

## **Intron 1 Inversion among Hemophilia a Patients in Basrah at the South of Iraq**

**Murtadha Ali Hadi<sup>1</sup>, Wijdan Nazar<sup>2</sup>, MeàadKadhun Hassan<sup>3</sup>**

<sup>1</sup>MSC. Department of Microbiology, College of Medicine, University of Basrah, Basrah, Iraq.  
mortadha.alaskare@gmail.com

<sup>2</sup>Ph.D. Department of Microbiology, College of Medicine, University of Basrah, Basrah, Iraq.

<sup>3</sup>Ph.D. Department of Pediatrics, College of Medicine, University of Basrah, Basrah, Iraq.

### **Abstract**

**Background/Aim:** Hemophilia A is an inherited bleeding disorder that results from a mutation in FVIII gene, intron 1 inversion is one of the mutations of FVIII gene. We aimed to detect the prevalence of Intron 1 inversion among hemophilia A patients in Basra/ south of Iraq

**Subjects and Methods:** A screen for the presence of intron 1 inversion at the F8 gene in 95 hemophilia A patient from Basrah at the south of Iraq. All patients have A total genomic DNA was extracted from EDTA fresh whole blood samples drawn from all participants, then detection of intron 1 inversion by conventional PCR and use of touchdown technique.

**Results:** Not there any one of Hemophilia A patients has Intron one Inversion

**Conclusions:** Absent of intron 1 inversion compare to other studies may be referred to geographic distribution of patients in spite of low prevalence percentage of intron 1 inversion, also more number of patients increase the accuracy of the research.

**Keywords:** intron 1 inversion, Basrah, Hemophilia, FVIII mutation, touchdown PCR.

### **Introduction**

The most common congenital bleeding disorder is hemophilia A, which is caused by a lack of the clotting protein Factor VIII (Callaghan, Sidonio and Pipe, 2018).

Factor VIII (FVIII) deficiency is an X-linked recessive condition that affects one out of every 5000 male births, regardless of ethnicity. (Witmer and Young, 2013).

F8 genotype mutations included large deletions (single or multiple exons), intron 1 and 22 inversions, nonsense mutations (light chain or non-light chain), small deletions/insertions/combined deletions and insertions (in poly-A runs or outside poly-A runs), splice site (conserved or non-conserved nucleotide positions), and missense mutations. (Gouw *et al.*, 2012).

Intron1 Inversion of factor VIII has been documented as one of the causes of severe hemophilia, and it appears to be responsible for roughly 2-5 percent of severe HA cases(Hart and Giangrande, no date; Hassan, 2017) and up to 7 % of HP in Asia (CHEN *et al.*, 2010).Intron 1 inversion mutations are caused by homologous recombination between a 1041-bp region (Int1h-1) and a 140-kb telomeric region (Int1h-2) that splices the FVIII gene promoter from exon 1.(Goodeve and Peake, 2003; Faridi, Kumar and Husain, 2012). This mutation occurs during male meiosis and results in the production of short mRNA, which results in the absence of FVIII.(Oldenburg, Pezeshkpoor and Pavlova, 2014).

Materia and method

### **Patients:**

The study included 95 male patients with Hemophilia A, ranging in age from 2 months to 63 years. All of the patients were diagnosed and registered at the Basra Center for Hereditary Blood Diseases, where they were assessed throughout their follow-up visits.

Patients with hemophilia A were split into three categories based on the severity of the disease: mild 37 (39%), moderate 33 (34.7 %), and severe 25 percent (26.3 % )

Regarding clinical presentation of hemophilia A patients in relation to the severity of the condition,96% of the sever cases presented with heamarthrosis, and 88% of them had Mucous membrane bleeding.while(69.7%) of moderate Hemophilia A patient presented with Muscle bleeding and (32.4%) of mild cases had bleeding after circumcission

### **Sample collection and there processing:**

Two milliliters of venous blood collected in an EDTA tube for Total genomicDNA extracted by wizard kit (Promega, USA). The extracted DNA was frozen in 30- C° until molecular analysis.

DNA loading and electrophoresis: 7 microliters of DNA were combined with 3 microliters of loading dye before being carefully placed in each gel well. The present power supply was switched on after that (70 volts for 30 minutes). From the cathode(-) to the anode(+) pole, the DNA band migrated. The Ethidium Bromide-stained bands inthe gel were visualized under UV transilluminator and photographed by adigital camera and the concentration of DNA was detected by Nanodrop. A Nano-dropSpectrophotometerwasusedtomeasuretheextractedDNAat260nm,whichwasthe wavelengthofmaximum absorption forDNA.

### **PCR Amplification**

To detect intron 1 inversion, a single PCR test run was employed to amplify a specific target using standard PCR and touchdown PCR techniques.The primers used in this study were

described in **Table 1** according to study of Wed elbahar with some modifications (Centre and Uk, 2016).

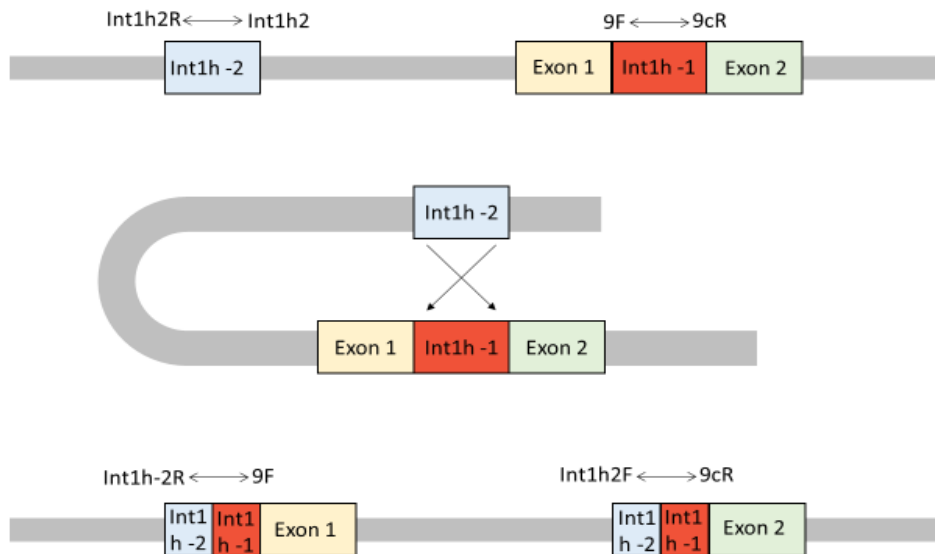
The PCR reaction tube contain 12.5 $\mu$  of master mix, 4 $\mu$  of primers mixture, 3 $\mu$  of DNA and 5.5 $\mu$  RNase-Free Water and use the condition of PCR reaction in the **Table 2**. In the reaction used two PCR tubes that just different in Primers mixture to detect of inversion in intron 1, that mean for each sample used 2 tubes in the reaction, first tube contain primers mix were (9F, 9cR, h-2F) produce region of 1908 bp for case without int1 inversion or 1776 bp for case with int1 inversion, while the second tube contain (h-2F, h-2R, 9F) primers mix that produce region of 1191 bp for case without int1 inversion or 1323 bp for case with int1 inversion according to **Figure 1**. The product of PCR diagnosed on agarose gel 2% after 1 hour at 70 Watt.

**Table 1: primers used to detect of FVIII gene Intron 1 inversion**

| Fragment | Primer name | Primer sequence                |
|----------|-------------|--------------------------------|
| Int1h-1  | 9F          | 5'-GTTGTTGGGAATGGTTACGG-3'     |
|          | 9cR         | 5'-CTAGCTTGAGCTCCCTGTGG-3'     |
|          | Int1h-2F    | 5'-GGCAGGGATCTTGTTGGTAAA-3'    |
| Int1h-2  | Int1h-2F    | 5'-GGCAGGGATCTTGTTGGTAAA-3'    |
|          | Int1h-2R    | 5'-TGGGTGATATAAGCTGCTGAGCTA-3' |
|          | 9F          | 5'-GTTGTTGGGAATGGTTACGG-3'     |

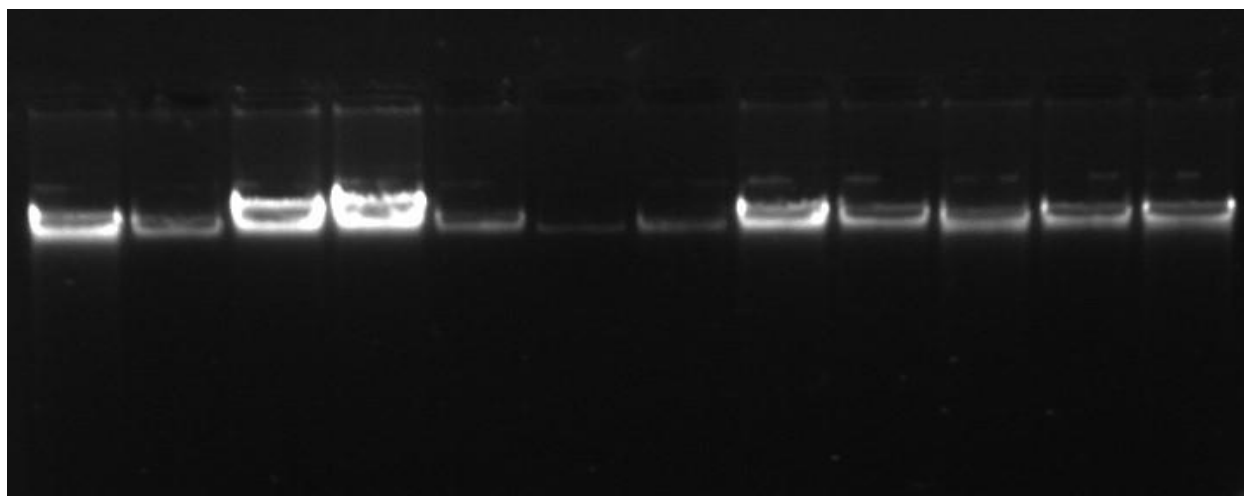
**Table 2 Procedure of conventional PCR reaction to detect of intron 1 inversion**

| Step                 | Temp (°C) | Time      | Cycle |
|----------------------|-----------|-----------|-------|
| Initial Denaturation | 94        | 5 minutes | 1     |
| Denature             | 94        | 30 second | 10    |
| Anneal               | 55-65     | 30 second |       |
| Extension            | 72        | 2 minutes |       |
| Denature             | 94        | 30 second | 25    |
| Anneal               | 55        | 30 second |       |
| Extension            | 72        | 2 minutes |       |
| Final Extension      | 72        | 5 minutes | 1     |



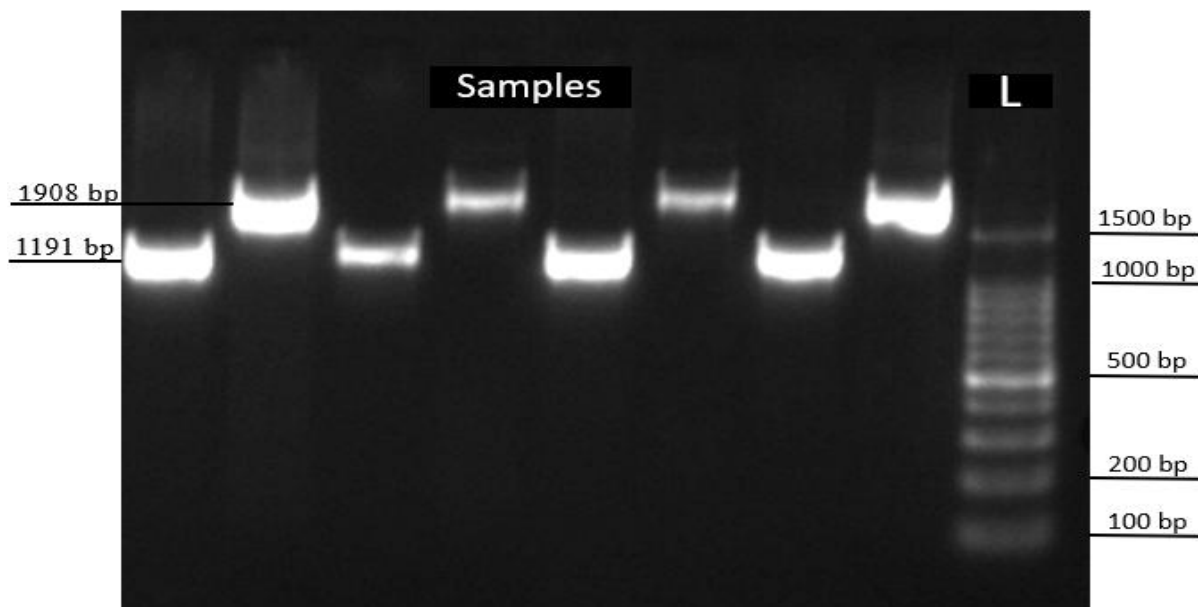
**Figure 1: inversion of intron 1 and primers position**

## Result



**Figure 2: (0.8%) of agarose was used for demonstration of genomic DNA**

There is no inversion in Intron I in any of the participants with hemophilia A (severe, moderate, or mild) as appear in picture 1 (9F,9cR and Int1h-2F) produce 1908 bp band while the second band appear 1191 bp with primers mixture (Int1h-2F, Int1h-2R and 9F). For that we not found direct relation between intron.



**Figure 3: PCR product on agarose gel for 75 minutes at 70 volts, all hemophilia patients without inversion, L: ladder 1500bp.**

## Discussion

Intron 1 inversion (Inv1) is a large molecular defect in FVIII gene resulting in severe HA. The pathogenic mechanism associated with this inversion involves homologous recombination between a 1041 bp region of intron 1 (int1h-1) of the FVIII gene and an extragenic copy (int1h-2) of region 5 approximately 140 kb telomeric to the FVIII gene.

In this study all the 95 with hemophilia A patients were analyzed for Intron 1 inversion and none of the severe cases nor the mild and moderate has this type of inversion. This result differs that found by Abdulqader *et al.*, who reported 3.3% (2/60) had Inv1 among Iraqi Kurdish patients with Hemophilia A (Abdulqader *et al.*, 2020), and Qadir *et al.*, in Kurdistan, Iraq also, found intron 1 inversion in only one out of 28 (3.5%) patients with hemophilia A (Qadir *et al.*, 2020).

The reported frequency of Inv 1 among patients with hemophilia A varies in different countries. In Pakistan, Inv. 1 was reported in 0.77% of severe HA patients (Sattar *et al.*, 2017). In India, Inv. 1 was found in 3.6% of PWH (Faridi, Kumar and Husain, 2012).

The results of the current study are similar to Castaman *et al.*, study in Albania (Castaman *et al.*, 2007) and Mantilla-Capacho *et al.* study in Mexico (Faridi, Kumar and Husain, 2012) who did not report Inv. 1 in their patients with hemophilia A. These variations in the results may be related to the sample selection, beside samples size and the geographical area of patients, also due to the immigration from the middle and north provinces to Basrah city in the last years

## Acknowledgments

The author thanks the employers in the center of hereditary blood disease specially for static part and laboratory.

## Reference

1. Abdulqader, A. M. R. *et al.* (2020) 'Identification of the Intron 22 and Intron 1 Inversions of the Factor VIII Gene in Iraqi Kurdish Patients With Hemophilia A', *Clinical and Applied Thrombosis/Hemostasis*, 26. doi: 10.1177/1076029619888293.
2. Callaghan, M. U., Sidonio, R. and Pipe, S. W. (2018) 'Novel therapeutics for hemophilia and other bleeding disorders', *Blood*, 132(1), pp. 23–30. doi: 10.1182/blood-2017-09-743385.
3. Castaman, G. *et al.* (2007) 'Spectrum of mutations in Albanian patients with haemophilia A: identification of ten novel mutations in the factor VIII gene', *Haemophilia*. Wiley Online Library, 13(3), pp. 311–316.
4. Centre, E. and Uk, D. (2016) 'European Journal of Biology and Medical Science Research', 4(6), pp. 1–6.
5. CHEN, Y. *et al.* (2010) 'Genetic analysis of haemophilia A in Taiwan', *Haemophilia*. Wiley Online Library, 16(3), pp. 538–544.
6. Faridi, N. J., Kumar, P. and Husain, N. (2012) 'Prevalence of intron 1 inversion of cases with hemophilia a in north Indian population', *Clinical and Applied Thrombosis/Hemostasis*, 18(6), pp. 599–603. doi: 10.1177/1076029611435094.
7. Goodeve, A. C. and Peake, I. R. (2003) 'The molecular basis of hemophilia A: Genotype - Phenotype relationships and inhibitor development', *Seminars in Thrombosis and Hemostasis*, 29(1), pp. 23–30. doi: 10.1055/s-2003-37936.
8. Gouw, S. C. *et al.* (2012) 'F8 gene mutation type and inhibitor development in patients with severe hemophilia A: systematic review and meta-analysis', *Blood, The Journal of the American Society of Hematology*. American Society of Hematology Washington, DC, 119(12), pp. 2922–2934.
9. Hart, D. P. and Giangrande, P. L. F. (no date) 'Chapter 17 The molecular basis of hemophilia Introduction : clinical features', pp. 221–234.
10. Hassan, M. M. (2017) 'Detection of Intron22 Mutations in Iraqi Female Carriers in Wasit province with Hemophilia A', 17(1).
11. M Kavitha, Z. H. Mahmoud, Kakarla Hari Kishore, AM Petrov, Aleksandr Lekomtsev, Pavel Iliushin, Angelina Olegovna Zekiy, Mohammad Salmani. application of Steinberg Model for Vibration Lifetime Evaluation of Sn-Ag-Cu-Based Solder Joints in Power Semiconductors. *IEEE Transactions on Components, Packaging and Manufacturing Technology*. 2021; 11(3):444-450.
12. Oldenburg, J., Pezeshkpoor, B. and Pavlova, A. (2014) 'Historical Review on Genetic Analysis in Hemophilia A', *Seminars in Thrombosis and Hemostasis*, 40(8), pp. 895–902. doi: 10.1055/s-0034-1395161.
13. Qadir, O. H. *et al.* (2020) 'Genotyping of intron 1 and 22 inversion of factor VIII gene using IS-PCR in Kurdish patients of Iraq', *EurAsian Journal of BioSciences*. Foundation for Environmental Protection and Research, 14(1), pp. 1181–1185.
14. Sattar, A. *et al.* (2017) 'Screening of intron 1 inversion of the factor VIII gene in 130 patients with severe hemophilia a from a Pakistani cohort', *Turkish Journal of*

- Hematology*, 34(3), pp. 270–281. doi: 10.4274/tjh.2017.0031.
15. Witmer, C. and Young, G. (2013) ‘Factor VIII inhibitors in hemophilia A: Rationale and latest evidence’, *Therapeutic Advances in Hematology*, 4(1), pp. 59–72. doi: 10.1177/2040620712464509.
  16. Z. H. Mahmoud. The Magnetic Properties of Alpha Phase for Iron Oxide NPs that Prepared from its Salt by Novel Photolysis Method. *Journal of Chemical and Pharmaceutical Research*, 2017, 9(8):29-33