0.514-0.933 in Al-Dawody and Al-Baraka respectively. Shannon- Weiner index (H') recorded a significant positive correlation with the Simpson index .

Keywords: Al-Hammer marsh, Biodiversity index, Environmental parameters, Phytoplankton

Introduction:

Aquatic ecosystems are the main sources of the continuance of life for most organisms that live in the aquatic system¹. Phytoplankton is the primary producers and significant source of nutrients in an aquatic ecosystem as they involve in the biogeochemical cycles of several elements to supply heterotrophic organisms with organic material. They are the essential biological characteristic that regulates productivity and nutrient cycling within food webs and carbon usage, along with water quality in determining the water ecosystems' ecological state. Phytoplankton availability is 40%

of the world's primary production, and it is the foundation of the aquatic food chain^{2,3}. Due to minor changes in nutrients and their brief life cycle, Phytoplankton reacts swiftly to environmental changes^{4,5}. Changes in hydro-climatic, biological, and chemical factors of the aquatic ecosystem have affected the distribution and abundance of Phytoplankton communities which indicate the water quality⁶. Since Phytoplankton communities are generally more sensitive to pollution, therefore, they are the best biotic indicators of pollution in the aquatic habitat⁷.

The study of the environment of the marshes was stopped for a long time due to the drying of the marshes 1990-2003, but after the rehabilitation of the Al-Hammer marsh there are not enough studies of the phytoplankton diversity in the Eastern Al-Hammer marsh, however, there are many studies covered physical and chemical factors and primary production. This study came to diagnose phytoplankton and the accompanying qualitative changes after drying and flooding, as in this study new species that are not registered in the Iraqi environment were recorded⁸. Some of these studies include limnological studies of traits of Al-Hammer marsh after restoration⁹, which studied¹⁰ Al-Hammer marsh after the flood; studied¹¹ evaluating water quality of Al-Hammer marsh with the GIS technique; studied¹² using indicators based on satellite images to evaluate the restoration plan for Al-Hammer marsh. Studied¹³ Carlson's Trophic State Index. Following a significant rise in salinity over the 2018 summer, studied¹⁴ the Eastern Al-Hammer marsh's Water Quality Index (WQI) was used as an indication and studied¹⁵ of the primary productivity of phytoplankton in the southern marshes of Iraq. There are recent studies in the other marshes of Iraq, and some studies showed the phytoplankton diversity in the different marshes of Iraq and^{16, 17} studied phytoplankton diversity in Auda marsh and studied¹⁸ the effect of physical and chemical factors in the water of the Chabayish marsh.

This work aimed to study the phytoplankton diversity in the Eastern Al-Hammer marsh as well as aquatic environmental factors after it flooded in 2003, whereby the traits of its ecosystem have changed.

Materials and Methods:

Study area

In southern Iraq, there are a lot of marshes, permanent ones and seasonal ones, but the significant marshes are that three marshes act as Iraq's wetlands: Marsh of Al-Hammer, Hawizeh marsh, and the central marshes¹⁹. The Iraqi marshes have changed significantly during the past 40 years for many reasons, such as drying and constructing dams, etc., which resulted in changing several of its traits, flooded areas, environmental system, and biological diversity composition²⁰.

East Al-Hammer marsh Fig. 1 acts as part of the South Al-marshes. It stretches for more than 33 kilometers, with the maximum deep water extent of 1.00 – 6.00 meters, depending on the tide. During the conditions in the 1990s, the marsh was exposed to desiccating. The marsh flooded in 2003 and recovered its vegetation and wildlife to varying

degrees¹³. The study was continued from December 2019 to November 2020, Phytoplankton was collected from four different sites by net (Al-Sada, Al-Mansoury, Al-Dawody, and Al-Baraka), Al-Sada site (N° 30°37.24.2 E° 47°40.07.7) is a far 6 km away from the Karma Bridge. The depth is approximately 6 m, it contains animals, birds, and Crustaceans, as well as various fishing operations, fishing Fish, shrimp, and birds that were recently damaged due to the progression of the saline tongue. Al-Mansoury site (N°30°40.31.4 E°47°37.22.9) is a distance from the site. Al-Sada is about 2 km away, and it is less area than the Al-Baraka site, but it is deeper than that and characterized this area is elongated in shape and called channel marsh, which contains branches and fish farms on the edges. It is one of the large marshes that are called Openness marsh. Al-Dawody site (N° 30°41.30.5 E° 47°35.52.1) is a narrow water channel of less depth, the height of the muddy land on two sides, the presence of many aquatic plants and the presence several different birds on the site Al-Baraka site (N° 30°41.47.8' E° 47°32.59.6) is an open and shallow area. The depth does not exceed three meters; it reaches its depth of 0.5m on the Islands. It is a far 13 km from the first station and much aquatic life, fish, shrimp, and many types of birds, reptiles, and mammals are inhabited it.

Field and laboratory study

water temperature (°C), electrical conductivity (EC) and pH were measured monthly using TRI METER (PH/EC-983) and DO determine by using an Oxygen meter (WTW). Nitrate, phosphate and silica were calculated based on²¹. The phytoplankton samples for this study were collected using a net of the size of about 20µm, we throw the net into the water and pulled it at a proper speed for 15 minutes, then collect the contents in the polyethylene bottle is clean and fixation in the field using formalin solution with a concentration of % 4. For microscope research and species identification, all organic material in diatom and silico-flagellate frustules must be oxidized and removed so that the valve structure can be observed. The frustules were cleaned in this study by boiling them in hydrogen peroxide. The 10 mL diatom suspension was heated for 30-45 minutes in 20 mL 30 % H₂O₂ or until the suspension became transparent²² 10 ml. The diatom suspension was allowed to settle at room temperature before being washed four times with distilled water and filtered using Whatman No. 1 filter paper to eliminate all organic contaminants permanent slides were made using Naphrax®. Algae morphology was screened (400x magnification). The Haemocytometer technique was used to count the non-diatom algae²³. One drop of the sample was

placed on a glass slide, covered with a glass cover slip, and viewed under a microscope with magnification 40x. using a microscope digital

camera (SCMOS03000KPA) and cell dimension (diameter, length, and width) of investigated algae, and algae were diagnosed based on the results²⁴⁻²⁶.

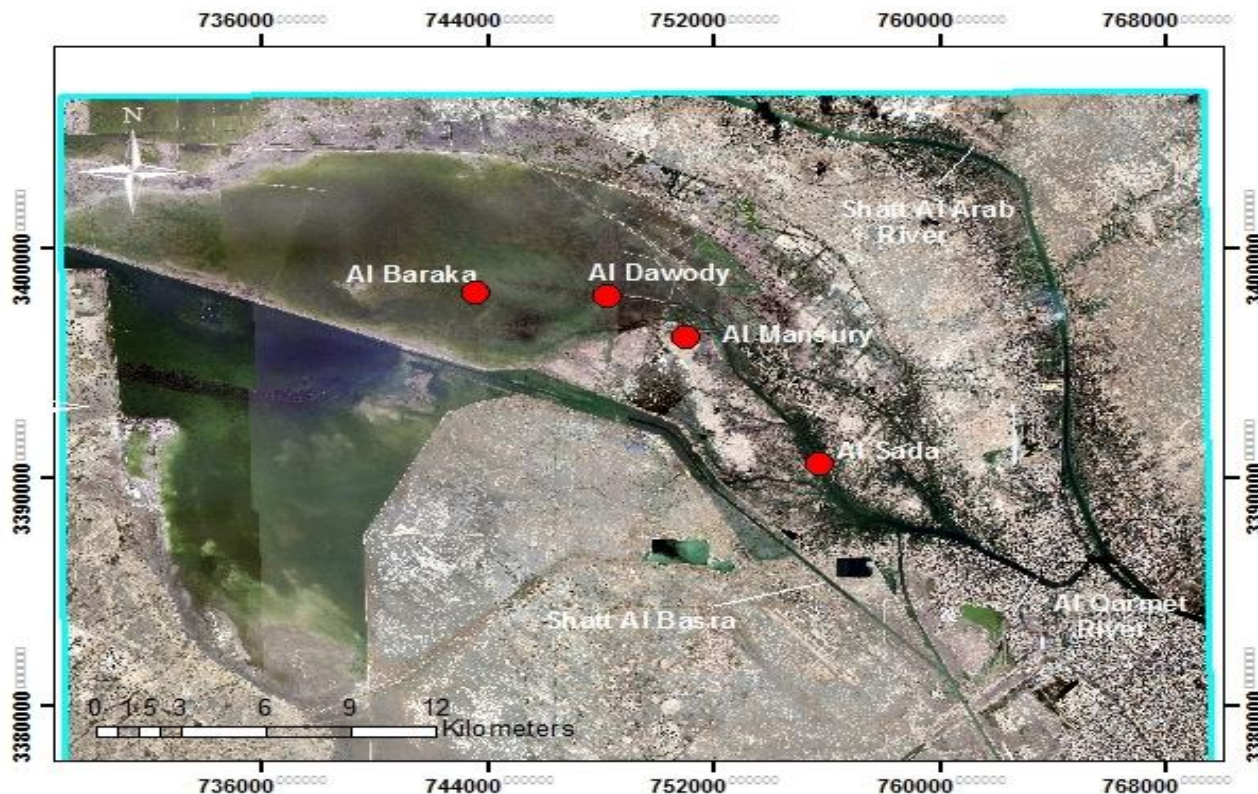


Figure 1. A map of sample collection sites from East Al-Hammer marsh, South of Iraq

Shannon - Weiner diversity index (H'):²⁷
index value was calculated by using the formula.*

$H' = -\sum p_i \ln p_i$ Where, 'pi' equal n_i/N

n_i : is the number of individuals in each species

N : the total number of individuals in the sample

Simpson's diversity index (D):²⁸

This index represents the abundance ratio of individual species to that of total* abundance values. It was calculated using the following formula $D = 1/\sum p_i^2$

p_i : is the preparation of the species to total abundance value.

Pielou's evenness index (J):^{29*}

$J = H'/\ln S$

Where:

S : total species H' : diversity index

Margalef Index (D):^{30*}

$D = S-1 / \ln N$

S : total species

N : the total number of individuals in the sample

Results and Discussion:

Values of physical-chemical parameters showed variations in the current study among four sites over ten months. WT ranged from 14.200 °c in Al-Mansoury during January and 33.900 °c in Al-Sada

during July, while ranging between 15.100-32.900°c in Al-Baraka and Al-Dawody respectively. WT recorded no Significant differences in all sites $p > 0.05$. some species appeared in the hot months such as *Nitzschia palea*, and *Oscillatoria princeps*, and some species appeared in the cold months such as *Gomphosphaeria semen-vitis*, *Dicloster acuatus* because WT is important for the physiology, behavior, distribution, survival, growth, and reproduction processes^{31,32}. The solubility of gases and salts in water is influenced by temperature, this is a confirmed negative correlation between WT and DO ($r = -0.603$) which is a significant component. Higher Electrical conductivity values of 11.900 ms/cm were observed in Al- Mansoury during May and a lower value of 2.790 ms/cm was observed in Al-Sada during November while ranging between 2.990 -11.400 ms/cm in Al-Baraka and Al-Dawody respectively due to decreased water discharges from Tigris and Euphrates and water entry from Arabian gulf across Shatt Al-Arab^{33,34}. EC recorded Significant differences in all sites $p < 0.05$, most diatom species recorded in the current study adapted to be found in saline and brackish water³⁵. EC showed significant negative correlation with NO_3 ($r = -0.328$) this agree with³³⁻³⁶ Table 1. Higher pH values of

8.600 were observed in Al-Mansoury during January and a lower value of 7.750 was observed in Al-Dawody during July while ranging between 7.840 - 8.530 in Al-Sada and Al-Baraka respectively. Increasing in pH may be the increase in the density of phytoplankton, the increase in the process of photosynthesis for algae and aquatic plants, and an increase in the consumption of carbon dioxide according to to^{6,37}. This confirmed a significant positive correlation between PH and DO ($r=0.630$). Higher Dissolved Oxygen values 13.000 mg/l were observed in Al-Mansoury during January and a lower value of 5.95 mg/l was observed in Al-Dawody during July while ranging between 7.540 -11.800 mg/l in Al-Sada and Al-Baraka respectively, the low values for (DO) in the hot months return to high temperatures and salinity³⁸ as well as an increase in activity organisms (bacteria) that decompose organic matter and consume oxygen, higher values for (DO) may be to increased photosynthesis rates by phytoplankton and aquatic plants, as well as a larger surface water area that allows for better mixing and oxygen compensation from the atmosphere³⁸, the current study agreed with the studies^{36,39}.

The maximum reactive Nitrate was 14.730 $\mu\text{g/l}$ in Al-Sada in December, and the slightest in Al-Dawody 0.570 $\mu\text{g/l}$ in August while ranging between 0.800 -11.740 $\mu\text{g/l}$ in Al-Baraka and Al-Mansoury respectively. NO₃ recorded Significant differences in all sites $p < 0.05$. The maximum reactive phosphate was 0.154 $\mu\text{g/l}$ in Al-Dawody in January slightest in Al-Mansoury 0.003 $\mu\text{g/l}$ in May while ranging between 0.004 -0.141 $\mu\text{g/l}$ in Al-Sada and Al-Baraka respectively, whereas the maximum reactive silicate 198.60 $\mu\text{g/l}$ in Al-Baraka in July and lowest in Al-Dawody 51.200 $\mu\text{g/l}$ in February while ranged between 61.6 -174.6 $\mu\text{g/l}$ in Al-Mansoury and Al-Sada respectively, Sio₃ showed a significant positive correlation with Wt ($r=0.630$). Po₄ and sio₃ recorded no Significant differences in

all sites $p > 0.05$ these results agreed with³⁶. In addition to the release of silica from sediments or the decomposition of diatom cells⁴⁰, these nutrients are rapidly eaten by the aquatic plants growing in the marsh. Phosphate (PO₄) and nitrate NO₃ concentrations were low¹⁰. Phytoplankton production is influenced by nitrogen, phosphorus, and inorganic phosphate. They respond quickly to the environment, disagreeing due to delicate nutrient alterations and their short life cycle^{41,42}. The growth of phytoplankton lowers the levels of micronutrients in the surface layers, such as nitrates, phosphates, and silicates, and limits primary production. This, too, is largely influenced by local weather conditions and topography⁴³.

Phytoplankton identified approximately 223 taxa belonging to 93 genera, Bacillariophyta represented the highest percentage 44% belonging to 88 taxa, 41 genera, they respond quickly to changes in the environment⁴⁴ the common genera *Nitzschia* 13 species and was one of common genera found at all sites due to the large range of environmental tolerance⁴⁵ and was from the common species *Gyrosigma* and *Tryblionella* found with eight species for both, *Navicula* six species, *Halamphora*, *Luticola* and *Surirella* found with three species for both, *Amphora*, *Campylodiscus*, *Cocconeis*, *Cyclotella*, *Cymatopleura*, *Cymbella*, *Entomoneis*, *Gomphonema*, *Rhopalodia*, *Tabularia* found with two species for both and recorded only one species for *Achnanthes brevipes* var. *intermedia*, *Bacillaria paxillifera*, *Coscinodiscus gigas*, *Diploneis smithii*, *Epithemia turgida*, *Mastogloia braunii*, *Pleurosigma elongatum* in the most sites, Bacillariophyta are dominant over the rest of the other groups, whether they are floating or attached⁴⁶⁻⁴⁹ and most of species recorded in eastern Al-Hammar marshes were brackish water species agreement with^{45,50}.

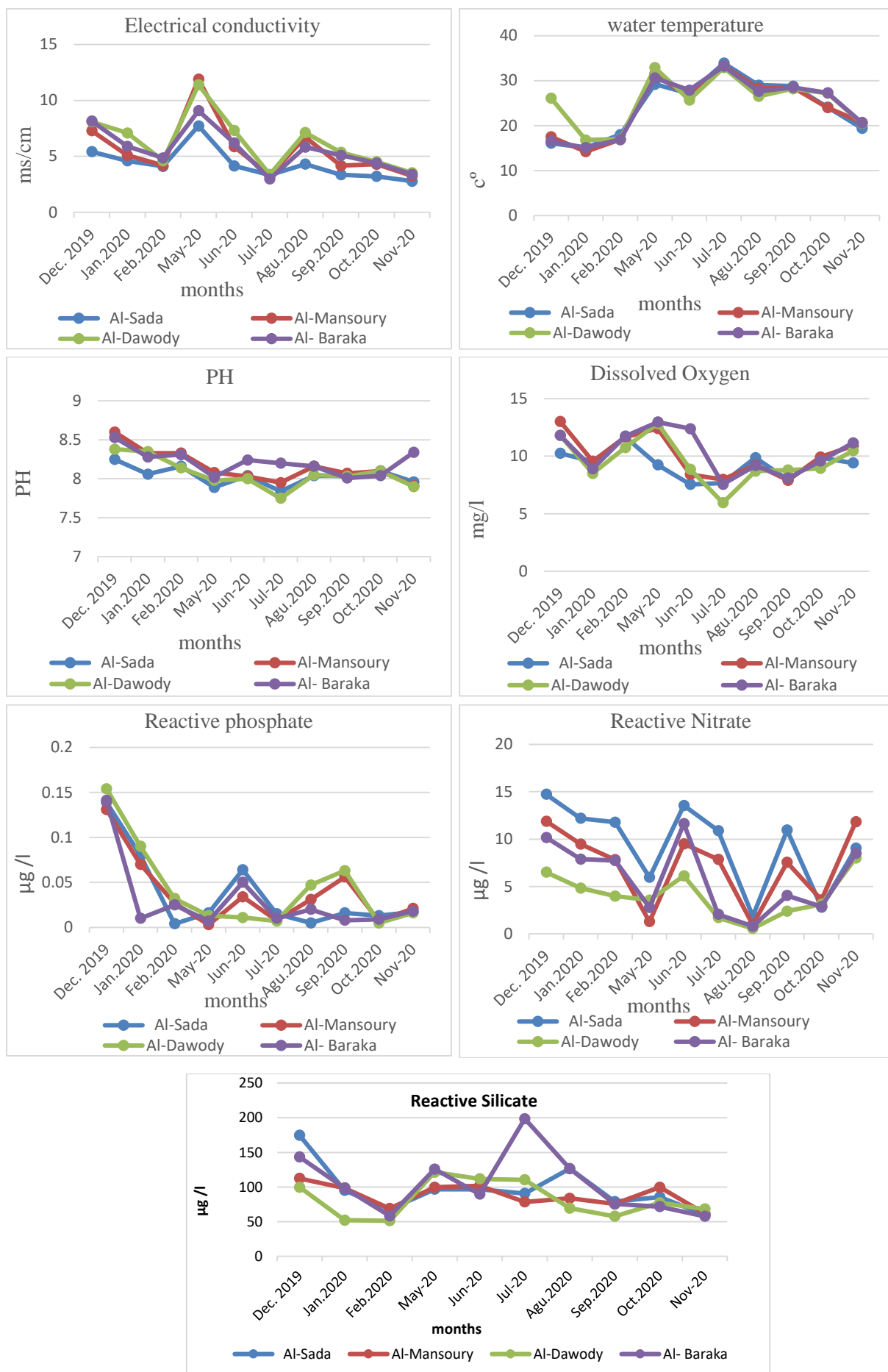


Figure 2. Physicochemical and *nutrients* parameters were variations in current study 2019-2020

The second group, Chlorophyta 29 %, belongs to 70 taxa, and 27 genera. The common genera are *Scenedesmus* 17 species; *Cosmarium* seven species, *Oocystis* and *Tetraedron*, found with six species for both, *Pediastrum* five species *Crucigenia* and *Ankistrodesmus*, found with four species for both, The other groups were Cyanophyta 16 % belonging to 39 species 15 genera, common genera that *Oscillatoria* eight species. *Chroococcus* six species, *Merismopedia* and *Gomphosphaeria* found with four species for both, *Anabaenopsis* and *Spirulina* found with two species for both and recorded only one species for *Komvophoron minutum*, *Pseudanabaena galeata*. Euglenozoa 4% belonging to 12 species three, common genera that *Euglena* eight species and was from the common species, *Phacus* three species and recorded only one species for *Lepocinclis glabr*, Miozoa(3%)belonging to four species three genera, common genera that *Peridinium* two species., recorded only one species for, *Glenodinium armatum*, *Noctiluca scintillans* Table. 2.

The correlation between the common species and environmental parameters at all sites was studied using (CCA) *N. palea* distributed evenly in sites of the current study and has a strong positive correlation between EC, silicate, and WT. On the other hand, this species shows a strong negative correlation with NO₃, PO₄, and pH because a plateful of Silicate makes diatoms take NO₃ and PO₄ at a much faster rate to build their cellular materials⁵¹ Fig 3. *S. quadricauda* has a strong positive correlation with NO₃ and EC because can be grown in a great range of brackish or saline water⁵². *O. princeps* has a strong positive correlation with PO₄ and PH, and a moderate positive correlation with NO₃ because these factors are responsible for the growth⁵³. On the other hand, this species includes a strong negative correlation with EC. *E. Proxima* recorded a significant positive correlation with PH and WT agreement with¹⁸. *P. bipes* distributed evenly in sites of the current study and showed a significant positive correlation with pH, NO₃, and PO₄ and showed a negative correlation with WT and EC. This indicates that the WT, pH, EC, nutrient nitrogen and phosphor play a pivotal role in the productivity and community structure of phytoplankton^{6,54}. phytoplankton recorded during intermittent periods include *Gomphosphaeria semen-vitis*, *Diclostera acuatus*, *Tetrastrum*

heteracanthum, and *Dictyocha fibula* have a strong positive correlation with NO₃ and PO₄ because these nutrients limiting factors for the growth of phytoplankton. On the other hand, these species include a strong negative correlation with WT and EC, WT is important for the physiology, behavior, distribution, survival, growth, and reproduction processes^{31,32}.

The Shannon diversity indicator is widely used to determine diversity that considers both the number and evenness of species in a community. It is a number that ranges from 0 to 4. The Shannon index (H') for phytoplankton was found to be the highest in Al-Dawody in May at 3.162 and indicates (less pollution) and lowest in Al-Mansoury in October at 2.275 indicate (more pollution) according to⁵⁵ that 0–1 (high pollution) 1–2 (moderate pollution), 2–3 (small pollution) and of 3–4 for inceptive pollution while ranged between 2.354 -3.045 in Al-Baraka and Al-Sada respectively.

The Simpson index (D) value ranges between 0 and 1, Simpson diversity index increases as a species evenness and increases richness⁶ a strong positive significant correlation between the Simpson diversity index with the evenness index (r=.490) for phytoplankton was found to be the highest in Al-Dawody in May (0.950) and lowest in Al-Mansoury in October 0.856 while ranged between 0.873 -0.945 in Al-Baraka and Al-Sada respectively. Evenness Index was found to be the highest in Al-Baraka in May 0.933 and lowest in Al-Dawody in July (0.514) while ranging between 0.572 -0.877 in Al-Mansoury and Al-Sada respectively. The evenness index indicates all species have equal abundance in the sample of site⁵⁶. Higher diversity values show the organism's suitability for the environment, more complicated ecosystems, as well as a more stable community.

The marsh has suffered from drying. As a result, the characteristics of its ecosystem have changed, and all its organisms have been affected, including algae. Other researchers noticed the phytoplankton species composition reaction to re-flooding¹⁷. After 2003, uncontrolled releases of Tigris and Euphrates River waters partially recovered some old marsh areas in southern Iraq, but restoration in others is failing due to the excessive soil and water salt concentrations⁵⁷.

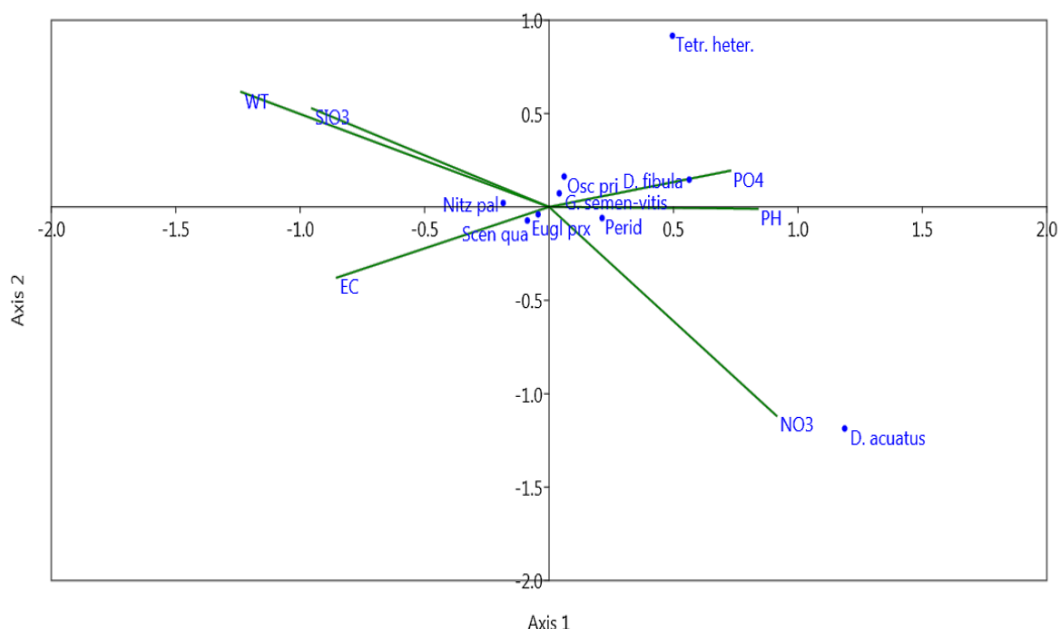


Figure 3. CCA- diagram of correlation between common species and phytoplankton recorded during intermittent periods with environmental parameters

Table 1. correlation between the environmental parameters and phytoplankton in the eastern AL- Hammar marsh, southern Iraq during 2019-2020

	Wt	Ec	PH	DO	NO3	PO4	SIO3	shan	Simp	even
Wt	1									
Ec	.130	1								
PH	-.591**	.225	1							
DO	-.603**	.231	.630**	1						
NO3	-.463**	-.328*	.113	.226	1					
PO4	-.103	.031	.247	.012	.164	1				
SIO3	.480**	.189	-.097	-.328*	-.328*	-.139	1			
shan	-.015	.111	.009	.184	-.017	-.069	-.138	1		
Simpson	.036	.108	.039	.175	.071	-.101	-.012	.934**	1	
even	.141	.127	-.017	.030	.157	-.115	.328*	.258	.490**	1

** . Correlation is significant at the 0.01 level (2-tailed).

* . Correlation is significant at the 0.05 level (2-tailed).

Table 2. Phytoplankton species were identified in the eastern Al-Hammar marsh, southern Iraq during 2019-2020

Phylum	Cyanobacteria(Cyanophyta)	3Order	Oscillatoriales
Class	Cyanophyceae	Family1	Gomontiellaceae
Order1	Chroococcales	Genus1	<i>Komvophoron</i> Anagn. and Kom. 1988 <i>K. minutum</i> (Skuja) Anagnostidis & Komárek
Family1	Chroococaceae	Family2	Oscillatoriaceae
Genus1	<i>Chroococcus</i> Naeg, 1849 <i>C. dispersus</i> (Keissler) Lemmermann <i>C. giganteus</i> West <i>C. limneticus</i> Lemmermann <i>C. minor</i> (Kützing) Nägeli <i>C. minutus</i> (Kützing) Nägeli <i>C. mipitanensis</i> (Wolszynska) Geitler <i>C. turgidus</i> (Kützing) Nägeli <i>C.sp.</i>	Genus1	<i>Lyngbya</i> C.Agardh ex Gomont, 1892 <i>L. aerugineocaerulea</i> Gomont
Family2	Cyanothrichaceae	Genus2	<i>Phormidium</i> Kützing ex Gomont, 1892 <i>P. ambiguum</i> Gomont
Genus1	<i>Johannesbaptistia</i> De Toni 1934 <i>J. pellucida</i> (Dickie) W.R.Taylor & Drouet	Genus3	<i>Oscillatoria</i> Vaucher, 1892 <i>O. agardhii</i> Gomont <i>O. brevis</i> Kützing ex Gomont <i>O. curviceps</i> C.Agardh ex Gomont <i>O. limnetica</i> Lemmermann <i>O. limosa</i> C.Agardh ex Gomont <i>O. princeps</i> Vaucher ex Gomont <i>O. splendida</i> Greville ex Gomont <i>O. tenuis</i> C.Agardh ex Gomont
Family3	Gomphosphaeriaceae	4Order	Spirulinales
Genus1	<i>Gomphosphaeria</i> Kützing, 1836 <i>G. aponina</i> Kützing <i>G. aponina</i> var. <i>cordiformis</i> Wolle <i>G. aponina</i> var. <i>delicatula</i> Virieux <i>G. semen-vitis</i> Komárek	Family1	Spirulinaceae
Family4	Microcystaceae	Genus1	<i>Spirulina</i> Tupin and. Gardner, 1827 <i>S. gigantea</i> Schmidle <i>S. major</i> Kützing ex Gomont
Genus1	<i>Gloeocapsa</i> Kuetzing 1843 <i>G. aeruginosa</i> Kützing	5Order	Synechococcales
Genus2	<i>Microcystis</i> Kuetzing, 1846 <i>M. aeruginosa</i> (Kützing) Kützing <i>M. aeruginosa</i> f. <i>flos-aquae</i> (Wittrock) Elenkin	Family1	Coelosphaeriaceae
2Order	Nostocales	Genus1	<i>Coelosphaerium</i> Naeg. 1849 <i>C. dubium</i> Grunow <i>C. kuetzingianum</i> Nägeli
Family1	Aphanizomenonaceae	Family2	Merismopediaceae
Genus1	<i>Anabaenopsis</i> V.V.Miller,1923 <i>A. circularis</i> (G.S.West) Wołoszyńska & V.V.Miller <i>A. elenkinii</i> V.V.Miller	Genus1	<i>Merismopedia</i> Meyen, 1839 <i>M. convoluta</i> Brébisson ex Kützing <i>M. elegans</i> A.Braun ex Kützing <i>M. glauca</i> (Ehrenberg) Kützing <i>M. punctata</i> Meyen, nom. illeg
Family2	Nostocaceae	Family3	Pseudanabaenaceae
Genus1	<i>Anabaena</i> Bory ex Bornet & Flahault, 1886 <i>A. flos-aquae</i> (Bornet & Flahault) Elenkin	Genus1	<i>Pseudanabaena</i> Lauterborn, 1915 <i>P. galeata</i> Böcher

Phylum	Chlorophyta	<i>C. astroideum</i> De Notaris
Class1	Chlorophyceae	<i>C.microporum</i> Nägeli
Order1	Chlamydomonadales	<i>C.reticulatum</i> (P.A.Dangeard) Senn
Family1	Chlamydomonadaceae	Genus2 <i>Scenedesmus</i> Meyen, 1829
Genus1	Carteria Diesing, 1866	<i>S. abundans</i> (O.Kirchner) Chodat
	<i>C. cordiformis</i> (H.J.Carter) Diesing	<i>S.acuminatus</i> (Lagerheim) Chodat
Genus2	Chlamydomonas Ehrenb, 1833	<i>S. acuminatus</i> var. <i>inermius</i> (Playfair) Playfair
	<i>C. angulosa</i> O.Dill	<i>S.acuminatus</i> var. <i>minor</i> G.M.Smith
	<i>C. globosa</i> J.W.Snow	<i>S.acutus</i> Meyen
Family2	Chlorococcaceae	<i>S.arcuatus</i> (Lemmermann) Lemmermann
Genus1	Chlorococcum Meneghin, 1842	<i>S.armatus</i> (Chodat) Chodat
	<i>C. humicola</i> (Nägeli) Rabenhorst	<i>S.bernardii</i> G.M.Smith
Family3	Goniaceae	<i>S.bijugus</i> (Turpin) Lagerheim
Genus1	<i>Gonium</i> O.F. Müller, 1773	<i>S.bijugatus</i> Kützing, nom. illeg.
	<i>G. pectorale</i> O.F.Müller	<i>S.brasiliensis</i> Bohlin
Family4	Volvocaceae	<i>S.dimorphus</i> (Turpin) Kützing
Genus1	<i>Eudorina</i> Ehrenberg, 1832	<i>S.ellipticus</i> Corda
	<i>E. elegans</i> Ehrenberg	<i>S. opoliensis</i> P.G.Richter
Genus2	<i>Pandorina</i> Bory, 1826	<i>S. obliquus</i> (Turpin) Kützing
	<i>P. morum</i> (O.F.Müller) Bory	<i>S.quadricauda</i> (Turpin) Brébisson
Genus3	<i>Pleodorina</i> W.R.Shaw, 1894	<i>S.quadricauda</i> var. <i>maximus</i> West & G.S.West
	<i>P.californica</i> W.R.Shaw	Genus3 <i>Tetrastrum</i> Chodat, 1895
Order2	Sphaeropleales	<i>T. heteracanthum</i> (Nordstedt) Chodat
Family1	Hydrodictyaceae	<i>T. staurogeniiforme</i> (Schröder) Lemmermann
Genus1	<i>Pediastrum</i> Meyen, 1829	Family5 Schroederiaceae
	<i>P.boryanum</i> (Turpin) Meneghini	Genus1 <i>Schroederia</i> Lemmermann, 1898
	<i>P. duplex</i> Meyen	<i>S. setigera</i> (Schröder) Lemmermann
	<i>P.duplex</i> var. <i>reticulatum</i> Lagerheim	Family6 Selenastraceae
	<i>P. simplex</i> Meyen	Genus1 Ankistrodesmus Corda, 1838, emRalfs, 1849
	<i>P. tetras</i> (Ehrenberg) Ralfs	<i>A. acicularis</i> (Braun) Korshikov
Genus2	<i>Tetraëdron</i> Kützing, 1845	<i>A. convolutus</i> Corda
	<i>Tetraëdron caudatum</i> (Corda) Hansgirg	<i>A.falcatus</i> (Corda) Ralfs
	<i>T. incus</i> (Teiling) G.M.Smith	<i>A.falcatus</i> var. <i>acicularis</i> (A.Braun) G.S.Wes
	<i>T.minimum</i> (A.Braun) Hansgirg	Genus2 <i>Kirchneriella</i> Schmidle 1893
	<i>T.muticum</i> (A.Braun) Hansgirg	<i>K. contorta</i> (Schmidle) Bohlin
	<i>T. regulare</i> Kützing	<i>K. obesa</i> (West) West & G.S.West
	<i>T.trigonum</i> (Nägeli) Hansgirg	Genus3 <i>Monoraphidium</i> Komárková-Legnerová, 1969
Family2	Neochloridaceae	<i>M. contortum</i> (Thuret) Komárková-Legnerová
Genus1	<i>Golenkinia</i> Chodat, 1894	
	<i>G. radiata</i> Chodat	
Family3	Radiococcaceae	
	<i>Gloeocystis</i> Nägeli, 1849	
	<i>G. gigas</i> (Kützing) Lagerheim	
Family4	Scenedesmaceae	
Genus1	<i>Coelastrum</i> Nägeli, 1849	

Class2	Trebouxiophyceae		<i>C. granatum</i> Brébisson ex Ralfs
Order1	Chlorellales		<i>C. hammeri</i> Reinsch
Family1	Chlorellaceae		<i>C. meneghinii</i> Brébisson ex Ralfs
Genus1	<i>Actinastrum</i> Lagerheim, 1882	Phylum	Euglenozoa
	<i>A. hantzschii</i> Lagerheim	Class1	Euglenophyceae
Genus2	<i>Chlorella</i> Beyerinck [Beijerinck], 1890	Order1	Euglenida
	<i>C. ellipsoidea</i> Gerneck	Family1	Euglenidae
	<i>C. vulgaris</i> Beijerinck	Genus1	<i>Euglena</i> Ehrenberg, 1830
Genus3	<i>Closteriopsis</i> Lemmermann, 1899		<i>E. acus</i> (O.F.Müller) Ehrenberg
	<i>C. longissima</i> (Lemmermann) Lemmermann		<i>E. convoluta</i> Korshikov
Genus4	<i>Dicloster</i> C.-C.Jao, Y.S.Wei & H.C.Hu, 1976		<i>E. elongata</i> W.Schewiakoff
	<i>D. acuatus</i> C.-C.Jao, Y.S.Wei & H.C.Hu		<i>E. gracilis</i> G.A.Klebs
Genus5	<i>Dictyosphaerium</i> Nägeli, 1849		<i>E. oxyuris</i> Schmarda
	<i>D. ehrenbergianum</i> Nägeli.		<i>E. polymorpha</i> P.A.Dangeard
	<i>D. pulchellum</i> H.C.Wood		<i>E. proxima</i> P.A.Dangeard
Family2	Oocystaceae		<i>E. spirogyra</i> Ehrenberg
Genus1	<i>Chodatella</i> Lemmermann, 1898	Family2	Phacaceae
	<i>C. ciliata</i> (Lagerheim) Lemmermann	Genus1	<i>Lepocinclis</i> Perty, 1849
Genus2	<i>Lagerheimia</i> R.Chodat, 1895, nom. Illeg		<i>L. glabra</i> Drezepolski
	<i>L. ciliata</i> (Lagerheim) Chodat	Genus2	<i>Phacus</i> Dujardin, 1841
Genus3	<i>Oocystis</i> Nägeli ex A.Braun, 1855		<i>P. acuminatus</i> A.Stokes
	<i>O. borgei</i> J.W.Snow		<i>P. caudatus</i> Hüner
	<i>O. crassa</i> Wittrock		<i>P. curvicauda</i> Svirenko
	<i>O. elliptica</i> West	Phylum	Miozoa(<i>Dinophyta</i>)
	<i>O. lacustris</i> Chodat	Class1	Dinophyceae
	<i>O. natans</i> G.M.Smith	Order1	Peridinales
	<i>O. solitaria</i> Wittrock	Family1	Peridinales incertae sedis
Order2	Trebouxiophyceae ordo incertae sedis	Genus1	<i>Glenodinium</i> Ehrenberg, 1836
Family1	Trebouxiophyceae		<i>G. armatum</i> Levander
Genus1	<i>Crucigenia</i> Morren, 1830	Family2	Peridiniaceae
	<i>C. crucifera</i> (Wolle) Kuntze	Genus1	<i>Peridinium</i> Ehrenberg, 1830
	<i>C. quadrata</i> Morren		<i>P. bipes</i> F.Stein
	<i>C. rectangularis</i> (Nägeli) Gay		<i>P. cinctum</i> (O.F.Müller) Ehrenberg
	<i>C. tetrapedia</i> (Kirchner) Kuntze	Class1	Noctilucophyceae
Phylum	Charophyta	Order1	Noctilucales
Class1	Zygnematophyceae	Family1	Noctilucaceae
Order1	Desmidiales	Genus1	<i>Noctiluca</i> Suriray, 1816
Family1	Closteriaceae		<i>N. scintillans</i> (Macartney) Kofoid & Swezy
Genus1	<i>Closterium</i> Nitzsch ex Ralfs, 1848	Phylum	Ochrophyta
	<i>C. moniliferum</i> Ehrenberg ex Ralfs	Class1	Chrysophyceae
Family1	Desmidiaceae	Order1	Chromulinales
Genus1	<i>Cosmarium</i> Corda ex Ralfs, 1848	Family1	Dinobryaceae
	<i>C. abbreviatum</i> Raciborski	Genus1	<i>Dinobryon</i> Ehrenberg, 1834
	<i>C. angulosum</i> Brébisson		<i>D. sertularia</i> Ehrenberg
	<i>C. botrytis</i> Meneghini ex Ralfs	Class2	Dictyochophyceae
	<i>C. contractum</i> O.Kirchner	Order1	Dictyochales
		Family1	Dictyochaceae

Genus1	<i>Dictyocha</i> Ehrenberg, 1837	Genus2	<i>Cymbella</i> C.Agardh, 1830
	<i>D. fibula</i> Ehrenberg		<i>C. cymbiformis</i> C.Agardh
Phylum	Bacillariophyta		<i>C.tumida</i> (Brébisson) van Heurck
Class1	Bacillariophyceae	Family2	Gomphonemataceae
Order1	Achnanthes	Genus1	<i>Gomphonema</i> Ehrenberg, 1832
Family1	Achnantheaceae		<i>G. intricatum</i> var <i>vibrio</i> (Ehrenberg) Cleve
Genus1	<i>Achnanthes</i> Bory, 1822		<i>G.parvulum</i> var. <i>lagenula</i> (Kützing) Frenguelli
	<i>A. brevipes</i> var. <i>intermedia</i> (Kützing) Cleve	Family3	Rhoicospheniaceae
Family2	Achnanthidiaceae	Genus1	<i>Rhoicosphenia</i> Grunow, 1860
Genus1	<i>Lemnicola</i> Round & Basson, 1997		<i>R. abbreviata</i> (C.Agardh) Lange-Bertalot 1980
	<i>L. hungarica</i> (Grunow Round & Basson	Order4	Licmophorales
Family3	Cocconeidaceae	Family1	Ulnariaceae
Genus1	<i>Cocconeis</i> Ehrenberg, 1836	Genus1	<i>Ctenophora</i> (Grunow) D.M.Williams & Round, 1986
	<i>C. placentula</i> Ehrenberg		<i>C. pulchella</i> (Ralfs ex Kützing) Williams & Round
	<i>C. placentula</i> var. <i>euglypta</i> (Ehrenberg) Grunow	Genus2	<i>Tabularia</i> (Kützing) D.M.Williams & Round, 1986
Order2	Bacillariales		<i>T. fasciculata</i> (C.Agardh) Williams & Round
Family1	Bacillariaceae		<i>T.tabulata</i> (C.Agardh)Snoeijs
Genus1	<i>Bacillaria</i> J.F.Gmelin, 1791	Genus3	<i>Ulnaria</i> (Kützing) Compère, 2001
	<i>Bacillaria</i> <i>paxillifera</i> (O.F.Müller) T.Marsson		<i>U.biceps</i> (Kützing) Compère
Genus2	<i>Nitzschia</i> Hassall, 1845	Order5	Mastogloiales
	<i>N. amphibian</i> Grunow	Family1	Mastogloiaceae
	<i>N. amplexens</i> Hustedt	Genus1	<i>Mastogloia</i> Thwaites ex W.Smith, 1856
	<i>N. clausii</i> Hantzsch		<i>M. braunii</i> Grunow
	<i>N. dissipata</i> (Kützing) Rabenhorst	Order6	Naviculales
	<i>N. fasciculata</i> (Grunow) Grunow	Family1	Naviculaceae
	<i>N. filiformis</i> (W.Smith)Hustedt	Genus1	<i>Caloneis</i> Cleve, 1894
	<i>N. hybrid</i> Grunow		<i>C. permagna</i> (Bailey) Cleve
	<i>N.kurzeana</i> Rabenhorst	Genus2	<i>Gyrosigma</i> Hassall, 1845
	<i>N. obtusa</i> W.Smith		<i>G. acuminatum</i> (Kützing) Rabenhorst
	<i>N. palea</i> (Kützing) W.Smith		<i>G. attenuatum</i> (Kützing) Rabenhorst
	<i>N. sigma</i> W.Smith		<i>G. balticum</i> (Ehrenberg) Rabenhorst
	<i>N. sigmoidea</i> (Nitzsch) W.Smith		<i>G. eximium</i> (Thwaites) Boyer
	<i>N.tryblionella</i> Hantzsch		<i>G. fasciola</i> (Ehrenberg) Griffith & Henfrey
Genus3	<i>Tryblionella</i> W.Smith, 1853		<i>G. macrum</i> (W.Smith) Griffith & Henfrey
	<i>T. apiculata</i> Gregory		<i>G. scalproides</i> (Rabenhorst) Cleve
	<i>T. cf. coarctata</i> (Grunow) Mann		<i>G. sinensis</i> (Ehrenberg) Desikachary
	<i>T. cocconeiformis</i> (Grunow) Mann	Genus3	<i>Haslea</i> Simonsen, 1974
	<i>T. compressa</i> (Bailey) Poulin		<i>H. spicula</i> (Hickie) Bukhtiyarova
	<i>T. granulata</i> (Grunow) Mann	Genus4	<i>Homoeocladia</i> C.Agardh, 1827
	<i>T. hungarica</i> (Grunow) Frenguelli		<i>H. subcohaerens</i> var. <i>scotica</i> Grunow
	<i>T.levidensis</i> W.Smith	Genus5	<i>Navicula</i> Bory, 1822
	<i>T. littoralis</i> (Grunow)Mann		<i>N. digitoradiata</i> (Gregory) Ralfs
Order3	Cymbellales		
Family1	Cymbellaceae		
Genus1	<i>Brebissonia</i> Grunow, 1860		
	<i>B. lanceolata</i> (C.Agardh) Mahoney & Reimer		

	<i>N. radiosa</i> Kützing		<i>C. cf. bicostatus</i> W.Smith ex Roper
	<i>N. rhynchocephala</i> Kützing		<i>C. sp.</i>
	<i>N. salinarum</i> Grunow	Genus2	<i>Cymatopleura</i> W.Smith, 1851
	<i>N. schroeterii</i> Meister		<i>C. elliptic</i> (Brébisson) W.Smith
	<i>N. subrhynchocephala</i> Hustedt		<i>C. solea</i> (Brébisson)W.Smith
Genus6	<i>Seminavis</i> D.G.Mann, 1990	Genus3	<i>Petrodictyon</i> D.G.Mann, 1990
	<i>S. strigosa</i> (Hustedt) Danieledis & Economou-Amilli		<i>P. gemma</i> (Ehrenberg) D.G.Mann
Family2	Amphipleuraceae	Genus4	<i>Surirella</i> Turpin, 1828
Genus1	<i>Vanheurckia</i> Brébisson, 1868		<i>S.robusta</i> Ehrenberg
	<i>V. lewisiana</i> (Greville) Brébisson		<i>S.striatula</i> Turpin
Family3	Berkeleyaceae		<i>S. tenera</i> W.Gregory
Genus1	<i>Parlibellus</i> E.J.Cox, 1988	Family2	Entomoneidaceae
	<i>P. crucicula</i> (W.Smith)Witkowski,Lange-Bertalot Metzeltin	Genus	<i>Entomoneis</i> Ehrenberg, 1845
Family4	Diadesmidaceae		<i>E. alata</i> (Ehrenberg) Ehrenberg
Genus	<i>Luticola</i> D.G.Mann, 1990		<i>E. paladosa</i> (W.Smith) Reimer
	<i>L. cf. mutica</i> (Kützing) Mann	Order8	Thalassioophysales
	<i>L. nivalis</i> (Ehrenberg) Mann	Family1	Catenulaceae
	<i>L. cf. ventricosa</i> (Kützing) Mann	Genus1	<i>Amphora</i> Ehrenberg ex Kützing, 1844
Family5	Diploneidaceae		<i>A. fluminensis</i> Grunow
Genus	<i>Diploneis</i> Ehrenberg ex Cleve, 1894		<i>A. ovalis</i> (Kützing) Kützing
	<i>D. smithii</i> (Brébisson)Cleve	Genus2	<i>Halamphora</i> (Cleve) Mereschkowsky, 1903
Family6	Naviculales_incertae_sedis		<i>H. ghanesis</i> Levkov
Genus	<i>Pseudofallacia</i> Y.Liu, J.P.Kociolek & Q.X.Wang, 2012		<i>H. paraveneta</i> (Lange-Bertalot, Cavacini, Tagliaventi & Alfinito) Levkov
	<i>P. tenera</i> (Hustedt) Liu, Kociolek & Wang		<i>H. veneta</i> (Kützing) Levkov
Genus	<i>Sieminskia</i> D.Metzeltin & Lange-Bertalot, 1998	Class2	Coscinodiscophyceae
	<i>S. wohlenbergii</i> (Brockmann) D.Metzeltin &Lange-Bertalot	Order1	Coscinodiscales
Family7	Stauroneidaceae	Family1	Coscinodiscaceae
Genus	<i>Craticula</i> Grunow, 1868	Genus1	<i>Coscinodiscus</i> Ehrenberg, 1839
	<i>C. cuspidate</i> (Kützing) Mann		<i>C. gigas</i> Ehrenberg
Family8	Pleurosigmataceae	Class3	Mediophyceae
Genus	<i>Pleurosigma</i> W.Smith, 1852,	Order1	Stephanodiscales
	<i>P. elongatum</i> W.Smith	Family1	Stephanodiscaceae
Order6	Rhopalodiales	Genus1	<i>Cyclotella</i> (Kützing) Brébisson, 1838,
Family1	Rhopalodiaceae		<i>C. meneghiniana</i> Kützing
Genus1	<i>Epithemia</i> Kützing, 1844		<i>C. straita</i> (Kützing) Grunow
	<i>E. turgida</i> (Ehrenberg) Kützing	Genus2	<i>Stephanodiscus</i> Ehrenberg, 1845
Genus2	<i>Rhopalodia</i> O.Müller, 1895, nom. cons.		<i>S. neoastreae</i> Håkansson & Hickel
	<i>R. gibba</i> (Ehrenberg) O.Müller	Order2	Thalassiosirales
	<i>R.musculus</i> (Kützing) O.Müller	Family1	Thalassiosiraceae
Order7	Surirellales	Genus1	<i>Thalassiosira</i> Cleve, 1873
Family1	Surirellaceae		<i>T. eccentric</i> (Ehrenberg) Cleve
Genus1	<i>Campylodiscus</i> Ehrenberg ex Kützing, 1844		

Conclusions:

The current study indicated the diversity of phytoplankton species in the eastern Al-Hammar marsh following various environmental changes. High salinity in May due to the lack of water drainage from the Tigris and Euphrates rivers and the entry of the waters of the Arabian Gulf through the Shatt al-Arab. Bacillariophyta has been identified as the dominant phylum of phytoplankton and *most species* recorded in eastern Al-Hammar marshes were brackish water *species*. Some species discovered in the current study point to the environment having altered and that saltwater organisms have evolved to survive in freshwater. The physicochemical parameters were varied in the current study among four different *sites*. Common species affected with *Ec*, *Sio3 WT* while *phytoplankton* recorded during intermittent periods affected with *PO4*, *NO3*, *DO*, and *PH*. The biodiversity indices differed among the stations of the East Al-Hammar marsh, except that the third station, Al-Dawody, was characterized by the highest diversity in the Shannon index and *indicated (less pollution)*.

Authors' declaration:

- Conflicts of Interest: None.
- We hereby confirm that all the Figures and Tables in the manuscript are mine ours. Besides, the Figures and images, which are not mine ours, have been given the permission for re-publication attached with the manuscript.
- Ethical Clearance: The project was approved by the local ethical committee in University of Basrah.

Authors' contributions statement:

EMJ analyzed the data, NJAM. and IJMAS suggested and developed the project. Both authors participated in writing the paper.

References:

1. Toma JJ, Aziz FH. Algal study in springs and streams from Shaqlawa district, Erbil Province, Iraq I-Euglenophyta Baghdad Sci J. 2022; 19(3): 483-492
2. Djumanto, Rasul E, Inoue T, Aoki S. Phytoplankton distribution in Mikawa Bay of Japan in relation to temperature and salinity variables. Wetlands 2017;1: 16-25.
3. Schaum C, Barton S, Bestion E. Adaptation of phytoplankton to a decade of experimental warming linked to increased photosynthesis. Nat Ecol Evol. 2017; 1:0094.
4. Taipale S J, Vuorio K, Aalto S L, Peltomaa E, Tirola M. Eutrophication reduces the nutritional value of phytoplankton in boreal lakes. Environ Res. 2019; 179: 108836.
5. Bergstrom AK, Jonsson A, Isles PDF, Creed IF, Lau CP. Changes in nutritional quality and nutrient limitation regimes of phytoplankton in response to declining N deposition in mountain lakes. Aquat Sci. 2020; 82(2): 31.
6. Inyang AI, Wang YS. Phytoplankton diversity and community responses to physicochemical variables in mangrove zones of Guangzhou Province, China, Springer. Ecotoxicology. 2020; 29(6): 650–668. <https://doi.org/10.1007/s10646-020-02209-0>.
7. Radwan A M, Abdelmoneim M A, Basiony A I, El-Alfy M A. Water Pollution Monitoring in Idku Lake (Egypt) using Phytoplankton and NSF-WQI. Egypt J Aquat Biol Fish. 2019; 23(4): 465 – 481
8. Jaffer E M, Al-Mousawi N J, Al-Shawi I J M. A Qualitative Study of Non-diatom Phytoplankton in East Al-Hammar Marsh. Egypt J Aquat Biol Fish, 2022; 26 (4): 449 – 468
9. Talib AH. Some limnological features of al-hammar marsh south of Iraq after restoration. Iraqi J Agric Sci. 2017; 48(3): 1331-1313.
10. Yaseen ST, Shaban AH, Jasim KA. Flood Behavior of Al-Hammar Marshes J Phys Conf Ser. 2021; 1879: 032062
11. AL-Musawi NO, AL-Obaidi SK, AL-Rubaie FM. Evaluating water quality index of Al-Hammar marsh, south of Iraq with the application of GIS technique. J Eng Sci Tech. 2018; 13 (12): 4118 – 4130
12. Dhaidan B A, Alwan I A, Al-Khafaji M S. Utilization of Satellite Images-Based Indices for Assessment of Al-Hammar Marsh Restoration plan. Eng Technol. 2021; 39 (08): 1328-1337.
13. Hussain N A, Sabbar A A. Trophic levels of Tidal and Non-Tidal Marshes of Southern Mesopotamia Basrah J Agric Sci. 2020; 33(2): 172-181.
14. Ghassan A Al-Nagar, Ali A Douabul, Sajed S Al-Noor. Water Quality Index (WQI) as indicator of the East Hammar marsh after sharpe salinity increase during summer 2018. Mar Bull. 2020 April; 15(1): 1–11
15. Maulood Bk, Hassan FM. Phytoplankton and Primary Production in Iraqi Marshes In : Jawad LA (ed.), Southern Iraq's Marshes, Coast Research Library, Springer Cham 2021; 36: 217-231.: https://doi.org/10.1007/978-3-030-66238-7_12
16. Albueajee AI, Hassan F M, Douabul AAZ. phytoplankton species composition and biodiversity indices in auda marsh- southern Iraq. Iraqi J Sci. 2020; 51(Special Issue): 217-228.
17. Al-Thahaibawi BMH, Al-Mayaly IKA, Al-Hiyaly SAK. Phytoplankton community within Al-Auda marsh in maysan province southern Iraq. IOP Conf Series: Environ Earth Sci. 2021; 722: 012026
18. Jaafar FA, Abdulwahhab AS. Impacts of the Physicochemical Properties of Al-Chibayish Water Marshes on The Biodiversity of Phytoplankton. Iraqi J Sci. 2021; 62(2): 402-414
19. Hussein ZE, Hasan RH, Aziz NA. Detecting the Changes of AL-Hawizeh Marshland and Surrounding Areas Using GIS and Remote Sensing Techniques Assoc. Arab Univ J Eng. Sci. 2018; 25: 53

20. Shimal S, Shaban AH .Estimation of groundwater pollution in Baiji / Salah Al-Deen province Iraq. AIP Conf Proc. 2019 July; 2123(1): 020058.
21. APHA (American Public Health Association). Standard methods for examination of water and wastewater. 21th ed, Washington D C. 2005; 1193p
22. Al-Handal AY, Wulff A. Marine epiphytic diatoms from the shallow sublittoral zone in Potter Cove, King George Island, Antarctica Bot Mar. 2008; 51: 411-435.
23. Martinez M R, Chakroff R P, pantastico JB. Note on direct phytoplankton counting technique using the haemocytometer. Phil Agric. 1975. 59: 1 – 12.
24. Desikachary TV. Cyanophyta, Indian Council of Agricultural Research, New Delhi. 1959; 686 p.
25. Prescott GW. Algae of the Western Great Lake Area, William, c. Brown co., publ. Dubuque, Iowa, 1982, 977p.
26. Guiry MD, Guiry GM. AlgaeBase. World-wide electronic publication. National University of Ireland, Galway. 2019; <http://www.algaebase.org>
27. Shannon CE, Wiener W. The mathematical theory of communication. Urbana university of Illinois Press, Chicago, USA; 1949. 117 p.
28. Simpson EH. Measurement of diversity. Nature. 1949; 163(688):688p. <http://dx.doi.org/10.1038/163688a0>
29. Pielou EC. Mathematical ecology. John Wiley & Sons, New York-London-Sydney-Toronto. Biom. J. 1978; 20(6): 627-628. <https://doi.org/10.1002/bimj.4710200616>
30. Margalef, R. perspectives in ecology. University of Chicago press. Chicago. *Limnol. Oceanogr.* 1969; 14(2): 179-316. <https://doi.org/10.4319/lo.1969.14.2.0313>
31. Ali H A, Owaid M N, Ali S F. Recording Thirteen New Species of Phytoplankton in Euphrates River Environment in Iraq. *Walailak J Sci Tech* 2020; 17(3): 200-211
32. Ashour M, Mabrouk M M, Ayoub H F, El-Feky M M, Zaki SZ, Hoseinifar S H, et al. Effect of dietary seaweed extract supplementation on growth, feed utilization, hematological indices, and non-specific immunity of Nile Tilapia, *Oreochromis niloticus* challenged with *Aeromonas hydrophila*. *J Appl Phycol* . 2020; 32(5): 3467-3479.
33. Alshaaban Z A A, Al-Hejuje M M. Comparison of the Application of two Trophic Status Indices at East Al - Hammar marsh - southern Iraq. *Mar Bull.* 2021 Sep; 16(2) : 161–172
34. Adlan NH, Al-Abbawy DA. Changes in physicochemical characteristics of water along shatt Al-Arab river. *Indian J Ecol.* 2022; 49 Special Issue (18): 300-307
35. Rasheed S S. Qualitative and quantitative study of the planktonic and epiphytic diatoms in east Hammar marsh /southern Iraq Master's Thesis, College of Science, University of Basra; 2019. https://www.researchgate.net/publication/353374544_Qualitative_and_quantitative_study_of_the_planktonic_and_epiphytic_diatoms_in_east_hammer_marsh_southern_Iraq?channel=doi&linkId=60f8dd2b1e95fe241a7d4709&showFulltext=true
36. Hussain NA, Abdalhsan H TH, Abduijaleel SA. Fish zonation patterns in East AL-Hammar tidal marsh/Basrah-Iraq, *Mar Bull* 2020 April; 15(1): 88-28.
37. Sivakumar K, Karuppasamy R. Factors affecting productivity of Phytoplankton in a Reservoir of 354 Tamilnadu , India *Amer Eur J Bot.* 2008; 1(3): 99-103
38. Koralay N, Kara O, Kezik U. Effects of run-of-the-river hydropower plants on the surface water quality in the Solakli stream watershed, Northeastern Turkey. *Water Environ J.* 2018; 32: 412-421
39. Hassan F M, Al-Kubaisi A A , Talib A H , Taylor W D , Abdulah D S. Phytoplankton primary production in southern Iraqi marshes after restoration, *Baghdad Sci J.* 2011; 8(1): 1111
40. Lind OT. Hand book of common methods in limnology. 2nd ed. London; 1979. 109 p.
41. Bergstrom AK, Jonsson A, Isles PDF, Creed IF, Lau DCP. Changes in nutritional quality and nutrient limitation regimes of phytoplankton in response to declining N deposition in mountain lakes. *Aquat Sci.* 2020; 82: 31
42. Taipale S J, Vuorio K, Aalto SL, Peltomaa E , Tiitola M . Eutrophication reduces the nutritional value of phytoplankton in boreal lakes. *Environ Res.* 2019; 179: 108836
43. Sourina A, Richard M. Phytoplankton and its contribution to primary production in two coral reef areas of French Polynesia. *J Exp Mar Biol Ecol.* 1976; 21: 129-140
44. Sanal M, Demir N. Use of the epiphytic diatoms to estimate the ecological status of Lake Mogan. *Appl Ecol Environ Res.* 2018; 16(3): 3529–3543
45. Al-Ahmady SSR, Al-Abbawy DAH, Al-Shaheen MAG . Relationships between environmental variables and both of planktonic and epiphytic diatoms in the East Hammar marshes, Southern Iraq *Mar Bull.* 2019 April; 14(1): 22-43
46. Al-Hussieny A A, Ali H A. List of Algae Species of Ramadi City within the Environment of the Euphrates River – Iraq. *Curr Res Microbiol Biotechnol.* 2017; 5(6): 1364-1374. https://www.researchgate.net/publication/323934961_List_of_Algae_Species_of_Ramadi_City_within_the_Environment_of_The_Euphrates_River_-_Iraq
47. Hassan F M, Salman J M and Al-Nasrawi, S. Community Structure of Benthic Algae in a Lotic Ecosystem, Karbala Province Iraq. *Baghdad Sci J.* 2017; 14(4): 2017.
48. Abdalhameed TA, Al Hassany JS. The Qualitative and Quantitative Composition of Epiphytic Algae on *Ceratophyllum demersum* L. in Tigris River within Wasit Province, Iraq *Baghdad Sci J.* 2019; 16(1): 1-9
49. ALbueajee AI, Hassan FM, Douabul AAZ. Epiphytic Algae Composition and Biodiversity in Auda Marsh / Southern of Iraq, *J Res Lepid.* 2020 June; 51 (2): 1135-1150.
50. Al-Shaheen MA, Al-Handal AY. Influence of environmental variables and different hosting substrate on diatom Assemblage in the Shatt Al-Arab River, Sothern Iraq. *Biol Appl Environ Res.* 2017; 1(1): 69-87

51. Al-Hassany J S, Alrubai G H, Jasim I M. The potential use of the diatom *Nitzschia palea* (Kützing) W. Smith For the Removal of Certain Pollutants from AlRustumeyah Wastewater Treatment Plant in Baghdad-Iraq IOP Conf. Series: Environ Earth Sci. 2021; 779: 012114
52. Ajlala SO, Alexander ML. Assessment of *Chlorella vulgaris*, *Scenedesmus obliquus*, and *Oocystis minuta* for removal of sulfate, nitrate, and phosphate in wastewater. Int J Energy Environ Eng. 2020; 11:311–326 2020
53. Bhat N A, Wangane A, Raina R. Variability in Water Quality and Phytoplankton Community during Dry and Wet Periods in the Tropical Wetland, Bhopal, India. J Ecosys Ecograph 2015; 5:2
54. Yang J, Li B, Zhang C, Luo H, Yang Z. pH-associated changes in induced colony formation and growth of *Scenedesmus obliquus* Fundam. Appl Limnol. 2016; 187(3):241–246
55. setyono P, Himawan W. Analyses of bioindicators and physicochemical parameters of water of Lake Tondano, North Sulawesi Province, Indonesia, Biodivers. 2018 May ;19 (3) : 867-874
56. Akhter S, Brraich OS. Spatial and Temporal Distribution of Phytoplankton from Ropar Wetland (Ramsar Site) Punjab, India. Appl Ecol Environ Sci, 2020; 8(1): 25-33
57. Richardson C J, Reiss P, Husain N A, Alwash A J and Pool D J. "The Restoration Potential of the Mesopotamian Marshes of Iraq. Science. 2005; 307:5713: 1307-1310

تأثير المتغيرات الفيزيائية والكيميائية على تنوع الهائمات النباتية في هور شرق الحمار/جنوب العراق

عماد جاسم الشاوي³

نداء جاسم الموسوي²

ابتهاال موسى جعفر¹

¹قسم البيئة، كلية العلوم، جامعة البصرة، البصرة، العراق

²قسم علوم الحياة، كلية العلوم، جامعة البصرة، البصرة، العراق

³قسم العلوم البحرية التطبيقية، كلية علوم البحار البصرة، العراق

الخلاصة :

ان التنوع الحيوي من العوامل البيولوجية المهمة في تحديد نوعية المياه والحفاظ على التوازن البيئي. لوحظ في الدراسة الحالية وجود 223 نوعا من الهائمات النباتية وكانت تنتمي الى 88 نوعا *Bacillariophyta* بنسبة 44% تليها *Chlorophyta* تنتمي الى 70 نوعا بنسبة 29% وفي المرتبة الثالثة *Cyanophyta* وتنتمي الى 39 نوعا بنسبة 16%، اما *Euglenozoa* تنتمي الى 12 نوعا بنسبة 4%، ثم تنتمي الى اربع أنواع *Miozoa* بنسبة 3%، اما الأقسام *Charophyta* و *Ochrophyta* فجاءت بالمرتبة الأخيرة والتي تنتمي الى ثمانية أنواع للافولوى ونوعان للثانية بنسبة مئوية 2%. من الأنواع الشائعة للهائمات النباتية التي سجلت في محطات الدراسة *Nitzschia palea*, *Scenedesmus quadricauda*, *Oscillatoria princeps* and *Peridinium bipes* و *Gomphosphaeria semen-vitis* *Dicloster acuatus*, *Tetrastrum heteracanthum* *Ec*, *Sio3 WT* و *Dictyocha fibula*، اظهرت علاقة ارتباط موجبة قوية مع الاوكسجين المذاب، الاس الهيدروجيني، الفوسفات الفعالة والنترات الفعالة تتراوح قيمة درجة حرارة الماء بين 14.200 - 33.900 °C في المنصوري والسدة على التوالي، تتراوح قيمة التوصيلية الكهربائية بين 2.790 - 11.900 ms/cm في السدة والمنصوري. تتراوح درجة الاس الهيدروجيني بين 7.750 - 8.600، ويتراوح الاوكسجين المذاب (DO) بين 5.950-13.000 mg/l سجلت درجة حرارة الماء علاقة ارتباط سالبة مع NO_3 ($r = -0.463$)، pH ($r = -0.591$) و DO ($r = -0.603$)، بلغت اعلى قيمة للنترات الفعالة 14.730 $\mu g/l$ بينما كانت ادنى قيمة 0.570 $\mu g/l$. بلغ الحد الأعلى للفوسفات الفعالة 0.154 $\mu g/l$ بينما كانت قيمة الحد الأدنى 0.003 $\mu g/l$ ، وبلغت اعلى قيمة للسليكات الفعالة 198.600 $\mu g/l$ بينما كانت ادنى قيمة 51.200 $\mu g/l$. سجلت NO_3 علاقة ارتباط سالبة مع $Sio3$ ($r = -0.328$) و EC ($r = -0.382$) بلغت اعلى قيمة لمؤشر شانون وواينر (H) 3.162 وكانت ادنى قيمة 2.275، بلغت اعلى قيمة لمؤشر سمبسون 0.950 وكانت ادنى قيمة 0.856، وبلغ الحد الأعلى لمؤشر التكافؤ 0.933 اما الحد الأدنى 0.514 وسجل دليل شانون وواينر علاقة ارتباط موجبة مع دليل سمبسون.

الكلمات المفتاحية: هور الحمار، ادلة التنوع الحيوي، المتغيرات البيئية والهائمات النباتية