

Control of *Salmonella Spp* isolated from Garmat Ali sewage by superoxide radicals produced in vitro.

Rafid M. Karim^{1*}, Ghazi M. Al-Maleky¹ & Ahmed J. Shabeeb¹

¹Department of Marine Biology, Marine Science Center, University of Basrah, Iraq.

Abstract

A modified method was used in the present study to generate superoxide radicals (O_2^-) in vitro, the method involves exposing of riboflavin to lighting and generate O_2^- . *Salmonella Spp* was isolated from sewage of Garmat Ali region in Basrah city, Iraq. This study aimed to reduced *Salmonella Spp* by submitted it to superoxide radicals. A significant reduction in the number of bacteria was revealed after exposed to superoxide radicals at (10, 20, 30, 40 and 50) $\mu\text{g/mL}$ of riboflavin for 10 min of lighting with fluorescent lamp (40 watt), as well as the reduction in bacteria number was observed when the bacteria was exposed to superoxide radicals at $30\mu\text{g/mL}$ of riboflavin for intervals (5, 10, 15, 20, 25 and 30) min of lighting with fluorescent lamp. This study suggesting that reduction in bacteria number was increased as increased riboflavin concentration or increased lighting time.

Introduction

Sewage contamination in drinking and recreational waters is of great concern, because the presence of enteric pathogens threatens public health, Among these organisms are *Salmonella*, which are pathogenic enteric bacteria that can cause salmonellosis in animals and humans, if concentrations able to give rise to infections are present. *Salmonella* is the bacterial pathogen most commonly studied in sewage . In addition, they can survive for long periods of time in sewage sludge and soil, perhaps years under cool moist conditions. Infected humans and animals shed salmonellae into the environment via faeces and ingestion of salmonellae-contaminated food and water (Clyde *et al.*, 1997). Salmonellosis due to infected food handlers has been reported by many authors in different countries (Luby & Jones, 1993; Al-Turki *et al.*, 1998; Senthilkumar & Prabakaran, 2005).

Free radicals such as O_2^- and its derivatives are believed to be the cause of oxygen toxicity. O_2^- is a strong oxidant factor and toxic to cells as the cause of killing bacteria (Kanafani & Martin, 1985). Damage to *Azotobacter chroococcum* and *Escherichia coli* by O_2^- has been demonstrated (Buchanan, 1977; Van Hemmen & Meuling, 1977).

Riboflavin also known as vitamin B₂, is an easily absorbed micronutrient with a key role in maintaining health in humans and animals. Riboflavin has been used as part of the phototherapy treatment of neonatal jaundice, a muscle pain reliever, improve the safety of transfused blood by reducing pathogens found in collected blood and other uses (Powers, 2003; Ball, 2006).

Aim of the study

The aim of the present study was not to survey on presence of *Salmonella Spp* in sewage but the main aim was control of (reducing) this bacteria by submitted it to superoxide radicals produced in vitro according to the modified assay as shown in Fig (1).

Materials and Methods

Isolation and diagnosis:

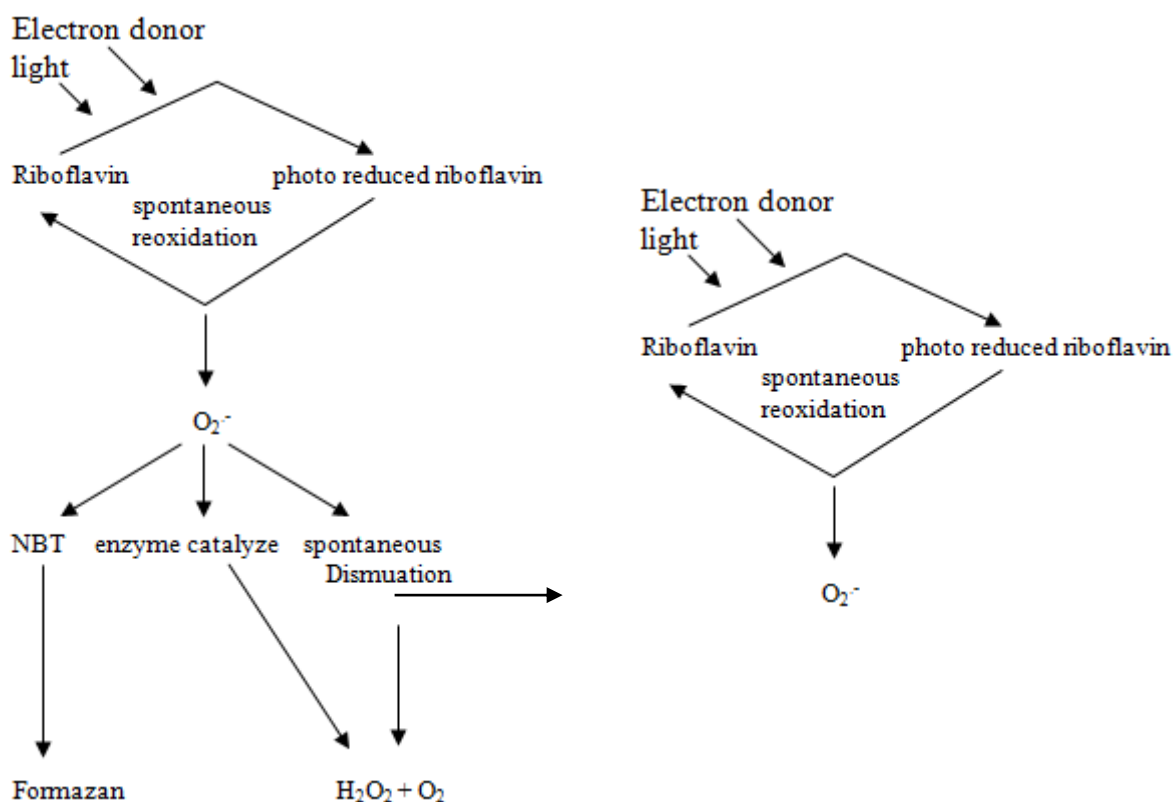
Ten sewage samples were collected from Garmat Ali region in Basrah city, and immediately cultured on *Salmonella Shigella* agar (SS agar), a differentially selective medium for the isolation of pathogenic enteric bacilli, especially those belonging to the genus *Salmonella*, then dishes were incubated at 37 Co for 24 - 48h to obtain on *Salmonella Spp* colonies. Preliminary diagnosis of *Salmonella Spp* depended on cultural and morphological characteristics, it appear as clear, translucent colonies, some with black centers indicating H₂S production.

biochemical tests:

After obtained of colonies from SS agar, in order to detect *Salmonella Spp*, biochemical tests such as gram stain, catalase activity, indole production, carbohydrate fermentation and other tests were used for this aim, table (1) was showed that.

Superoxide production:

For produce superoxide radicals in vitro, the assay of superoxide dismutase activity by Beyer & Fridovich, (1987) was modified as figure below. The procedure was required to the box padded from the inside with aluminum foil and contained fluorescent lamp (40 watt) in addition to riboflavin (B₂ vitamin) which reduced by light and then photo reduced riboflavin interacts with oxygen to generate superoxide radicals (fig 1).



Before modify
(Donnelly *et al.*, 1989)

After modify

Fig(1): Production of superoxide radicals in vitro.

Experiment:

Tubes of dilution 1×10^4 of *Salmonella Spp* were prepared (according to World Health Organization, (2004)) with normal saline at concentrations (0, 10, 20, 30, 40 and 50 $\mu\text{g}/\text{mL}$ riboflavine), the experiment tubes were exposed to lighting (inside the box) for 10 min and cultured on SS agar, then dishes were incubated at 37 C° for 24h.

Tubes of dilution 1×10^4 of *Salmonella Spp* were prepared with normal saline at concentration 30 $\mu\text{g}/\text{mL}$ riboflavine. the experiment tubes were exposed to lighting (inside the box) for intervals 5, 10, 15, 20, 25 and 30 min and cultured on SS agar, then dishes were incubated at 37 C° for 24h.

Results and Discussion

All sewage samples (10) that have been collected from Garmat Ali region were given a heavy growth of *Salmonella Spp* on SS agar, and the table (1) was shown the biochemical tests for diagnosis of *Salmonella Spp*. A significant reducing in Colonies Forming Unit (CFU) of bacteria was revealed after exposed to superoxide radicals at 30 $\mu\text{g}/\text{mL}$ riboflavin for 10 min of lighting (40 watt fluorescent lamp) as compared with non treatment bacteria (0 $\mu\text{g}/\text{mL}$ riboflavin or control), and the reducing in CFU was increased as increased concentrations of riboflavin as it clear in Fig (2). The present study was also revealed a significant diminution in CFU of *Salmonella Spp*

after submitted it to the superoxide radicals at 30µg/mL of riboflavin for different times of lighting and the diminution in CFU was increased as increased lighting time (Fig 3), and the Fig (4) was showed a significant differences between treated and untreated *Salmonella Spp* with O₂⁻ radicals. from these results we can suggesting that production of superoxide radicals were increased by increased riboflavin concentrations, and this was explained the reduction in CFU of *Salmonella Spp* was increased as increased of riboflavin concentrations (Fig, 2). As well as increasing of lighting time that's mean increasing of Exposing the bacteria for long period with superoxide radicals and reasoned CFU reduction (Fig, 3).

The present study was resembled to previous studies, such as the study of (Jaquette *et al.*, 1996) which revealed a significant reduced in population of *Salmonella stanley* inoculated onto alfalfa seeds after the last was treated with different concentrations of chlorine, as well as the study of (Gandhi & Matthews, 2003) was revealed that chlorine or calcium hypochlorite were reduced of *Salmonella* population in alfalfa seeds artificially contaminated with *Salmonella*, also the study of (Zhuang & Beuchat, 1996) was revealed a significantly reduced in population of *Salmonella montevideo* in core tissue of tomatoes dipped in 4–15% TSP (trisodium phosphate), and the study of (Sapers & Jones, 2006) was revealed that hydrogen peroxide (H₂O₂) had efficacy in reduced population of *E. coli* on dip-inoculated tomatoes.

Table (1): biochemical tests for diagnosis of *Salmonella Spp*

The test	Result
Motility	+
Gram's stain	-
Cellular morphology	Rods
Catalase activity	+

Indole production	-
Voges proskauer	-
Triple sugar iron	K/A with H ₂ S
Sugar fermentation	
Glucose	+
Lactose	-
Maltose	-
Xylose	-

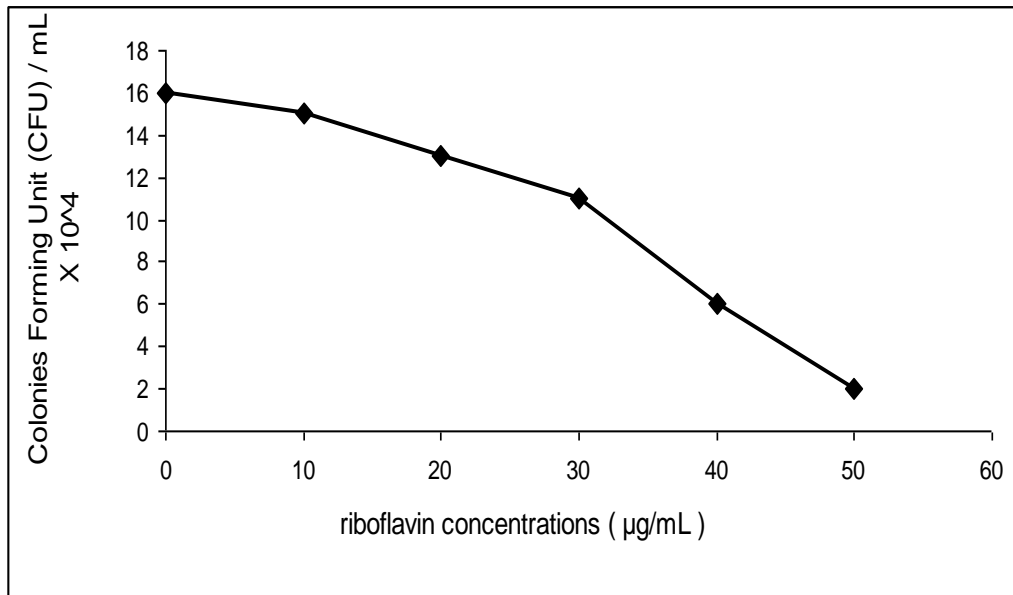
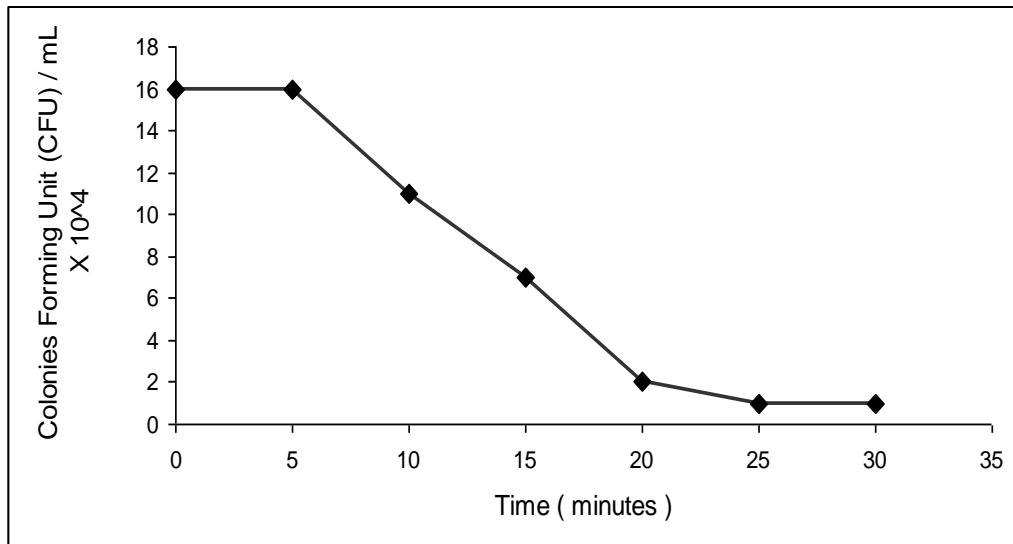
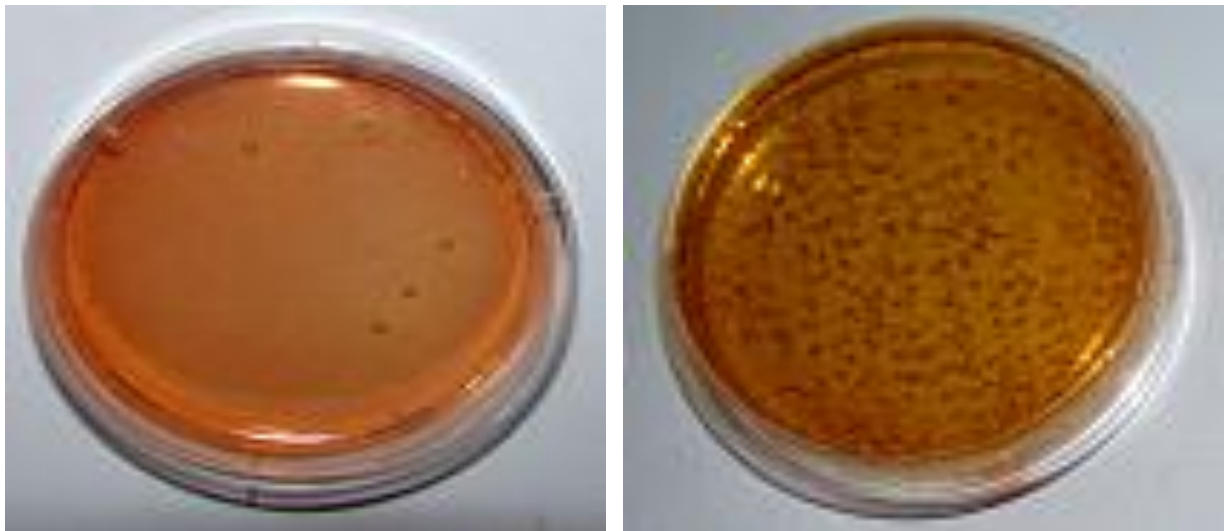


Fig (2): exposure of bacteria to superoxide radicals at different concentration of riboflavin for 10 min.



Fig(3): exposure of bacteria to superoxide radicals at 30µg/mL of riboflavin for different times.



(A)

(B)

Fig(4): comparison between *Salmonella Spp* exposed and non exposed to superoxide radicals.

Where:-

(A) exposed to superoxide radicals at 30µg/mL riboflavin for 15min.

(B) non exposed to superoxide radicals.

Conclusion

From the present study, one can conclude that superoxide radicals have efficacy in elimination on *Salmonella Spp* in sewage samples from Garmat Ali region in Basrah city, and the modified method which was used in current study was very efficient in the production of superoxide radicals, as well as the production of superoxide radicals was increased as increased of riboflavin concentrations according to the modified method of this study.

References

- Al-Turki K.; El-Tahir A. & Bushait S. (1998). Bacterial food poisoning. Saudi medical journal, 19:581-4.
- Ball F.M. G. (2006). Riboflavin in Vitamins in Foods, Analysis, Bioavailability, and Stability. Taylor and Francis Group, New York. P168-175.
- Beyer W. F. & Fridovich I. (1987). Assaying for Superoxide Dismutase activity : some large consequences of minor changes in conditions . Anal. Biochem. 161: 559 – 566 .
- Buchanana G. (1977). The response Of *Azotobacter chroococcum* to oxygen : superoxide- mediated effects. Canadim Journal of Micro- biology 23,1548-1553.
- Clyde V.; Ramsay E. & Bemis D. (1997). Fecal sheddding of *Salmonella* in exotic felids. Journnal of zoology & wildlife medicine, 28:148-52.
- Donnelly J. K.; McLellan K. M.; Walker J. L. & Robinsons D. S. (1989). Superoxide dismutases in foods. A review. Food Chemistry . 33 : 243-270.

- Gandhi M. & Matthews K. R. (2003). Efficacy of chlorine and calcinated calcium treatment of alfalfa seeds and sprouts to eliminate *Salmonella*. International Journal of Food Microbiology, Volume 87, Issue 3, 1 November 2003, Pages 301-306.
- Jaquette C. B.; Beuchat L. R. & Mahon B. E. (1996). Efficacy of Chlorine and Heat Treatment in Killing *Salmonella stanley* Inoculated onto Alfalfa Seeds and Growth and Survival of the Pathogen during Sprouting and Storage. Applied and Environmental Microbiology, July 1996, p. 2212–2215.
- Kanafani H. & Martin SE. (1985). Catalase and superoxide dismutase activities in virulent and nonvirulent *Staphylococcus aureus* isolates. j clin microbiol Apr;21(4):607-10.
- Luby S. & Jones J. (1993). Outbreak of gastroenteritis due to *Salmonella enteritidis* from locally produced grade A eggs, South Carolina. Southern medical journal, 86:1350–3.
- Powers J. H. (2003). Riboflavin (vitamin B-2) and health, Review Article. Am J Clin Nutr. 77:1352–60.
- Sapers G. M. & Jones D. M. (2006). Improved Sanitizing Treatments for Fresh Tomatoes. Journal Of Food Science-Vol. 71: 252-256.
- Senthilkumar B. & Prabakaran G. (2005). Multidrug resistant *Salmonella typhi* in a symptomatic typhoid carriers among food handlers in namakkal district, tamil nadu. Indian Journal of medical microbiology. 23 (2): 92-94.
- Van Hemmen J. J. & Meuling W. J. A. (1977). Inactivation of *Escherichia coli* by superoxide radicals and their dismutation products. Archives of Biochemistry and Biophysics 182, 743-748.
- World Health Organization. (2004). Guidelines for drinking water quality. 3rd ed. Vol. 1.

Zhuang R. Y. & Beuchat L. R. (1996). Effectiveness of trisodium phosphate for killing *Salmonella montevideo* on tomatoes. The Society for Applied Microbiology. 232:97-100.

الخلاصة

السيطرة على بكتريا *Salmonella Spp* المعزولة من مياه الصرف الصحي من منطقة كرمة علي بواسطة جذور السوبرأوكسايد المنتجة خارج الجسم الحي.

رافد محمد كريم¹ * ، غازي مالح¹ و أحمد جري شبيب¹

¹قسم الأحياء البحرية، مركز علوم البحار، جامعة البصرة، العراق.

تم استخدام طريقة محورة في الدراسة الحالية لإنتاج جذور السوبرأوكسايد (O_2^-) مختبرياً، إذ تضمنت الطريقة معاملة الراييوفلافين (B_2) إلى الإضاءة وتحرير جذور السوبرأوكسايد. وهدفت الدراسة إلى استخدام هذه الجذور في السيطرة (قتل) على بكتريا *Salmonella Spp* التي تم عزلها من مياه الصرف الصحي من منطقة كرمة علي في مدينة البصرة. أظهرت الدراسة انخفاض واضح في أعداد البكتريا بعد تعريضها إلى جذور السوبرأوكسايد عند تراكيز (10، 20، 30، 40 و 50) مايكرو غرام/ مليلتر من الراييوفلافين لفترة 10 دقائق من الإضاءة بمصباح فلوريسنت 40 واط. كما لوحظ أيضاً انخفاض كبير في أعداد البكتريا بعد تعريضها إلى جذور السوبرأوكسايد بتركيز 30 مايكرو غرام/ مليلتر من الراييوفلافين لفترات إضاءة مختلفة (5، 10، 15، 20، 25 و 30) دقيقة بمصباح الفلوريسنت، وهذا يشير إلى إن الانخفاض في أعداد البكتريا يزداد بزيادة تركيز الراييوفلافين أو بزيادة فترة التعرض للإضاءة.