

Effect of Paclobutrazol and potassium on growth

of Date Palm Phoenix dactylifera L. seedlings

*Azhar M. Abdul-Sahib **Mahmood Sh. Hashim **Ibtisam M. Abdul-Sahib

* Date Palm Research Centre, University of Basrah, Basrah, Iraq

**Marin Sciences Centre., University of Basrah, Basrah, Iraq.

Abstract

A field trial was lead at the Centre of Date Palm Researches, Basrah University, to investigate the effect of Paclobutrazol (PBZ) at 0, 100 and 200 mg. L^{-1} and Potassium Sulfate (K₂SO₄) at 0, 2.5 and 5 g. L^{-1} on growth characteristics of *Phoenix dactylifera* L. plants grown from seeds in a factorial experiment designed with R.C.B.D. Most growth parameters recorded were affected by the application of Paclobutrazol and/ or potassium sulfate. Results showed a significant decrease in the number of leaves, leaf length, leaf width, leaf area, and shoot height when applying PBZ. In contrast, shoot dry weight, root length, and root dry weight were increased. Whereas the application of Potassium significantly improved all the previous parameters. On the other hand, total chlorophyll content, dry matter, total soluble carbohydrates, protein content, potassium content in leaves and C/N ratio in leaves showed a significant increase in PBZ and K₂SO₄ treatments. The interaction between the two factors in this trial was found significantly effective in some growth parameters.

Keywords: Paclobutrazol, PBZ, Potassium fertilizer, K₂SO₄, Date palm seedling.

Introduction

Palm trees are of economic importance both as a source of agricultural produce and as an ornamental component in landscaping projects. Palms are globally important as socioeconomic plants (Henderson et al., 1995). Based upon archaeological evidence, the date palm (*Phoenix dactylifera*) was domesticated some 6000 years ago in the Mesopotamian Region (Zohary and Hopf, 2000). Date Palm Phoenix dactylifera L. is a useful traditional medicinal plant belonging to the family Arecaceae; the genus Phoenix contains 12 species, including P. dactylifera which is a subtropical fruit tree widely grown in Iraq and some middle east regions (Barreveld, 1993; Copley et al., 2001; Sirisena and Ajlouni, 2015). Date palm trees require relatively great amounts of macro and micronutrients to achieve good growth and reasonable economic production. Fertilization is, therefore, one of the important practices that increase date production and improve fruit quality. Properly applying of macro and micro nutrients is necessary to increase the quantitative, qualitative, and economic output of date production in palm groves (Shaaban and Mahmoud, 2012). Seeds were the means to intentionally reproduce date palms for millennia in Mesopotamia, and later in North Africa, from native wild species or through seed introductions. From these two areas, seeds were transported, in association with human migrations or invading armed forces, east to present-day Pakistan, into southern Europe, and later to South and North America, on southern Africa, and to distant Australia and New Caledonia (Johnson, 2010). Paclobutrazol, a plant growth regulator from imperial chemical industries, has been very effective for dwarfing many crops (Lever et al., 1982; Menhenett and Hanks 1982). It has been documented to reduce plant growth without directly interfering with secondary metabolite biosynthetic pathways, as it inhibits gibberellin synthesis downstream in the chain of reactions leading to the production of secondary metabolites, i.e., the biosynthesis of tannins, phenolic compounds, and terpenoids inhibit GA biosynthesis and act as inhibitors of mono-oxygenases catalyzing the oxidative steps from ent-kaurene to ent-kaurenoic acid of biosynthesis path way of GA and employed magnificently to reduce the height of date palm (Rademacher, 2000; Cohen et al., 2013). Potassium is necessary for basic physiological functions, such as the formation of sugars and starch, the synthesis of proteins, cell division and growth, fruit formation and could improve fruit size, flavor and color (Abbas and Fares, 2008). Moreover, Potassium is also an important nutrient for date palm growth and productivity (Al-Kharusi et al., 2009). Researchers have obtained many positive and clear indications that the date palm does not differ from other plants in response to foliar spraying. All these experiments recommended using their results and conducting other operations to develop the reality of date palm cultivation globally; one of the nutrients used in feeding date palms is potassium sulfate (Shareef et al., 2021). In this regard, potassium is a mobile nutrient, it is well adapted to foliar fertilization because it is rapidly translocated from the leaves to other plant parts and consequently increases yield and components (Mengel, 2002).

The addition of nutrients to plants through spray application ensures the nutrient is inputted directly into the plant and enhances plant tissue metabolism, reducing energy consumption. The foliar technique increases the potential for mixing nutrients with growth regulators, saving much effort and time (Salvagiotti*et al.*, 2008), as the power of plant leaves to absorb nutrients has resulted in the fact that the foliar application of nutrients becomes a recurrent method for supplying nutrients to plants (Swietlik and Faust, 1984).

For the importance of the date palm plant, this experiment was applied to investigate the effect of Paclobutrazol and Potassium Sulfate on the growth of *Phoenix dactylifera* L. seedlings and likewise to find mathematical relationships that serve environmental studies.

Materials and Methods

A field trial was carried out at the of Date Palm Research Center -University of Basrah during 2016/2017 to study the effect of Paclobutrazol (the commercial product from the German company Syngenta Agro AG) and Potassium Sulfate and their treatment combinations on growth characteristics of Phoenix dactylifera L. seedlings. One-year-old Date Palm Barhi cv. seedlings were transplanted to 25 cm plastic pots (one plant per pot) containing a 6:2:1 (v/v) mix of sand: peat moss: organic manure. Pots were kept in a greenhouse and were watered whenever they need. Three concentrations of PBZ (0, 100 and 200 mg. L^{-1}) and K₂SO₄ at 0, 2.5 and 5 g. L^{-1} were prepared as distilled water was used for control plants. Two weeks after transplanting, seedlings were sprayed with the PBZ and K₂SO₄ solutions applied until run off once a week, for a duration of eight weeks as a foliar spray. At the end of the experiment, seedlings were extracted from pots carefully using water and data was tabulated on: shoot height (cm): was measured from the soil surface until the bottom of the leaves; number of leaves, leaf length (cm), leaf width (cm), average of one leaf area (cm²): was measured according to Dvorinic (1965); total chlorophyll content in leaves (mg. 100 g⁻¹ fresh wt.): was estimated according to Goodwin (1976); dry weight of shoot (without leaves as leaves were separated from shoot): was measured by replacing in a circulatory air oven at 65 C° for 72 hours until the weight stabilized; dry matter in leaves (%): fresh weight of leaves was recorded then leaves were dried in a circulatory air oven at 65 C° for 72 hours until the weight stabilized then dry matter was calculated; total soluble carbohydrates content in leaves (%): was estimated according to DuBois et al., (1956); protein (%) in leaves: nitrogen was estimated as described by Jackson (1958), then protein was evaluated according to the reported procedures in A.O.A.C. (1970) using the equation: Protein = N % X 6.25; K% in leaves: was estimated using flame photometer as described by (Page *et al.*, 1982); C/N ratio (as C/N = total soluble carbohydrate/ nitrogen content in leaves); root length (cm); roots dry weight (g): roots were dried in a circulatory air oven at 65 C° for 72 hours until the weight stabilized then dry matter was calculated.

The applied experimental design of this trial was a randomized complete block design (Al-Rawi and Khalaf-Alla, 2000). All data collected were analyzed using analysis of variance (ANOVA) with the GenStat Statistical Analysis System (2007). Seventh Edition (DE3). Means were compared using Least Significant Difference (LSD) ($P \le 0.05$).

Results and Discussion

Plant growth performances represented in Table (1) revealed that the number of leaves, leaf length, leaf width and means of leaf area were significantly reduced with the application of PBZ. While a significant increase in the same parameters took place when applying K_2SO_4 as compared to the control treatment. On the other hand, the combined treatments of PBZ and K₂SO₄ affected some of these growth characteristics significantly, as the highest values in number of leaves and those of leaf length (3.58 and 37.21 cm respectively) were recorded when spraying with 0 mg. L^{-1} of PBZ + K₂SO₄ at 5 g. L^{-1} . While the lowest values in same two parameters (2.04 and 21.33 cm respectively) were recorded when spraving with 200 mg. L^{-1} of PBZ + 0 g. L^{-1} of K₂SO₄. On the other hand, 0 + 0 and 0 + 5 g. L^{-1} of PBZ and K_2SO_4 scored the greatest values (0.95 cm and 43.81 cm²) in leaf width and leaf area, respectively and the lowest values $(0.56 \text{ cm and } 27.67 \text{ cm}^2)$ were found in 200 mg. L⁻ 1 + 0 and 200 + 5 g. L⁻¹, of PBZ and K₂SO₄ respectively. It has been suggested that the effectiveness of inhibition in shoot and leaf growth by PBZ is determined by the concentration of the chemical used, the plant species under study, and its stage of development (Lee et al., 1998; Chaney, 2004). On the other hand, the increases in physical characteristics when applying Potassium Sulfate may be due to K important role in pH stabilization, osmoregulation, enzyme activation, protein synthesis, stomata movement, photosynthesis, cell extension, cell division, cell size, cell number and cell turgidity. These results are in accordance with (Mengel and Kirkby, 2001; Dialami and Mohebi, 2010). El-Sabagh (2012) and Amro et al., (2014) informed that translocation of photosynthetic assimilations depends on the concentration of cell potassium. Potassium is involved in controlling cell water content and photosynthetic activity. Furthermore, the beneficial influence of K might be attributed to the enhancing of many metabolic which reproduced on yield development (David et al., 1998).

Treatment			No. of Leaves	leaf length cm	leaf width cm	leaf area cm²
DD7 com		0	3.31	30.97	1.34	37.91
$\mathbf{PBZ} \text{ con.}$		100	3.24	23.42	1.34	30.45
Nig. L		200	2.81	22.84	1.00	28.03
L.S.D.(<i>P</i> ≤0.05)			0.050	0.755	0.032	0.778
V SO		0	2.76	23.17	0.94	29.91
$K_2 SO_4 \text{ con.}$		2.5	3.20	25.88	1.24	32.81
g. L		5	3.40	28.18	1.50	33.69
L.S.D. $(P \le 0.05)$			0.050	0.755	0.032	0.778
		0	3.10	25.99	0.95	31.68
	0	2.5	3.25	29.69	1.20	38.25
PBZ con.		5	3.58	37.21	1.86	43.81
Mg. L		0	3.14	22.19	1.32	30.24
	100	2.5	3.22	23.82	1.35	31.54
K ₂ SO ₄ con. g. L ⁻¹		5	3.36	24.24	1.35	29.58
		0	2.04	21.33	0.56	27.80
	20(2.5	3.14	24.11	1.16	28.63
		5	3.26	23.08	1.29	27.67
L.S.D. $(P < 0.05)$			0.087	1.308	0.055	1.348

Table 1 . Effect of PBZ and K2SO4 on the morphological characteristics of leavesof Date Palm seedlings

It was obtained from the data in Table (2) that shoot height was significantly reduced in the high PBZ concentrations and increased when adding K_2SO_4 when compared to control plants in this study. Whereas root length and dry weights of shoots and roots were increasing significantly when applying PBZ or K_2SO_4 . Results also revealed that some treatments of PBZ and K_2SO_4 used in a combination caused a significant increase in shoot height, shoot dry weight, root length and root dry weight values.

The relationship between shoot height and root length of date palm seedlings is represented in Figure 1.

Treatment			Shoot Height Cm	Shoot Dry weight G	Root length Cm	Root Dry weight g	
DD7 con		0	4.39	12.93	31.14	12.31	
$M_{\alpha} L^{-1}$		100	3.57	13.28	36.01	12.24	
Mg. L		200	3.15	14.17	39.29	13.95	
L.S.D.(<i>P</i> ≤0.05)			0.116	0.180	2.038	0.144	
V CO		0	3.25	13.02	34.02	12.31	
$K_2 SO_4 \text{ con.}$ g. L ⁻¹		2.5	3.90	13.44	35.70	13.00	
		5	3.96	13.93	36.72	13.20	
L.S.D. _{(P}	L.S.D. $(P < 0.05)$		0.116	0.180	2.038	0.144	
		0	4.15	12.20	29.59	11.88	
	0	2.5	4.48	13.23	31.54	12.58	
PBZ con.		5	4.53	13.36	32.29	12.47	
Mg. L ⁻		0	3.51	12.89	34.33	11.57	
K-SO, con	10(2.5	3.56	12.87	37.25	12.58	
$K_2 SO_4 con.$ g. L ⁻¹		5	3.64	14.08	36.46	12.58	
		0	2.08	13.96	38.14	13.47	
	20(2.5	3.65	14.21	38.31	13.83	
		5	3.72	14.34	41.42	14.54	
$L_{1}S_{2}D_{2}(B < 0.05)$			0.202	0.312	3.530	0.249	

Table 2. Effect of PBZ and K₂SO₄ on the physical characteristics of shoot and roots of Date Palm L. seedlings.

Values are means of four replications



Figure 1. The relationship between shoot height and root length (cm) of date palm seedlings.

PBZ employed magnificently to reduce the height of the date palm (Cohen *et al.*, 2013). Also according to Berova and Zlatev (2000), the application of PBZ resulted

in both of the reduction of plant height and the increasing of stem thickness in young plants, it contributed to an enhancement of seedling quality at planting. Likewise, PBZ reduced the plant height, number of new shoots, length of internode, and total plant weight. The percentage of new shoots, length and diameter of terminal shoots, number of leaves and leaf area were significantly reduced in mango (*Mangifera indica* L.) plant (Khader, 1991; Singh *et al.*, 2004).

K is an activator of many important enzymes, such as protein synthesis, sugar transport, N and C metabolism and photosynthesis. Furthermore, K is very important for cell growth which is vital for the function and development of plants (Hepler et al., 2001; Marschner, 2012; Oosterhuis et al., 2014). About the growth-promoting mechanism of K, it is generally granted that K stimulates and controls ATPase in the plasma membrane to generate acid stimulation, then triggers cell wall loosening and hydrolase stimulation whih promots cell growth. K has strong mobility in plants and plays an important role in regulating cell osmotic pressure and balancing the cations and anions in the cytoplasm (Kaiser, 1982; Oosterhuis et al., 2014; Hu et al., 2016). Through these processes, K is involved in regulating stomatal opening, cell elongation, and other important physiological processes. Also adequate K supply can promote N metabolism and enhance the synthesis of amino acids and proteins (Ruan et al., 1998; Ruiz and Romero, 2002). Also, PBZ influences the isoprenoid pathway and changes the status of phytohormones by inhibiting gibberellin synthesis, decreasing ethylene production, and enhancing cytokinin and ABA contents (Kamountsis and Sereli 1999). PBZ has been testified to inhibit GA biosynthesis in plants by inhibiting kaurene oxidase, a Cyt P-450 oxidase, thus blocking the oxidation of kaurene to kaurenoic acid (Dalziel and Lawrence, 1984). Browning et al.,(1992) investigated the effect of PBZ on the translocation of endogenous IAA (indol- 3-acetic acid) as they found that PBZ caused a slower movement of IAA in shoot tips. In this study, the results obtained indicated that PBZ or K₂SO₄ applications led to a significant increase ($p \le 0.05$) in chlorophyll content, dry matter percentage, percentage of carbohydrates, protein content, potassium content and C/N ratio in leaf tissue, compared to the control plants values (Table 3). Regarding the treatment combinations, results showed that spraying with 200 mg. L^{-1} of PBZ + K_2SO_4 at 5 g. L⁻¹ scored the highest values in chlorophyll content (10.94 mg. g⁻¹

fresh wt.), dry matter in leaves (5.14 %), carbohydrates content (53.58 %), protein content (9.00 %) and C/N ratio (37.22). Whereas, the highest value of potassium content (0.74 %) was found when spraying with 200 mg. L^{-1} of PBZ + K₂SO₄ at 2.5 g. L^{-1} .

Treatment		Total chlorophyll mg .100 g ⁻¹ fresh wt.	Dry matter (%)	Carbohydrates (%)	Protein (%)	Potassium (%)	C/N ratio		
DD7		0	9.00	3.35 31.61		7.63	0.53	25.92	
$M_{\alpha} \mathbf{L}^{\cdot 1}$	•	100	10.48	3.72	37.85	8.56	0.57	27.66	
Mg. L		200	10.65	4.24	47.68	8.88	0.67	33.58	
L.S.D. _(P ≤0.05)		0.261	0.429	2.194	0.125	0.040	1.677		
$\begin{array}{c c} K_2 SO_4 & 0\\ con. & 2.5\\ g. L^{-1} & 5 \end{array}$		0	9.74	3.60	36.23	8.06	0.53	27.89	
		2.5	10.10	3.59	38.92	8.29	0.63	29.23	
		5	10.29	4.11	41.99	8.70	0.62	30.04	
L.S.D. $(P < 0.05)$		0.261	0.429	2.194	0.125	0.040	1.677		
		0	8.45	3.28	28.25	7.19	0.47	24.57	
PBZ	0	2.5	9.19	3.31	32.35	7.38	0.56	27.41	
con.		5	9.35	3.45	34.23	8.31	0.57	25.79	
Mg. L^{-1}		0	10.33	3.67	37.21	8.25	0.50	28.19	
$\begin{array}{c} \mathbf{X} \\ \mathbf{K}_2 \mathbf{SO}_4 \\ \mathbf{con.} \\ \mathbf{g}_2 \mathbf{L}^{-1} \end{array}$	10(2.5	10.54	3.74	38.17	8.63	0.59	27.65	
		5	10.57	3.75	38.18	8.79	0.62	27.13	
		0	10.44	3.84	43.22	8.75	0.61	30.90	
8. —	200	2.5	10.58	3.73	46.24	8.88	0.74	32.63	
		5	10.94	5.14	53.58	9.00	0.66	37.22	
L.S.D.(<i>P</i> ≤0.05)		0.452	0.743	3.799	0.216	0.070	2.905		

Table 3.	Effect	of PBZ	and	K ₂ SO ₄ on	the	chemical	analysis	in	leaves	of	Date
Palm see	dlings										

The relationship between Protein content (%) and C/N ratio of date palm seedlings is represented in Figure 2.



Figure 2. The relationship between Protein content (%) and C/N ratio of date palm seedlings.

An increased in photosynthetic rate in *Catharanthus* roseus (Abdul Jaleel *et al.*, 2007) and potato (Tsegaw*et al.*, 2005) exposed to PBZ has been reported previously. This was attributed to this higher chlorophyll content of the leaf tissue. Davis *et al.*, (1986, 1988) reported that PBZ application resulted in various physiological changes that are generally associated with optimized yield in various plants as the most prominent changes include: repression of senescence with enhanced concentrations of photosynthetic pigments in plant tissues, alteration of hormonal balance, improvement in mineral absorption and carbohydrate synthesis.Wang and Steffens (1987) informed that PBZ was also shown to shift assimilate partitioning from leaves to roots, increasing carbohydrates in all parts of seedlings, rising chlorophyll and protein content and mineral element concentrations in leaf tissue, intensifying root respiration and reducing water use.

References

- Abbas, F., Fares, A. (2008). Best management practices in citrus production. Tree For. Sci. Biotech. 3, 1-11.
- Abdul Jaleel, C; Manivannan, P; Sankar, B; Kishorekumar, A; Sankari, S and Panneerselvam, R. (2007). Paclobutrazol enhances photosynthesis and ajmalicine production in Catharanthusroseus. Process Biochemistry, 42: 1566-1570.
- Al-Kharusi, L.M., Elmardi, M.O., Ali, A., Al-Julanda, F., Al-Said, L., Abdelbasit, K., Al-Alpha, J.M., Chen. J, Zhang, G. (2009). Effect of nitrogen fertilizer forms on growth, photosynthesis, and yield of rice under cadmium stress. J. Plant Nutrition, 32 (2): 306-317.

- Al-Rawi, K.M. and A. Khalaf-Alla (2000). Design and Analysis of Agricultural Experiments. College of Agriculture and Forestry. University of Mosul, Iraq, 487 pp. (In Arabic).
- Amro, S., M. Salama, Omima, M. El-Sayed and Osama, H.M. El-Gammal, (2014). Effect of effective microorganisms (EM) and potassium sulphate on productivity and fruit quality of "Hayany" date palm grown under salinity stress. J. of Agric. and Veterinary Sci., 7: 90-99.
- A.O.A.C. (1970). Official Methods of Analysis. 11th ed. Association of Official Analytical Chemist. USA.
- Barreveld, W. H. (1993). Date palm products, FAO Agricultural Services . Bulletin No. 101.
- Berova, M. and Zlatev, Z. (2000). Physiological response and yield of Paclobutrazol treated tomato plants (*Lycopersiconesculentum* Mill.). Plant Growth Regulation, 30: 117-123.
- Browning, G.; Kuden, A. and Blanke, P. (1992). Site of (2RS, 3RS) paclobutrazol promotion of axillary flower initiation in pear cv. Doyenne du Comice. Journal of Horticultural Science. 67(1):121-128
- Chaney, WR. (2004). Paclobutrazol: More than Just a Growth Retardant. Pro-Hort Conference. Peoria, Illinois.
- Cohen, Y.; Aloni, D D.; Adur, U; Hazon, H. and Klein, J D. (2013). Characterization of growthretardant effects on vegetative growth of date palm seedlings. J. Plant Growth Regulation, 32: 533-541.
- Copley, MS. ; Rose, PJ. ; Clapham, A. ; Edwards, DN.; Horton, MC. and Evershed, RP. (2001). Detection of palm fruit lipids in archaeological pottery from QasrIbrim, Egyptian Nubia. ProcBiol Sci. 268 (1467): 593-597
- Dalziel, J. and Lawrence, DK. (1984) . Biochemical and biological effects of kaurene oxidase inhibitors such as paclobutrazol. Monograph of British Plant Growth Regulators Group 11:43-57
- David, D.W., B.C. Darst, R.T. Roberts, S.O. Fox, W.R .Agerton, S.J. Couch, S.K. Rogers. (1998). Better Crops with Plant Food, 82(3):28-29.
- Davis, T. D.; Sankhla, N. and Upadhyaya, A. (1986). Paclobutrazol: A promising plant growth regulator. In S. S. Purohit (Ed.), Hormonal regulation of plant growth and development (pp. 311–332). Bikaner: Agrobotanical Publishers.
- Davis, T. D., Steffens, G. L., and Sankhla, N. (1988). Triazol plant growth regulators. Horticultural Review, 10, 151–188.
- Dialami, H. and Mohebi, A.H. (2010). Increasing yield and fruit quality of date palm 'Sayer' with application of nitrogen, phosphorus and potassium optimum levels. Members of scientific staff at date palm and tropical fruits research institute of Iran.
- DuBois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. (1956).Colorimetric method for determination of sugars and related substances. Anal Chem. 28(3):350–356.

- Dvorinic, V. (1965) Lucrări practice de ampelografie. EdituraDidacticășiPedagogică, București, R. S. Romania, pp. 1–15.
- El-Sabagh, A.S. (2012). Effect of bunches spraying with some macro and micronutrient on fruit retention and physical characteristics of DegletNour date palm cultivar during Kimiri stage. Res. J. Agric. Biol. Sci., 8: 138-146.
- Goodwin, T. W. (1976). Chemistry and biochemistry of plant pigments. 2nd ed., Academic Press, London, N.Y., San Farncisco. USA, p. 373.
- Henderson, A.; Galeano G. and Rodrigo B. (1995). Field guide to the palms of the Americas. Princeton: Princeton University Press, p 351.
- Hepler, P. K., Vidali, L., and Cheung, A. Y. (2001). Polarized cell growth in higher plants. Annu.Rev. Cell. Dev. Biol. 17, 159–187. doi: 10.1146/annurev.cellbio.17. 1.159
- Hu, W., Jiang, N., Yang, J., Meng, Y., Wang, Y., Chen, B., et al. (2016). Potassium (K) supply affects K accumulation and photosynthetic physiology in two cotton (*Gossypiumhirsutum* L.) cultivars with different K sensitivities. Field Crop. Res. 196, 51–63. doi: 10.1016/j.fcr.2016.06.005
- Jackson M. L. (1958). Soil chemical analysis prentice. Hall & Englewood, Cliffs, N. T. USA.
- Johnson, D. V. (2010). Worldwide dispersal of the date palm from its homeland. Acta Hort. 882:369-375.
- Kaiser, W. M. (1982). Correlation between changes in photosynthetic activity and changes in total protoplast volume in leaf tissue from hygro-, mesoand xerophytes under osmotic stress. Planta 154, 538–545. doi: 10.1007/ BF00402997
- Kamountsis, A. P., and Sereli, C. (1999). Paclobutrazol affects growth and flower bud production in gardenia under different light regimes. Hort Science, 34, 674–675.
- Khader, S. E. S. A. (1991). control of tree height, trank girth, shoot growth and total assimilation in young grafted mango trees by paclobutrazol, Indian J. Hort., 48: 112-115.
- Kishore, K. Singh, HS.Kurian, RM. (2015). Paclobutrazol use in perennial fruitcrops and its residual effects: A review.Indian Journal of Agricultural Sciences.85(7):863-872
- Lee, I.; Foster, K R. and Morgan, P W. (1998). Effect of gibberellin biosynthesis inhibitors on native gibberellin content, growth and floral initiation in Sorghum bicolor. Plant Growth Regulation, 17: 185–195.
- Lever, BG.; Shearing, SJ.; Batch, JJ. (1982). PP333 a new broad-spectrum growth retardant. Proceedings of British Crop Protection Conference Weeds. British Crop Protection Council, Croydon, pp 3- 10.
- Marschner, H. (2012). Marschner's Mineral Nutrition of Higher Plants. Cambridge, MA: Academic press.

- Mengel, K. (2002). Alternative or complementary role of foliar supply in mineral nutrition. ActaHortic: 33–47.
- Mengel, K. and Kirkby, E.A. (2001). Principle of plant Nutrition, 5th Ed., Kluwer Academic Publishers.
- Menhenett, R.; Hanks, GR. (1982). Comparisons of a new triazole retardant PP333 with ancymidol and other compounds on pot-grown tulips. Plant Growth Regulation 1: 173-181.
- Oosterhuis, D., Loka, D., Kawakami, E., and Pettigrew, W. (2014). The physiology of potassium in crop production. Adv. Agron. 126, 203–234. doi: 10.1016/B978-0-12-800132-5.00003-1
- Page A. L., Miller R. H., Keeny D. R., (1982) Methods of soil analysis. Part 2, 2nd ed., Madison, Wisconson, U.S.A., p. 1159.
- Rademacher, W. (2000). Growth retardants: Effects on gibberellin biosynthesis and other metabolic pathways. Annual Review of Plant Physiology and Plant Molecular Biology 51:501–531.
- Ruan, J., Wu, X., Ye, Y., and Hardter, R. (1998). Effect of potassium, magnesium and sulphur applied in different forms of fertilisers on free amino acid content in leaves of tea (*Camellia sinensis* L.). J. Sci. Food Agric. 76, 389–396. doi: 10.1002/(SICI)1097-0010(199803)76:3<389::AID-JSFA963<3.0.CO;2-X</p>
- Ruiz, J., and Romero, L. (2002). Relationship between potassium fertilisation and nitrate assimilation in leaves and fruits of cucumber (Cucumissativus) planleets. Ann. Appl. Biol. 140, 241–245. doi: 10.1111/j.1744-7348.2002.tb00177.x
- Salvagiotti, F., Cassman, K.G., Specht, J.E., Walters, D.T., Weiss, A. (2008). Nitrogen uptake, fixation and response to fertilizer N in soybeans. A review. Agronomy and Horticulture — Faculty Publications, 133.
- http://digitalcommons.unl.edu/agronomyfacpub/133
- Shaaban, S.H.A. and Mahmoud, M.S. (2012). "Nutritional evaluation of some date palm (*Phoenix dactyliferaL.*) cultivars grown under Egyptian conditions", *Journal of American Science*, Vol. 8 No. 7, pp. 135-139.
- Shareef, J.; Ibtihaj, H. Al-Tememi; GholamrezaAbdi. (2021). Foliar nutrition of date palm: advances and applications. A review. FOLIA OECOLOGICA – vol. 48, no. 1 . 2021. doi: 10.2478/foecol-2021-0010
- Singh, N.P., Malhi, C.S., Dhillion, W-s. (2004). Effect of plant bioregulators on the promotion of flowering in mango cv. Dashehari. Journal of Research - Punjab Agricultural University, 41(3): 341-344.
- Sirisena S, Ng K, Ajlouni S. (2015). The Emerging Australian Date Palm Industry: Date Fruit Nutritional and Bioactive Compounds and Valuable Processing By-Products. Compr Rev Food Sci Food Saf; 14 (6): 813-823.
- Swietlik, D. and M. Faust, (1984). Foliar nutrition of fruit crops. Hort. Rev., 6: 287-355.

- Tsegaw, T; Hammes, S and Robbertse, J (2005). Paclobutrazol-induced leaf, stem, and root anatomical modifications in potato. American Society for Horticultural Science, 40: 1343-1346.
- Wang CY, Steffens GL. (1987): Postharvest responses of 'Spartan' apples to preharvestpaclobutrazol treatment. HortScience. 22:276-278
- Zohary, D. and M. Hopf. (2000). Domestication of plants in the Old World. 3rd ed. Oxford University Press, U.K.

تأثير الباكلوبوترازول والبوتاسيوم في نمو بادرات نخلة التمر . Phoenix dactylifera L

أحمد ماضي وحيد المياحي st

أزهار مهدي عبد الصاحب ^{*}

ابتسام مهدي عبد الصاحب ^{*}

محمود شاکر هاشم ً

*مركز أبحاث النخيل ، جامعة البصرة ، العراق

**مركز علوم البحار ، جامعة البصرة ، العراق.

الخلاصة

Paclobutrazol الدراسة في مركز أبحاث النخيل / جامعة البصرة ، لبحث تأثير البكلوبيترازول PBZ) تم إجراء هذه الدراسة في مركز أبحاث النخيل / جامعة البصرة ، لبحث تأثير البكلوبيترازول (PBZ) (PBZ) بالتراكيز 0 و 2.5 و 5 مغم. لتر⁻¹ على بعض خصائص نباتات النخيل . *Phoenix dactylifera* L النامية من البذور في تجربة عم. لتر⁻¹ على بعض خصائص نباتات النخيل . R.C.B.D. ملفر *dactylifera* L النامية من البذور في تجربة عاملية مصممة باستخدام تصميم القطاعات العشوائية الكاملة .R.C.B.D. تأثرت معظم صفات النمو التي تم معنويا في عدد الأوراق وطول الورقة وعرض الورقة ومساحة الورقة وارتفاع النبات عند استعمال النائج انخفاضا معنويا في عدد الأوراق وطول الورقة وعرض الورقة ومساحة الورقة وارتفاع النبات عند استعمال Advertice دفي حين أدت معنويا في عدد الأوراق وطول الورقة وعرض الورقة ومساحة الورقة وارتفاع النبات عند استعمال الماد . ولا أحدة معنويا في عدد الأوراق وطول الورقة وعرض الورقة ومساحة الورقة وارتفاع النبات عند استعمال PBZ ، من الحية ثانية، حدثت زيادة معنوية في الوزن الجاف للساق وطول الجذر والوزن الجاف للحزر . في حين أدت الحماية النبات عاد التعمال . ولا العذر . في حين أدت معنويا في عدد الأوراق وطول الورقة وعرض الورقة ومساحة الورقة وارتفاع النبات عند استعمال PBZ ، من الحية ثانية، حدثت زيادة معنوية في الوزن الجاف للساق وطول الجذر والوزن الجاف للجذر . في حين أدت المافة البوتاسيوم إلى زيادة معنوية في الوزن الجاف الساق وطول الجذر والوزن الجاف للجزر . في حين أدت الحافة البوتاسيوم إلى زيادة معنوية في جميع الصفات السابقة. من ناحية أخرى ، أظهر محتوى الكاوروفيل واضافة البوتاسيوم إلى زيادة معنوية في جميع الصفات السابقة من ناحية أخرى ، أظهر محتوى الكاوروفيل واضافة البوتاسيوم إلى زيادة معنوية في جميع الصفات السابقة في معاملات كالوراق والوليل محموى الكاروفيل الخري والمادة الجافة والكربوهيدرات الذائبة الكلية ومحتوى البروتين ومحتوى البوتاسيوم في الأوراق ونسبة الكاربوهيدرات الى النتروجين C/N في بعض صفات النماني والمان في هذه التجربة في الأوراق زيادة معنوية في معاملات كاع ولي في هذه التجربة فعال بشكل معنوي في بعض صفات النمو .

الكلمات المفتاحية: PBZ ، Paclobutrazol ، سماد البوتاسيوم ، K2SO₄ ، بادرات نخلة التمر