

Research Article

Effect of stocking density and supplementation of potato peel (*Solanum tuberosum* L.) powder and extract on the physiological and immunological performance of Japanese quail, *Coturnix coturnix japonica*

Zainab A. MUTTER¹, Rabia J. ABBAS²

¹Department of Public Health, College of Veterinary Medicine, University of Basrah, Basrah, Iraq.

²Department of Animal Production, College of Agriculture, University of Basrah, Basrah, Iraq.

*Email: zainb.mutter@uobasrah.edu.iq

Abstract

This study aimed to investigate the effects of stocking density and supplementation of potato peel (*Solanum tuberosum* L.) powder and extract on Japanese quail's physiological and immunological performance. A total of 810 quail chicks were randomly assigned to six dietary treatments and two stocking densities in a complete randomized design with a 6×2 factorial scheme. The quail were kept with a stocking density of 333cm²/birds (15 birds/replicate), and 167cm²/birds (30 birds/replicate). The first treatment served as a control diet without supplementation (T1). T2 and T3 were supplemented with potato peel extract (PPE) at 15, and 30ml/l in drinking water; T4, T5 with 15, and 30g/kg potato peel powder (PPP) in basal diet, and T6 with PPE at 15ml/l+PPP at 15g/kg. A significant improvement in WBC (T2 and T6), total protein and globulin, and a significant decrease in relative weight of spleen in T3 and T6, bursa of fabricus and bursa index in all treatments, albumin in T2 and T3, glucose, and cholesterol were observed compared to control. Immune response, and value of antibodies titer against Newcastle and Gumboro disease were significantly better than control. In conclusion, the supplementation of PPE, PPP and their mixture showed the best physiological and immune response. Furthermore, the low density performed better in most studied traits.

Keywords: Quail, Potato peel powder and extract, Biochemical parameters, Immunity.

Citation: Mutter, Z.A.; & Abbas, R.J. 2022. Effect of stocking density and supplementation of potato peel (*Solanum tuberosum* L.) powder and extract on the physiological and immunological performance of Japanese Quail, *Coturnix coturnix japonica*. Iranian Journal of Ichthyology 9(Special Issue 1, 2022): 282-290.

Introduction

Stocking density, sensitivity to pathogens, metabolic complaints, breeding ability, and suitable slaughter age could affect overall economic performance in the poultry industry (Baeza et al. 2012; Berg & Yngvesson 2012; Kryeziu et al. 2018). Environmental stressors, including stocking density, humidity, and temperature, are crucial factors for the overall performance in this industry too (Gharaghani et al. 2015; Pompeu et al. 2018). Stocking density is

one of the most common stressors in the poultry industry. The advantages of stocking density include enhancing productivity, using restricted space, and increasing income (Majid et al. 2020).

The potato peel has received excessive attention as an herbal antioxidant and is documented to have antioxidant effects 10 times greater than its flesh with approximately 50% of all of the polyphenolic contents found in potato tuber (Javed et al. 2019). There is interest in herbal antioxidants as useful

components because they provide cell safety to oxidative damages restricting the danger of degenerative illnesses (Liang et al. 2014, 2015). There is a strong relationship between total phenol content in the extract and antioxidant properties; however, total polyphenol is decreased in potato peel during storage at -20°C . Therefore, correct storage conditions in potato peels to keep their antioxidant properties are crucial (Al-Weshahy & Rao 2009; Al-Weshahy et al. 2013). Alansari (2020) pointed out that adding rosemary oil to potato starch edible films improves the oxidative stability of foodstuff. Since using by-products in food with dietary benefits is economically attractive (Pathak 2015). Therefore, this study aimed to investigate the effect of stocking density and supplementation of potato peel powder and its extract on some hemato-biochemical parameters and immune response of Japanese quails.

Materials and Methods

Animals, diets, and management: The experiment was conducted in a quail farm, the Department of Animal Production, College of Agriculture, the University of Basra, from 28 December 2020 to 7 February 2021. A total number of 810 Japanese quail (*Coturnix coturnix japonica*) chicks were used in this study. Six dietary treatments and two stocking densities were used in a completely randomized design with a 6×2 factorial arrangement of treatments. The quail chicks were kept in cages of $100 \times 50 \text{cm}^2$ with a stocking density of $333 \text{cm}^2/\text{birds}$ (15 birds/replicate, $167 \text{cm}^2/\text{birds}$, and 30 birds/replicate). Chicks were fed six experimental diets from 7 to 42 days as a control diet without supplementation (T1). In the second and third treatments, the potato peel extract (PPE) was supplemented at 15 and 30ml/l in drinking water; in T4 and T5, 15, and 30g/kg potato peel powder (PPP) were added in the basal diet, and for treatment six, potato peel extract at 15ml/l+potato peel powder at 15g/kg were added, respectively. A basal diet was formulated according to NRC (1994) (Table 1). All

chicks were kept under constant management conditions throughout the experiment period of 42 days.

Preparation of potato peel powder (PPP): Potato peels (*Solanum tuberosum* L.) were collected from some restaurants in Basra City. The collected potato peels were washed, sundried, crushed into a meal, and stored for later use.

Chemical composition of potato peels powder: Proximate composition of the potato peel powder was analyzed based on AOAC (2016) (Table 2).

Preparations of potato peel extract (PPE): To prepare the methanol extract of potato peel powder, 10g of the ground materials were used to extract by 100ml of methanol alcohol overnight in a shaker at room temperature, followed by filtration through Whatman No.1 filter paper. The residues were re-extracted under the same conditions. The combined filtrates were evaporated in a rotary evaporator below 40°C . The potato peel extracts obtained after evaporation of methanol were stored at -20°C for further use (Mohdaly et al. 2010).

Hematological and Biochemical traits: On day 42, six birds (three males + three females) were selected randomly from each group and slaughtered to collect blood samples in tubes containing anticoagulant (EDTA). The hematological analysis includes red blood cell count (RBC), white blood cell count (WBC), packed cell volume (PCV), and hemoglobin (Hb). For biochemical traits, the blood was collected in tubes without anticoagulant, allowed for an hour at room temperature for serum collection, then serum separated by centrifuge at a speed of 3000rpm for 15min and then stored at -20°C for further analysis. Serum contents, including total protein, albumin, globulin, cholesterol, and glucose, were analyzed by a spectrophotometer using a commercial kite (Biolabo, SAS, France).

Immunological tests: Regarding immune response, six birds from each group were randomly chosen, weighed, slaughtered, eviscerated, and dressed. The relative weight for lymphoid organs, spleen and

Table 1. Ingredients and nutrient composition of quail basal diets.

Ingredients	Basal diet (%)
Maize	50.00
Wheat	8.75
Soybean meal (48%)	34.00
¹ protein concentrates (44%)	5.00
Dicalicum phosphate	0.50
Limestone	1.00
Mineral premix	0.30
Sodium chloride	0.30
DL-methionine	0.15
Total	100
Calculated composition²	
Metabolizable energy (Kcal /Kg)	2921
Crude protein	24.00
Crude fat	2.97
Crude fiber	2.54
Calcium (%)	0.84
Phosphorus available	0.36
Lysine	1.30
Methionine	0.48
Methionine + Cysteine	0.82

¹Protein concentrate used from Al-Hayat Company, Jordanian origin, to provide the following per kg of diet: 44% protein, 2800 kcal/kg ME, 12% fat, 25% ash, 5% calcium, 2.9% phosphorus, 2.55% methionine+Cysteine, 2.8% lysine. The chemical composition ²Was calculated according to of feedstuff contained in NRC (1994).

bursa of Fabricius were expressed as a percentage of live body weight. Blood samples were collected during slaughtering, allowed to clot, and then centrifuged immediately to separate serum and stored at -20°C until the determination of antibody titer against Newcastle (ND) and Gumboro diseases (IBD) by the hem-agglutination inhibition test using ELISA test kit.

Statistical analysis: An experiment was assigned in a complete randomized design with a 6×2 factorial scheme (3 replications each). Data were analyzed based on variance analysis at a 5% significance level. Duncan's multiple ranges tests were used if the treatment indicated significant effects (Duncan's 1955).

Results and Discussion

The effect of stocking density and experimental treatments on the hematological parameters of quail are shown in Table 3. The results indicate that RBC and PCV did not affect, whereas a significant ($P \leq 0.05$) improvement was observed in WBC of T2 and T6. Birds fed diets supplemented with 15 PPP

g/kg+15 PPE m/l (T6) showed significantly ($P \leq 0.05$) higher hemoglobin concentration and lower heterophils: lymphocytes (H/L) ratio. Regarding stocking density effects, the low density (D1) showed a higher ($P \leq 0.05$) WBC count and lower heterophils: lymphocytes (H/L) ratio in comparison with the high density (D2). This positive effect of these parameters may be attributed to the vital function of potato peels. Phenolic compounds of potatoes are phenolic acids and flavonoids, including flavonols and anthocyanins (Deusser et al. 2012). The relationship between polyphenol intake and reduced incidence of some diseases has been noticed in several studies (Akyol et al., 2016); however, their positive effects could not be given only to their antioxidant properties. The health benefits of polyphenols are assigned to some “non-antioxidant” complex activities that could not be linked to free radical inhibition (Andre et al. 2014). Blood haematological parameters serve as indicators of the physiological state of birds (Mohammed et al. 2015). The antioxidant potential of potato peel extracts was examined as a defensive towards erythrocytes oxidative induced *in vitro*

Table 2. Chemical analysis of potato peel powder.

Compounds	Percentage%
Dry matter	88.60
Moisture	11.40
Crud protein	12.23
Crud fiber	2.31
Total Ash	1.82
Crud fat	0.22
Nitrogen free extract	83.42
Organic mater	98.18
*Metabolic energy kcal/kg	3431.74

*ME was calculated by the equation described by (Pauzenga 1985).

damage, observing morphological and structural variations in the cell membrane (Javed et al. 2019). Medicinal plants or their extract improve quails' productive and physiological traits (Abbas et al. 2017; AL-Salhie & AL-Waeli 2019). Our results disagree with Kpanja et al. (2020), who observed no effect of the test material on hematological parameters of broiler chicks when using graded levels of potato peels meal in their diet. Also, our findings disagree with Beloor et al. (2010), who found that the H:L ratio is independent of stocking density at 15, 20, and 25 birds/m² in broiler chicks.

Table 4 indicates the effect of stocking density and experimental treatments on blood biochemical parameters of quail. The biochemical changes showed significant ($P \leq 0.05$) improvements in total protein and globulin concentrations compared to control, while albumin (in T2, T3, and T6), glucose, and cholesterol levels were significant ($P \leq 0.05$) decreased by the dietary treatments of broilers. The improvements in total protein and globulin in chicks fed potato peel powder (PPP) and its aqueous extract (PPE) may be due to potato peel antioxidant and antimicrobial properties, supporting birds' immune systems. In a previous study, potato peel extract was inspected for antimicrobial activity and found to be potentially effective against bacterial and fungal species. As compared to antibiotic (Streptocycline), potato peel extract effects significantly against *Clavibacter microgenesis* and *Pseudomonas*

aeruginosa (Prasad & Pushpa 2007). Gebrechristos et al. (2020) reported that potato peel extract has antibacterial and antioxidant properties, and incorporated with potato starch, produces an active film that can be used for active food packaging.

Low stocking density (D1) significantly affected serum total protein and globulin levels. The result showed an interaction effect of stocking density and supplementary diets on the content of serum total protein, globulin, and cholesterol. A significant effect on the interaction of PPP and PPE and stocking density may be due to the potato peels content of Phenolic compounds, which have antioxidant activity and other features that could encourage health (Deusse et al. 2012). Also, chlorogenic acid, which is extensively found in potato samples, is caused an increase in insulin sensitivity, decreases gut glucose absorption, and prevents gluconeogenesis (Ong et al. 2013; Andre et al. 2014). In contrast, Raphael (2017) reported that serum albumin level was highest in broiler fed 100% sun-dried Irish potato peel meal.

Inclusion of PPE and PPP and their mixture in the diet decreased ($P \leq 0.05$) total plasma cholesterol and glucose levels. The polyphenol and anthocyanin are rich in purple potato that play an important role in the safety against oxidative harm in rats fed a high-cholesterol diet (Ezekiel 2013). Althawab (2019) showed that the antioxidant ingredients in the peel of the purple sweet potato improved the lipid status in

Table 3. Effect of stocking density and experimental treatments on hematological parameters of quail at 42 days.

Parameters	Experimental diets							Mean	SEM	<i>P-value</i> *
	Density	T1(control)	T2 (15 m/l PPE)	T3 (30 m/l PPE)	T4 (15g/kg PPP)	T5 (30 g/kg PPP)	T6 (15 PPP + 15 PPE)			
RBC (10 ⁶ /mm ³)	D1	2.69	3.52	3.46	3.75	3.32	3.40	3.35	0.11	T=0.37
	D2	3.62	3.60	3.24	3.46	3.46	3.58	3.49	0.07	D= 0.29
	Mean	3.15	3.56	3.35	3.61	3.39	3.49	3.42	0.06	T×D=0.12
WBC (10 ³ /mm ³)	D1	29.26	34.90	31.84	32.10	32.10	31.93	32.02 ^A	0.61	T = 0.01
	D2	27.66	32.16	29.83	30.60	30.04	32.58	30.48 ^B	0.57	D=0.05
	Mean	28.46 ^b	33.53 ^a	30.83 ^b	31.35 ^b	31.07 ^{ab}	32.25 ^a	31.25	0.43	T×D=0.84
Hb (g/100ml)	D1	14.10	16.02	16.14	16.01	15.80	15.50	15.59	0.28	T = 0.18
	D2	14.95	15.76	14.98	14.99	15.91	16.76	15.56	0.25	D= 0.9
	Mean	14.53 ^c	15.89 ^b	15.56 ^b	15.50 ^b	15.85 ^b	16.13 ^a	15.58	0.18	T×D=0.32
PCV%	D1	36.66	49.66	46.66	43.00	46.66	45.66	44.72	1.66	T = 0.39
	D2	52.66	43.33	50.00	44.00	42.66	54.33	47.83	1.47	D=0.12
	Mean	44.66	46.50	48.33	43.50	44.66	50.00	46.27	1.12	T×D=0.06
H/L	D1	0.29	0.19	0.16	0.19	0.19	0.16	0.19 ^B	0.010	T=0.001
	D2	0.34	0.21	.210	0.23	0.25	0.18	0.23 ^A	0.012	D=0.001
	Mean	0.31 ^a	0.2 ^c	0.18 ^d	0.21 ^b	0.22 ^b	0.17 ^e	0.21	0.008	T×D=0.01

^{abc} Means in the same row with no common superscript are different at $P \leq 0.05$, ^{ab}Means in the same column with no common superscript are different at $P \leq 0.05$. SEM: Standard error of the mean. PPE: Potato peel extract; PPP: Potato peel powder, *P-value**, T= Treat effect, D: Density effect, T×D = Interaction effect, D1: 16 chicks/replicate; D2: 30chicks/replicate.

the blood of mice fed a high-fat diet after giving them peel extract with fermented cow's milk at a level of 1 and 2%. Nasoetion et al. (2019) reported that the addition of purple sweet potato extract at a level of 25ml/ kg led to a reduction in cholesterol and low-density lipoproteins in the serum of broiler chickens raised in different densities. Furthermore, the control of lipid oxidation associated with stocking density entails the supplementary of normal antioxidant factors to block the production of free radicals (Xiao et al. 2021). Our results are in line with Arun et al. (2015) that reported that a freeze-dried powder of potato peel causes a remarkable decrease in blood glucose levels and efficiently reduces the diabetic change in rats. In contrast to our study, Park et al. (2018) reported that a high stocking density increased total cholesterol and triacylglycerol levels in the blood of broilers. Also, Majid et al. (2020) observed a mild and humid climate with 20 chick density

treatments (high density) had the highest cholesterol levels, and Qaid et al. (2016) also reported that blood glucose increased due to stress caused by high stocking density.

The present results indicate a significant improvement in the relative weight of lymphoid organs (spleen and bursa of Fabricus), bursa index, antibody titers against Newcastle (ND), and Gumboro diseases (IBD) compared to control one (Table 5). A significant improvement was found in antibody titers against IBD and ND for low stocking density, while higher values were achieved at the higher density in the spleen. A significant interaction effect of stocking density and supplementary diets on serum antibody titers' content against IBD was also observed. It is widely known that high stocking density induces stress responses in broilers (Al-Benna et al. 2006). The antioxidants found in herbs and their extracts reduce stress on birds. Potato peel

Table 4. Effect of stocking density and experimental treatments on blood biochemical parameters of quail at 42 days.

Biochemical Parameters	Experimental diets							Mean	SE M	P-value*
	Density	T1 (control)	T2 (15 m/l PPE)	T3 (30 m/l PPE)	T4 (15g/kg PPP)	T5 (30 g/kg PPP)	T6 (15PPP + 15 PPE)			
Total Protein (g/dl)	D1	4.37	5.15	5.58	5.57	4.63	5.58	5.15 ^A	0.12	T=0.02
	D2	3.97	5.29	4.35	4.17	5.44	5.49	4.78 ^B	0.17	D= 0.01
	Mean	4.17 ^c	5.22 ^{ab}	4.96 ^b	4.87 ^b	5.03 ^b	5.53 ^a	4.96	0.11	T×D=0.001
Albumin (g/dl)	D1	2.49	1.84	1.89	2.68	2.16	2.12	2.20	0.10	T = 0.01
	D2	2.27	1.68	1.68	1.95	2.74	1.93	2.04	0.11	D=0.23
	Mean	2.38 ^a	1.76 ^c	1.79 ^c	2.31 ^a	2.45 ^a	2.02 ^b	2.12	0.07	T×D =0.14
Globulin (g/dl)	D1	1.88	3.30	3.68	2.89	2.46	3.46	2.95 ^A	0.16	T = 0.001
	D2	1.70	3.61	2.67	2.22	2.69	3.55	2.74 ^B	0.17	D=0.02
	Mean	1.79 ^c	3.46 ^a	3.17 ^a	2.55 ^b	2.58 ^b	3.51 ^a	2.84	0.11	T×D = 0.001
Cholesterol (mg/dl)	D1	153.49	128.96	115.79	118.90	119.06	122.80	126.50	3.25	T = 0.001
	D2	138.46	127.51	115.74	128.62	122.32	118.81	125.24	2.00	D=0.43
	Mean	145.97 ^a	128.23 ^b	115.77 ^d	123.76 ^b	120.69 ^{cd}	120.81 ^{cd}	125.87	1.88	T×D 0.001
Glucose (mg/dl)	D1	179.20	150.83	137.91	154.58	158.52	159.34	156.73	4.73	T = 0.001
	D2	178.40	148.28	138.78	177.51	167.31	166.57	162.81	4.05	D=0.23
	Mean	178.80 ^a	149.55 ^{bc}	138.35 ^c	166.06 ^b	162.92 ^b	162.96 ^b	159.77	3.11	T×D =0.69

abc: Means in the same row with no common superscript are different at $P \leq 0.05$, AB: Means in the same column with no common superscript are different at $P \leq 0.05$. SEM: Standard error of the mean, PPE: Potato peel extract; PPP: Potato peel powder, P-value*, T= Treat effect, D: Density effect, T×D = Interaction effect, D1: 15 chicks/replicate; D2: 30chicks/replicate.

Table 5. Effect of stocking density and experimental treatments on immunological parameters of quails at 42 days.

Immunological Parameters	Experimental diets							Mean	SEM	P-value*
	Density	T1(control)	T2 (15 m/l PPE)	T3 (30 m/l PPE)	T4 (15g/kg PPP)	T5 (30 g/kg PPP)	T6 (15PPP + 15 PPE)			
Spleen%	D1	0.08	0.05	0.03	0.07	0.0	0.02	0.05 ^A	0.010	T=0.001
	D2	0.09	0.05	0.05	0.08	0.05	0.04	0.06 ^A	0.012	D= 0.001
	Mean	0.08 ^a	0.05 ^c	0.04 ^d	0.07 ^b	0.05 ^c	0.03 ^c	0.05	0.008	T×D= 0.17
Bursa of fabric us %	D1	0.04	0.03	0.03	0.02	0.02	0.02	0.03	0.001	T = 0.001
	D2	0.04	0.02	0.02	0.03	0.02	0.02	0.02	0.002	D=0.07
	Mean	0.04 ^a	0.03 ^b	0.03 ^b	0.03 ^b	0.02 ^b	0.02 ^b	0.03	0.002	T × D =0.05
Bursa Index	D1	1.00	0.82	0.85	0.74	0.68	0.56	0.77	0.04	T = 0.001
	D2	1.00	0.64	0.65	0.74	0.6	0.72	0.72	0.03	D= 0.11
	Mean	1.00 ^a	0.73 ^a	0.75 ^b	0.74 ^b	0.64 ^b	0.64 ^b	0.75	0.03	T × D = 0.03
Antibody titers against ND	D1	3675.67	6468.33	5706.33	4486.67	4589.33	6550.33	5246.11 ^A	264.19	T = 0.001
	D2	3418.00	5751.67	5249.33	4178.00	4275.66	6505.33	4896.33 ^B	258.37	D=0.001
	Mean	3546.83 ^c	6110.00 ^b	5477.83 ^c	4332.33 ^d	4432.5 ^d	6527.83 ^a	5071.22	184.38	T × D = 0.33
Antibody titers against IBD	D1	2113.33	3328.00	3962.67	2551.67	3043.33	3843.00	3140.33 ^A	163.81	T = 0.001
	D2	2056.00	3271.33	2976.00	2309.00	2635.67	3245.33	2748.89 ^B	125.05	D=0.001
	Mean	2084.67 ^d	3299.67 ^a	3469.33 ^a	2430.33 ^c	2839.50 ^b	3544.17 ^a	2944.61	106.75	T × D = 0.01

abc: Means in the same row with no common superscript are different at $P \leq 0.05$, AB: Means in the same column with no common superscript are different at $P \leq 0.05$. SEM: Standard error of the mean, PPE: Potato peel extract; PPP: Potato peel powder, P-value*, T= Treat effect, D: Density effect, T×D = Interaction effect, D1: 15 chicks/replicate; D2: 30 chicks/replicate, ND: Newcastle disease, IBD: Gumboro disease.

is rich in bioactive compounds, including antioxidants; the most important characteristic of peel is due to containing phenolic acids and flavonoids, including flavonols, and anthocyanins (Deusser et al. 2012). Potato peels seem to constitute a potential source of natural antioxidants used in the food industry (Carocho et al. 2018). The antioxidant effect of the extract resulted in low levels of peroxide value and carbonyl compounds (Sabeena et al. 2012). In this regard, Wang et al. (2016) revealed the antioxidant mechanism of immunomodulatory activity of sweet potatoes. Wang et al. (2016) found that water extract of sweet potato has the highest immunomodulatory activity by stimulated T- and B-cell proliferation and pro-inflammatory cytokine production, water extract restricted the production imbalance of the Th1-type, and Th2-type cytokine (Kim et al. 2015). Regarding density effect, Majid et al. (2020) noted that the density of 20 chicks/m² (high density) at 42 days of age had the lowest immune response, whereas, Houshmand et al. (2012) also showed the relative weight of the bursa and spleen were not affected by stocking density.

Conclusion

It is concluded that supplementation of PPE and PPP and their mixture achieved the best physiological and immune response in quails, with an interaction effect of stocking density and supplementary diets on the in heterophils: lymphocytes (H/L) ratio, Hb%, the content of total protein, globulin, cholesterol and antibody titers against IBD of quails.

Acknowledgments

The authors gratefully thank the staff of Quail farm for their assistance in carrying out this experiment.

References

A.O.A.C. 2016. Official Methods of Analysis of AOAC International George W., & Latimer, Jr. (Eds.), 20th ed Rockville, Maryland 20850-3250, 3172 p.

Abbas, R.J.; Al-Salhi, K.C.K. & Al-Hummod, S.K.M. 2017. The effect of using different levels of pomegranate (*Punica granatum*) peel powder on

productive and physiological performance of Japanese quail (*Coturnix coturnix japonica*). Livestock Research for Rural Development 29.

Alansari, B.M. 2020. Development of Antioxidant Activity of Potato Starch Edible Films Incorporate with Rosemary *Rosmarinus officinalis* L. Oil and Used it in Packaging Beef Pattis. Basrah Journal of Agricultural Sciences 33(2): 67-79.

Al-Benna, S.; Hamilton, C.A.; McClure, J.D.; Rogers, P.N.; Berg, G.A.; Ford, I.; Delles, C. & Dominiczak, A.F. 2006. Low-density lipoprotein cholesterol determines oxidative stress and endothelial dysfunction in saphenous veins from patients with coronary artery disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 26(1): 218-223.

Al-Salhi, K.C.K. & Al-Waeli, A.M. 2019. The effect of using different levels of red ginseng roots powder on some physiological characteristics of Japanese quail males (*Coturnix japonica*). *Basrah Journal of Agricultural Sciences* 32(1): 34-38.

Althawab, S.A.; Mousa, H.M.; El-Zahar, K.M. & Zaher, A.M.A. 2019. Protective effect of sweet potato peel against oxidative stress in hyperlipidemic Albino rats. *Food and Nutrition Sciences* 10: 503-516.

Al-Weshahy, A. & Rao, V.A. 2009. Isolation and characterization of functional components from peel samples of six potatoes varieties growing in Ontario. *Food Research International* 42: 1062-1066.

Al-Weshahy, A.; El-Nokety, M.; Bakhete, M. & Rao, V.A. 2013. Effect of storage on antioxidant activity of freeze-dried potato peels. *Food Research International* 50: 507-512.

Andre, C.; Legay, S.; Iammarino, C.; Ziebel, J.; Guignard, C.; Larondelle, Y.; Hausman, J.; Evers, D. & Miranda, L. 2014. The potato in the human diet: A complex matrix with potential health benefits. *Potato Research* 57: 201-214.

Arun, K.; Chandran, J.; Dhanya, R.; Krishna, P.; Jayamurthy, P. & Nisha, P.A. 2015. Comparative evaluation of antioxidant and antidiabetic potential of peel from young and matured potato. *Food Biosciences* 9: 36-46.

Baéza, E.; Arnould, C.; Jlali, M.; Chartrin, P.; Gigaud, V.; Mercierand, F.; Durand, C.; Méteau, K.; Le Bihan-Duval, E. & Berri, C. 2012. Influence of increasing slaughter age of chickens on meat quality, welfare,

- and technical and economic results. *Journal of Animal Science* 90(6): 2003-2013,
- Beloor, J.; Kang, H.K.; Kim, Y.J.; Subramani, V.K.; Jang, I.S., Sohn, S.H. & Moon, Y.S. 2010. The effect of stocking density on stress related genes and telomeric Length in broiler chickens. *Asian-Australasian Journal Animal Science* 23: 437-443.
- Berg, C. & Yngvesson, J. 2012. Optimal stocking density for broilers-optimal for whom? *World's Poultry Congress*. pp. 5-9. Salvador, Bahia, Brazil.
- Carocho, M.; Morales, P. & Ferreira, I.C.F.R. 2018. Antioxidants: reviewing the chemistry, food applications, legislation and role as preservatives. *Trends in Food Science and Technology* 71: 107-120.
- Deusser, H.; Guignard, C.; Hoffmann, L. & Evers, D. 2012. Polyphenol and glycoalkaloid contents in potato cultivars grown in Luxembourg. *Food Chemistry* 135(4): 2814-2824.
- Duncan, D.B. 1955. Multiple range test and F-test. *Biometrics* 11: 1-42.
- Ezekiel, R.; Singh, N.; Sharma, S. & Kaur, A. 2013. Beneficial phytochemicals in potato-A review. *Food Research International* 50: 487-496.
- Gebrechristos, H.Y., Ma, X.; Xiao, F.; He, Y.; Zheng, S.; Oyungere, G. & Chen, W. 2020. Potato peel extracts as an antimicrobial and potential antioxidant in active edible film. *Food Science and Nutrition. Food Science & Nutrition* 8: 6338-6345.
- Gharaghani, H.; Shariatmadari, F. & Torshizi, M.A. 2015. Effect of fennel (*Foeniculum vulgare Mill.*) used as a feed additive on the egg quality of laying hens under heat stress. *Revista Brasileira de Ciencia Avicola* 17: 199-208.
- Gholami, M.; Chamani, M.; Seidavi, A.; Sadeghi, A.A. & Aminafshar, M. 2020. Effects of stocking density and environmental conditions on performance, immunity, carcass characteristics, blood constituents, and economical parameters of Cobb 500 strain broiler chickens. *Italian Journal of Animal Science* 19(1): 524-535.
- Houshmand, M.; Azhar, K.; Zulkifli, I.; Bejo, M. & Kamyab, A. 2012. Effects of prebiotic, protein level, and stocking density on performance, immunity, and stress indicators of broilers. *Poultry Science* 91: 393-401.
- Javed, A.; Ahmad, A.; Tahir, A.; Shabbir, U.; Nouman, M. & Hameed, A. 2019. Potato peel waste-its nutraceutical, industrial and biotechnological applications. *Agriculture and Food* 4(3): 807-823.
- Kim, O-K.; Nam, D-E.; Yoon, H-G.; Baek, S.J.; Jun, W. & Lee, J. 2015. Immunomodulatory and antioxidant effects of purple sweet potato extract in LP-BM5 murine leukemia virus-induced murine acquired immune deficiency syndrome. *Journal of Medicinal Food* 18: 882-889.
- Kpanja, E.J.; Duru, S., Kotso, J. & Gonjoh, P.T. 2020. Effect of diets containing graded levels of Irish potato peel meal (IPPM) on carcass quality, haematological and blood biochemical profile of broiler chickens. *Nigerian Journal of Animal Science* 22(3): 242-249
- Kryeziu, A.J.; Mestani, N.; Berisha, S. & Kamberi, M.A. 2018. The European performance indicators of broiler chickens as influenced by stocking density and sex. *Agronomy Research* 16(2): 483-491
- Liang, S. & Armmandio, G. 2014. Chemical and thermal characterization of potato peel waste and its fermentation residue as potential resources for biofuel and bioproducts production. *Journal of Agricultural and Food Chemistry* 62: 8421-8429.
- Liang, S. & Armmandio, G. 2015. Anaerobic digestion of pre-fermented potato peel wastes for methane production. *Nuclear and Chemical Waste Management Journal* 46: 197-200.
- Majid, G.; Mohammad, C.; Alireza, S.; Ali, A.S. & Mehdi, A. 2020. Effects of stocking density and environmental conditions on performance, immunity, carcass characteristics, blood constituents, and economical parameters of Cobb 500 strain broiler chickens. *Italian Journal of Animal Science* 19: 524-535.
- Mohammed, G.; Igwebuikwe, J.U.; Adamu, B.S.; Ashiekh, L.G.; Garba, S.S. & d Kolo, U.M. 2015. Effect of Dietary replacement of maize with Yam and Irish potato peel meals on the growth and economic performance of growing rabbits. *Biokemistri. An International Journal of the Nigerian Society for Experimental Biology* 27(2): 106-110.
- Mohdaly, A.A.A.; Sarhan, M.A.; Smetanska, I. & Mahmoud, A. 2010. Antioxidant properties of various solvent extracts of potato peel, sugar beet pulp and sesame cake. *Journal of the Science of Food and Agriculture* 90: 218-226.

- Nasoetion, M.H.; Atmomarsono, U.; Sunarti, D. & Suthama, N. 2019. Growth performance and lipid profile of broilers fed different levels of purple sweet potato extract and raised under different stocking densities. *Livestock Research for Rural Development* 31(7).
- National Research Council. 1994. *Nutrient Requirements of Poultry*. 9th ed. National Academy of Science. Washington, D.C. 176 p.
- Ong, K.W.; Hsu, A. & Tan, B.K. 2013. Anti-diabetic and anti-lipidemic effects of chlorogenic acid are mediated by ampk activation. *Biochemical Pharmacology* 85(9): 1341-51.
- Park, B.S.; Um, K.H.; Park, S.O. & Zammit, V.A. 2018. Effect of stocking density on behavioral traits, blood biochemical parameters and immune responses in meat ducks exposed to heat stress. *Archive Animal Breeding* 61: 425-432
- Pathak Pranav, D.; Mandavgane Sachin, A. & Bhaskar, D. 2015. Fruit peel waste as a novel low-cost bio adsorbent. *Reviews in Chemical Engineering* 31(4): 361-381.
- Pauzenga, U. 1985. Feeding Parent Stock. *Zootech International publications*. pp: 22-25
- Pompeu, M.A.; Cavalcanti L.F.L. & Toral, F.L.B. 2018. Effect of vitamin E supplementation on growth performance, meat quality, and immune response of male broiler chickens: A meta-analysis. *Livestock Science* 208: 5-13.
- Prasad, A.G.D. & Pushpa H.N. 2007. Antimicrobial activity of potato peel waste. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences* 9: 559-561.
- Qaid, M.; Albatshan, H.; Shafey, T.; Hussein, E. & Abudabos, A.M. 2016. Effect of Stocking Density on the Performance and Immunity of 1- to 14-d- Old Broiler Chicks. *Brazilian Journal of Poultry Science* 18(04).
- Raphael, J.W.; Ojinnaka, P.E.; Tarimbuka, L.I.; Iliya, D.S. & Shehu, I.I. 2017. Growth performance, carcass characteristics and serum biochemistry of broiler chicken fed graded levels of sun-dried Irish potato peel meal. *Turkish Journal of Agriculture - Food Science and Technology* 5(5): 525-529.
- Sabeena Farvin, K.H.; Grejsen, H.D. & Jacobsen, C. 2012. Potato peel extract as a natural antioxidant in chilled storage of minced horse mackerel (*Trachurus trachurus*): Effect on lipid and protein oxidation. *Food Chemistry* 131: 843-851.
- Smetanskaa, I. 2010. Antioxidant efficacy of potato peels and sugar beet pulp extracts in vegetable oils protection. *Food Chemistry* 123: 1019-1026.
- Wang, S.; Nie, S. & Zhu, F. 2016. Chemical constituents and health effects of sweet potato. *Food Research International* 89(1): 90-116.
- Xiao, Z.; He, L.; Hou, X.; Wei J.; Ma, X.; Gao, Z.; Yuan, Y.; Xiao, J.; Li, P. & Yue, T. 2021. Relationships between Structure and Antioxidant Capacity and Activity of Glycosylated Flavonols. *Foods* 10: 849.