ISOLATION AND GENETIC DETECTION OF MORAXELLA BOVIS FROM

BOVINE KERATOCONJUNCTIVITIS IN BASRAH CITY

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ABSTRACT

This study was aimed to determine the most sensitive isolation procedures and evaluate the genetic diversity of *Moraxella bovis* because there are large number of pathogenic bacteria and several other infectious agents such as virus and Mycoplasma have been isolated from the eyes infected with infectious bovine keratoconjunctivitis (IBK). This study included examination of (40) eye swabs, from cows from different ages and regions in Basrah city showed clinical signs of an ocular infection. The isolated bacteria that obtained in pure cultures were non-motile, catalase and oxidase positive, and a clear zone of β -hemolytic colonies were produced on blood agar. According to the growth characteristics, morphology, staining and biochemical tests, the isolated bacteria were identified initially as *Moraxella spp.*, the genetic diversity among *Moraxella spp.* was assessed by 16S rRNA and sequencing as *M. bovis.* The study has also indicated that the isolates of *M. bovis* were resistant to tetracycline, chloramphenicol and erythromycin, However, they were sensitive to penicillin and gentamicin and has an intermediate sensitivity to streptomycin. This study concluded that PCR techniques and sequencing were verified to be most accurate tools to indicate the genetic diversity between *Moraxella spp.* in bovine keratoconjunctivitis.

Key words: Cows, Sequencing, Genetic diversity.

هادي وأخرون

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لمعدي في الأبقار في محافظة البصرة	تهاب القرنية والملتحمة ا	<i>Moraxella bo</i> من ال	عزل وتشخيص جيني لجراثيم <i>evis</i>
[*] فراس طه منصور	^{**} مؤيد حنون صيهود	[*] نورس نوري جابر	[*] ندی صالح هادي
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المستخلص

تهدف هذه الدراسة الى تحديد الطريقة الأكثر دقة في العزل وتقييم التنوع الجيني لجرثومة . Moraxella b وذلك لوجود عدد كبير من الجراثيم الممرضة والعوامل المعدية الأخرى مثل الفيروسات والميكوبلازما التي تم عزلها من العين المصابة بألتهاب القرنية والملتحمة المعدي في الابقار. اشتملت هذه الدراسة على فحص(40) مسحة عين من أبقار ذات اعمار مختلفة ومن مناطق مختلفة في محافظة البصرة والتي أظهرت علامات سريرية لعدوى في العين. الجراثيم المعزولة التي تم الحصول عليها من المزارع النقية كانت غير متحركة، موجبة لأنزيم الكتاليز وانزيم الاوكسيديز، وكذلك انتجت منطقة واضحة من التحلل الكلي لخلايا الدم الحمراء على وسط أجار الدم. وفقًا لخصائص النمو, الخصائص الشكلية, التصبيغ والاختبارات الكيميائية الحيوية، تم مبدئيا تشخيص الجراثيم المعزولة على أنها لخصائص النمو, الخصائص المعزولة على أنها الحصائم العرائيم المعزولة على أنها الحصائم عليها من المزارع النقية كانت غير متحركة، موجبة لأنزيم الكتاليز وانزيم الاوكسيديز، وكذلك انتجت منطقة واضحة من التحلل الكلي لخلايا الدم الحمراء على وسط أجار الدم. وفقًا لخصائص النمو, الخصائص الشكلية, التصبيغ والاختبارات الكيميائية الحيوية، تم مبدئيا تشخيص الجراثيم المعزولة على أنها منحصت على انها جرثومة Moraxella spp. كما أشارت الدراسة إلى أن عزلات M. كانت مقاومة لمضادات التتراسيكلين شخصت على انها جرثومة M. كما أشارت الدراسة إلى أن عزلات M. كانت مقاومة لمضادات التراسيكلين شخصت على انها جرثومة M. معن الظهرت حساسية للبنسلين والجنتاميسين وكذلك لديها حساسية متوسطة الجيني بين جراثيم . والكلورامفينيكول والإريثر وميسين، في حين الظهرت حساسية للبنسلين والجنتاميسين وكذلك لديها حساسية متوسطة تجاه الستربتومايسين. ولالكرامفينيكول والإريثر وميسين، في حين الغرب الحرام الجيني بين جراثيم .

الكلمات المفتاحية: الابقار، التسلسل الجيني، التنوع الجيني

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INTRODUCTION

Infectious bovine keratoconjunctivitis, also known as pink eye is a bacterial eye disease affect cattle distributed all over the world, and can cause considerable economic impact. The main factors that play a role in financial loss are lower weight gain, increase of treatment cost, and market discounts due to eve blindness and disfigurement. The disease is also considered as one of the most common cases affecting beef heifers, and the second most common disease of nursing calves more than three weeks old (12,14). The disease is seen all over the world but mainly in regions with high temperature climates. In addition, this disease in seasonal countries in the summer months are more predominant and it ordinarily appears in younger animals. The numbers of face fly population within the hot months are greater, also dust and intensive sunlight prompt eye infection (10). Moraxella bovis, is a Gram negative bacterium, is considered the main causative agent of IBK. Although this type of infection is usually accompanied with inflammation of eye and most corneal ulcers heal without loss of vision, it can cause permanent or temporary blindness in severe cases and chronic untreated cases (2,20). These bacteria produce hemolysin, which plays a role in damaging neutrophils that are accumulated into the infected area and can also discharge collagen hydrolysis enzymes that play a role in liquefying ulcers (5). The other possible virulence factors of this bacterium include phospholipases (7), iron acquisition systems, outer membrane proteins (16) and proteolytic and hydrolytic enzymes (8). This study was designed for isolation and molecular detection of Moraxella bovis from bovine keratoconjunctivitis in Basrah city, Iraq.

MATERIALS AND METHODS 1- Clinical diagnosis

In this study, keratoconjunctivitis was defined clinically by acute and rapid appearance of photophobia, lacrimation, conjunctival hyperemia, blepharospasm, opacity in the center of the cornea and chemosis that develop during a day of exposure followed by keratitis with corneal edema. The main complaint from the owner was the observation of eye problems in adults and young animals for a few weeks.

2- Sample collection

A total of 40 eye swabs, from cows from different ages and regions in Basrah city showed clinical signs of an ocular infection. The samples were collected using swab sticks soaked with sterile normal saline. The eyelid was opened and the swab stick was gentle rotated from front and back on the cornea and conjunctiva surfaces. The swab was then aseptically dipped in the sterile test tubes containing 5 ml of sterile nutrient broth. Subsequently, all samples were transferred to the lab of microbiology at the College of Veterinary Medicine/ University of Basrah, Iraq.

3- Bacterial isolation and identification

The tubes were incubated aerobically in the incubator at 37°C for 24 hours. The samples were then cultured on blood agar comprising 5% blood of sheep and incubated aerobically at 37°C for 24 hours (13) and then sub cultured to obtain pure colonies. The morphology and overlapping of colonies were determined after Gram staining (17). The pure colonies were kept in brain heart infusion accompanied with agar 1.5% and incubated for 24 hours at 37°C for extraction of DNA and for the purpose of conducting biochemical tests, gelatin hydrolysis, catalase, oxidase, and motility test (23).

4- Molecular detection of *Moraxella bovis* A- Extraction of the DNA

The DNA extraction was performed to all isolates using specific commercial DNA extraction kit, intron biotechnology, cat.no. 17045 following the manufacturer's instruction.

B- Detection of 16S rRNA gene using PCR

Moraxella bovis was detected and diagnosed by amplifying the 16S rRNA fragment using universal primers (18) as shows in Table 1. The reagents of PCR reaction were summarized in Table 2, and PCR condition was described in Table 3.

Primer	Sequence	Tm (°C)	GC (%)	Product size
Forward	5'- AGAGTTTGATCCTGGCTCAG- 3'	54.3	50.0	1250
Reverse	5'- GGTTACCTTGTTACGACTT- 3'	49.4	42.1	base pair

Ta	able 2. Reagents of PCR and	mplification	<u>(25 µl) for 16S r</u>	<u>RNA gene</u>			
	Components		Concentration				
	Taq PCR PreMix		5µl				
	Forward primer		(1 µl)				
	Reverse primer		(1 µl)				
	DNA		1.5µl				
	Nuclease -free Water	Nuclease -free Water					
	Final volume		25µl				
Table	Table 3. PCR condition for amplifying 16S rRNA gene of Moraxella bovis						
No.	Phase	Tm (°C)	Time	No. of cycle			
1-	Initial Denaturation	95°C	5 min.	1 cycle			
2-	Denaturation -2	95°С	45 sec	-			
3-	Annealing	52°C	1 min	35 cycles			
4-	Extension-1	72°C	1 min	-			
5-	Extension -2	72°C	7 min.	1 cycle			

C-Agarose gel electrophoresis

The amplified PCR product was detected using 1% agarose gel prepared with TBE buffer stained with ethidium bromide by used 1500 bp ladder (Promega/USA). The expected size of the amplicon (1250 bp) was estimated by comparison with the standard DNA ladder.

D- Sequencing and sequence alignment

Sequencing of the amplified DNA fragment was analyzed using national instrumentation center for environmental management (nicem) (http://nicem.snu.ac.kr/main/?en skin=index.h tml). Sequence analysis and alignment was carried out using program of Basic Local Alignment Search Tool (BLAST) which is existing at the National Center Biotechnology Information (NCBI) online at (http:// www.ncbi.nlm.nih.gov) and Bio Edit program.

5- Antibiotic susceptibility tests

The antibiotic susceptibility test was performed by disk agar diffusion method (4).

The isolates were tested for susceptibility to 6 different antimicrobial discs, which included Erythromycin E (15 mg), Gentamicin GN chloramphenicol C (30 mg), (10mg), Penicillin PI (30 mg), Streptomycin ST (10mg) and Tetracyclin TE (30 mg).

RESULTS AND DISCUSSION Visual and physical examinations

Forty eye swab from cattle were suffering from eye infection in one or both eyes. The affected eye(s) showed copious lacrimation, closure of the eyelids, photophobia and blepharospasm. Some calves showed copious watery discharge from the affected eve and matting the hair on the lateral aspect of the face . There was severe conjunctivitis and edema resulted in lateral deviation of the eyeball with lacrimation, and opacity of the cornea (Figure 1).



Figure 1. Calf with severe conjunctivitis and edema with corneal opacity

Many calves showed keratitis and yellow opacity of cornea and appearance of nictitating

membrane (third eye lid) (Figure 2)



Figure 2. Calf with yellow opacity of the cornea and ulcer in central cornea

Other calves had ocular discharge, edema and white opacity of cornea and matting of the eye lashes with copious lacrimation. In some cases, the cornea became conical in shape surrounded by a hyperemic zone (Figure 3). Most of the animals resented examination of the eyes had depressed appetite because of the ocular discomfort that resulted in inability to locate food, our clinical findings agree with the findings of Kahn et al (15).



Figure 3. Calf with and opacity of the cornea with hyperemic zone

Isolation and Identification of *Moraxella* bovis

Moraxella bovis was isolated on blood agar and confirmed by determination of the morphology of the characteristics colonies followed by Gram staining. The bacterium isolated from a total of forty eye swabs were non-motile, catalase and oxidase positive (Table 4). According to its growth characteristics, morphology, staining and biochemical tests it was identified initially as *Moraxella bovis* – the etiological agent of infectious bovine keratoconjunctivitis and these results are similar to the results of Mukhtar et al (19). All the isolated bacterium was highly virulent as recognized by the clear zone of hemolysis produced on the blood agar. These findings authenticated the findings of Postma et al (21) who reported that virulent strains of *Moraxella bovis* are formed β -

haemolysin toxins which lysed the corneal epithelial cells, and released cytotoxic toxin and pathogenic hyaluronidase, aminopeptidases, fibrinolysin, and phosphatase.

 Table 4. Laboratory investigation of

 Moraxella bovis

Morphological and biochemical	Result
tests Gram stain	-ve
Motility	-
Hemolytic activity	+ (β)
Oxidase	+
Catalase	+
gelatin hydrolysis	+

Molecular detection of *Moraxella bovis* Amplifying of the 16S rRNA gene by PCR technique: The antigenic and genetic difference among *M. bovis* and the assumed incidence of other micro-organisms existence in IBK can be assessed by comparing different sizes of DNA fragments produced from PCR amplification. Universal primer was used, and the amplification products were visualized on agarose gel with a size of approximately 1250bp (Figure 4). The isolated bacteria were genetically identified by PCR targeting 16S rRNA fragment were sequenced and aligned with the sequences available in GenBank (Figure 5), and this result agree with Helena et al (11) who reported PCR-derived tools verified to be most accurate instrument to indicate the presence of genetic variability between Moraxella spp., also Faraj et al (1) reported that PCR technique followed by phylogenic tree analysis good methods for detection and identification of genetic variants.

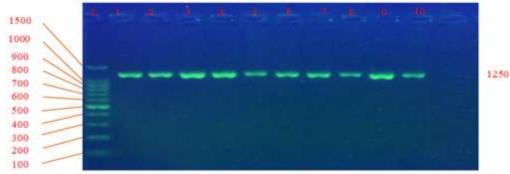


Figure 4. Amplification products of 16S rRNA gene on 1% agarose gel stained with ethidium bromide. The results showed the amplification of 1250 bp of 16S rRNA gene.

Score	Expect	Identities	Gaps	Strand
754 bits(408)	0.0	416/420(99%)	0/420(0%)	Plus/Plus
Query_1	ANTANGCACCOGCTARC	TCTGTGCCAGCAGCCGCGG	TAATACAGAGGGTGCAAGCGTTA	A 60
				1
Shist. 451	AATAAGCACCGGCTAAC	TCTGTGCCAGCAGCCGCGG	TAATACAGAGGGTGCAAGCGTTA	A 510
Query 61	TOGGAATTACTGGGCGT	ARAGCGRGCGTRGGTGGTT	ACTTANGTCAGATGTGAAAGCCC	C 120
			1.1111111111111111111111111111111111111	1
Shist 511	TCGGRATTACTGGGCGT	AAAGCGAGCGTAGGTGGTC	ATTTAAGTCAGATGTGAAAGCCC	C 570
Quarte 121	GGGCTTAACCTGGGAAT	TGCATCTGATACTGGGTGA	CTAGAGTAGGTGAGAGGGAAGTA	G 180
	11111111111111111			1
Skass 571	GGGCTTAACCTGGGAAC	TGCATCTGATACTGGATGA	CTAGAGTAGGTGAGAGGGAAGTA	G 630
Query 181	AATTCCAGGTGTAGCGG	TGAAATGCGTAGAGATCTG	GAGGAATACCGATGGCGAAGGCA	G 240
				1
Shist 621	AATTCCAGGTGTAGCGG	TGAAATGCGTAGAGATCTG	GAGGAATACCGATGGCGAAGGCA	G 690
Query 241	CTTCCTGGCATCATACT	GACACTGAGGTTCGAAAGC	GTGGGTAGCAAACAGGATTAGAT	A 300
				1
Shish 691	CTTCCTGGCATCATACT	GACACTGAGGTTCGAAAGC	STGGGTAGCAAACAGGATTAGAT	A 750

Figure 5. Gene sequencing of Moraxella bovis isolated in the present study

Antibiotic susceptibility tests

Antimicrobial susceptibility, as measured by the standard agar disk diffusion procedure, indicated that the isolates of *M. bovis* were resistant to tetracycline, chloramphenicol and erythromycin, but it was sensitive to penicillin and gentamicin and have an intermediate sensitivity to streptomycin (Table 5). These result disagreements with Grazieli et al (9); Shryock et al (22) which reported that *M. bovis* strains showed resistance to penicillin and gentamicin, respectively. While the results of our study is in agreement with Conceição et al (6); Angelos et al (3) they reported that *M. bovis* strains resistant to erythromycin and tetracycline antibiotic classes, respectively. **sensitivity test**

Table 5. Antibiotic sensiti

Antibiotics	Symbol	Disc content (mg)	Dian	Diameter of inhibition (mm)		
			(Mea	(Mean±standard deviation)		
			Sensitive	intermediate	Resistance	
Tetracycline	ТЕ	30	-	-	≥8	
Penicillin	PI	30	20.5±1.91	-	-	
Chloramphenicol	С	30	-	-	≥6	
Erythromycin	Ε	15	-	-	<u>≥</u> 3	
Gentamicin	GN	10	19.5±1	-	-	
Streptomycin	ST	10	-	12.25 ± 0.50	-	
DEFEDENCES			nhosnholingso	P activity from	n Morarall	

REFERENCES

1. A. A. Faraj, B. F. Hade and A. M. Al-Amery.2019. Conventional and molecular study of *Babesia Spp*. of natural infection in dragging horses at some areas of Baghdad city, Iraq. Iraqi Journal of Agricultural Sciences. 50(3):909-915

2. Ahmed S. 2018. Review on infectious bovine keratoconjunctivitis and its economic impacts in cattle. Biomedicine and Nursing. 4: 34-45

3. Angelos J.A., E.L. Dueger and L.W. George.2000. Efficacy offlorfenicol for treatment of naturally occurring infectious bovine keratoconjunctivitis. J Am Vet Med Assoc. 216:62-64

4. Bauer A. W., W. M. Kirby, J. C. Sherris, and M. Turck. 1966. Antibiotic susceptibility testing by a standardized single disk method. Am. J. Clin. Pathol. 36:493-496

5. Beard M.K. and L.J. Moore. 1994. Reproduction of bovine keratoconjunctivitis with a purified haemolytic and cytotoxic fraction of *Moraxella bovis*. Vet Microbiol. 42: 15-33. 11

6. Conceição F.R., D.M. Bertoncelli and B.O. Storch. 2004. Antibiotic susceptibility of *Moraxella bovis* recovered from out-breaks of infectious bovine keratoconjunctivitis in Argentina, Brazil and Uruguay between 1974 and 2001. Braz J Microbiol. 35:364-366

7. Farn J.L., R.A. Strugnell, P.A. Hoyne, W.P. Michalski and J.M. Tennent. 2001. Molecular characterization of a secreted enzyme with

phospholipase B activity from *Moraxella bovis*. J Bacteriol. 183: 6717-6720

8. Frank S.K. and J.D. Gerber. 1981. Hydrolytic enzymes of *Moraxella bovis*. J Clin Microbiol. 13: 269-371

9. Grazieli Maboni, Leticia T. Gressler, Julia P. Espindola, Marcelo Schwab, Caiane Tasca, Luciana Potter and Agueda Castagna de Vargas. 2015. Differences in the antimicrobial susceptibility profiles of *Moraxella bovis*, *M. bovoculi* and *M. ovis*. Brazilian Journal of Microbiology. 46, 2: 545-549

10. Haskell S. 2008. Blackwell's five-minute veterinary consult: Ruminant. Wiley-Blackwell, Oxford Merck Veterinary Manual, Infectious Keratoconjunctivitis, UK

11. Helena Brocardo Comin, Robert Domingues, Emanuelle Baldo Gaspar, João Rodrigo Gil De Los Santos and Fernando Flores Cardoso. 2020. Genetic differences among *Moraxella* bovis and Moraxella bovoculi isolates from infectious bovine keratoconjunctivitis (IBK) outbreaks in southern Brazil. Genetics and Molecular Biology . 43,2:1-7

12. Holzhauer M., I.J. Visser and K. van Maanen. 2004. Infectious bovine keratoconjunctivitis (IBK) in cows, clinical and lab review at four farms. Tijdschr Diergeneeskd. 129: 526-529.

13. Huimin Liu, Jing Yan, Yutian Wang, Qi Yan, Linping Zhao, Yan Ruoqian, and H.E. Hongxuan.2014. Isolation of *Moraxella bovoculi* from racehorses with keratoconjunctivitis. Journal of Veterinary Diagnostic Investigation. 26(4) 585–587 14. Jeyabal L., D. Debdatta Ray, S. Sureshkannan, K. Nagarajan, S. Visnuvinayagam, S. Ghosh, P.S. Banerjee, S.C. Sekar, M. Bagath, K. Padmanath, K. Rajarajan and P. Ravikumar. 2013. First report of *Morexella bovis* infection in Indian cattle. Adv. Anim. Vet. Sci. 1 (6): 202 – 204.

15. Kahn C.M., S. Line, S.E. Aiello, D.G. Allen, D.P. Anderson, L.B. Jeffcott, K.E. Quesenberry, O.M. Radostits, P.T. Reeves and A.M. Wolf. 2011. The Merck Veterinary Manual, 9th ed. W.B. Sunders Company Ltd., London. Whitehouse Station, NJ, USA

16. Luke N.R., J. Amy, J.A. Howlett, J. Shao and A.A. Campagnari. 2004. Expression of type IV pili by *Moraxella catarrhalis* is essential for natural competence and is affected by iron limitation. Infect Immun. 72: 6262-6270

17. M. F. Hamood, H. N. Jasim and A.S.A. AL-Hassani. 2018. Evalution of contamination statuse in imported and local table eggs. Iraqi Journal of Agricultural Sciences. 49(3):888-898

18. Miller C.S., K. M. Handley, K. C. Wrighton, K. R. Frischkorn, B. C. Thomas and J. F. Banfield. 2013. Short-Read Assembly of

Full-Length 16S Amplicons Reveals Bacterial Diversity in Subsurface Sediments. PLoS ONE 8(2): e56018. doi:10.1371

19. Mukhtar T. Abu Samra, Yassir A. Shuaib, Mazen I. M. Ryhan, Monther B. M. Bayomi, Ahmed O. A. Mohamed, Abdallah A. N. Ali and Amro E. A. Mohammed. 2016. Infectious bovine keratoconjunctivitis in Al-silaite area, Khartoum state. Sch J Agric Vet Sci. 3(3):227-233

20. O'Connor A.M., H.G. Shen, C. Wang and T. Opriessnig. 2012. Descriptive epidemiology of *Moraxella bovis*, *Moraxella bovoculi* and *Moraxella ovis* in beef calves with naturally occurring infectious bovine keratoconjunctivitis (Pinkeye). Vet Microbiol. 155: 374-380

21. Postma G.C., J.C. Carfagnini and L. Minatel. 2008. *Moraxella bovis* pathogenicity: An update. Comp Immunol Microbiol Infect Dis. 31:449–458

22. Shryock T.R., D.W. White and C.S. Werner. 1998. Antimicrobial sus-ceptibility of *Moraxella bovis*. Vet Microbiol. 61:305-309

23. Vanessa Sosa, Ana Umpiérrez, Sofía Acquistapace and Pablo Zunino.2015. Virulence genes in *Moraxella spp.* isolates from infectious bovine keratoconjunctivitis cases. J Infect Dev Ctries. 9(9):1028-1032.