

Green Synthesis of Silver Nanoparticles by *Enterobacter Aerogenes* Bacteria in Combination with Antibiotics Against Multidrug Resistance *Streptococcus Mitis* Isolated from Oral Cavity of Some Dental Caries Patients in Misan City.

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ABSTRACT

The present study aimed at the green, ecofriendly, cost-effective synthesis of silver nanoparticles (AgNPs) by using *Enterobacter aerogenes* bacterial isolated strains (S1, S2 and S3) from the oral cavity of dental caries patients in the center of Misan province, and also investigates their antibacterial effective combination with some traditional antibiotics against MDR *Streptococcus mitis* isolated from dental caries patients in the center of Misan province. The synthesized (AgNPs) nanoparticles were characterized by process of UV-Visible spectrophotometer Transform Fourier (FT-IR). The Scanning Electron Microscopy (SEM) image refers to a spherical shape with the size range of the nanoparticles (AgNPs) synthesized was 47.22 – 105.00 nm. The X-ray diffraction (XRD) patterns revealed the crystalline structure of nanoparticles was AgNPs. Transform Fourier Infrared Spectroscopy (FTIR) analysis refers to a functional group that is implicated with the reduction of silver ion to AgNPs. All of the clinical bacterial strains isolated from the oral cavity of dental caries patients exhibited resistance against conventional antibiotics. The synergistic effect of antibiotics combined with (AgNPs) mentioned an increasing percentage of antibiotics. However, the maximum of synergistic effect was done only with Erythromycin E 30.7mm for the AgNPs (0.1 mg/ml), when in contrast Streptomycin 40mm were Streptomycin with the AgNPs 0.1 mg/ml recorded 6.0mm, were Tetracycline 15.6mm has been recorded only 6mm inhibition zone of when combination with AgNPs 0.1 mg/ml, that refer to antagonistic effects. Whereas optochin OP, penicillin P, and cephalexin CN antibiotics recorded in different effects. Finally, with this study report. It can be concluded that AgNPs prepared from *Enterobacter aerogenes* showed good antimicrobial activity when combined with antibiotics against Multidrug resistance *Streptococcus mitis* isolates. Thus these AgNPs can be used as a wide application to treat many oral cavity diseases in the dental field.

Keywords: Green synthesis of silver nanoparticles combination with antibiotics, *Enterobacter aerogenes*, MDR *Streptococcus mitis* oral cavity dental caries.

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1. Introduction

At the beginning of 20th century the antibiotic penicillin was discovered, and since then humans have been working continuously to discover new antibiotics because bacteria at the same time, develop new mechanisms to resist these antibiotics. (Clatworthy *et al.*,2007). Although of these elevation a high accident rate to the rise of antibiotics resistance toward the traditional antibiotics is an important health care with globally serious impact (Mannaa *et al.*,2015). In recent years, nanotechnology as developing field of nanscience with important applications, including synthesis of nanomaterials and applications in medicine, biology due to their unique shape and size based on their chemical, biological and physical properties (Mahasneh,2013) Nanotechnology is considered the fourth industrial revolution through the history of human civil evolution Parthasarathi *et al* (2011), it deals with greater part of material types including ceramics, metals, polymers, and biomaterials. It has a great deal in elaboration and improvement of biogenic products such as certain medications like, antibiotics, sunscreens and sunblocks Vijay *et al.*, (2015). The green synthesis of nanoparticle metals has more favorable compared to chemical synthesis caused by use of eco-friendly deputy as reducing agents rather than hazardous of chemicals (Abdeen *et al.*,2014). Many studies refers to the evolution of nanoparticles products is distance presently, nanoparticles is establish in implant coatings, food containers, ointments and other items. US Food and Drug Administration have approval from it (Dunn & Edwards,2004). Different studies refers to the nanosilver bactericidal effective against gram positive and gram negative bacterial isolates (Morones *et al.*,2009).

2. Materials and Methods

2.1 Silver nitrate solutions

It's advised to prepare silver nitrate solution at real time. Silver nitrate was dissolved in double distilled water (DDW) in dark place to avoid photo-oxidation of silver nitrate. The solution was used as a substrate for silver nanoparticles biosynthesis (Chaudhari *et al.*, 2012).

2.2 Isolation and Identification of Bacteria:

Isolates samples were obtained from oral cavity of dental disease Dental Specialist Center of dental disease in Misan Governorate - Iraq. The isolates were diagnosed by Gram stain and biochemical tests. Finally by PCR with sequencing .These identified cultures were transferred to nutrient agar slant for preservation and then store in the refrigerator at 4°C.

2.3 Molecular Identification of Strains

One strain selected (S2) from three strains(S1 , S2 , S3) belonging to *Enterobacter aerogenes* bacterial isolated strains , were identified by 16S rDNA sequenced , extracted and DNA amplified using PCR, sequenced ,and aligned with other identified strains in Gene bank database by using BLAST tool an online to determine the similarity score .

2.4 Activation bacterial strains

Bacterial strains were activated in 10 ml of nutrient broth in serum tube by aseptic technique and incubated at 37°C overnight.

2.5 Biomass production

bacterial isolate (*Enterobacter aerogenes*) was inoculated in flask 500ml contained 6.5 gram/ml nutrient broth, incubated at 37°C in a shaking incubator at 200rpm for 48 h.

2.6 Harvesting the cell free medium

Supernatant of cell free was done through four steps: activation by inoculum , inoculation in the nutrient broth, culture of incubated for production biomass and centrifugation for cell excluding. Cell free supernatant was harvested by centrifugation of the broth culture pellet cells at bottom of the tubes leaving cell free supernatant. Supernatant then collected in a sterile flask for use in the production of silver nanoparticles (Singh *et al.*, 2018).

2.7Biogenic synthesis of silver nanoparticles:

cellular synthesis of silver Nanoparticles was accomplished by mixing 250ml of cell free supernatant from 48 hour liquid culture of *Enterobacter aerogenes* strain (filtered through 0.2µm pore dimension) with 250ml of 1mM silver nitrate (AgNo₃) solution. Then incubated in orbital shaker (200rpm) at 37°C in dark for 5 days. A flask with cell free supernatant without (AgNo₃) utilized as control (Saifudin *et al.*2009).

2.8 Characterization of biosynthesis(AgNPs) .

Biosynthesis characterization of (AgNPs) was performed by using UV-visible spectroscopy, The purified AgNPs were examined for the presence of biomolecules using FTIR spectrum (Thermo Scientific Nicolet 380 FT-IR Spectrometer X-ray diffraction(XRD) and scanning electron microscope (SEM) \ , particle sizes (Zhang *et al.*, 2016).

2.9 Antimicrobial activity test

The antimicrobial activity test was evaluated by two (well and disk diffusion methods). Johan *et al.* (2003).

2.10.1 Combination assay of antibiotics and AgNPs

Assays were performed in susceptibility of antibiotic with AgNPs it is Combination assay, the cells of bacterial were grown in nutrient broth .Disk-diffusion method was performed to assess the combination between antibiotics and AgNPs biologically synthesized with their antimicrobial activity against some bacterial isolates from dental caries and periodontitis. Nutrient broth were cultured with bacterial isolates *Streptococcus mitis* on Muller – Hinton lates (Johnson, 2010).

2.10.2 MIC Determination.

The synergy effects between antibiotics drugs usually expressed by sum of Fractional Inhibitory Concentration (FIC) of two combinations . FIC is calculated by following for combination A , B:

$$\text{FIC} = \frac{\text{MIC of antibacterial A in combination}}{\text{MIC of antibacterial A alone}}$$

FIC was also calculated for combination B with same formula

Thus [FIC= FIC of antibacterial A+ FIC of antibacterial B

3.Results and Discussion

3.1 Morphological and biochemical characterization.

In the present study the clinical bacterial strains isolates from oral cavity of dental disease patients were obtained from center of Misan City- Iraq Identification of the isolate was done based upon exploring the role of bacterial strain (S2) *Enterobacter aerogenes* in

synthesize AgNPs along with their biomedical application to controlling infectious bacteria , as shown in [Figure. 1,2].

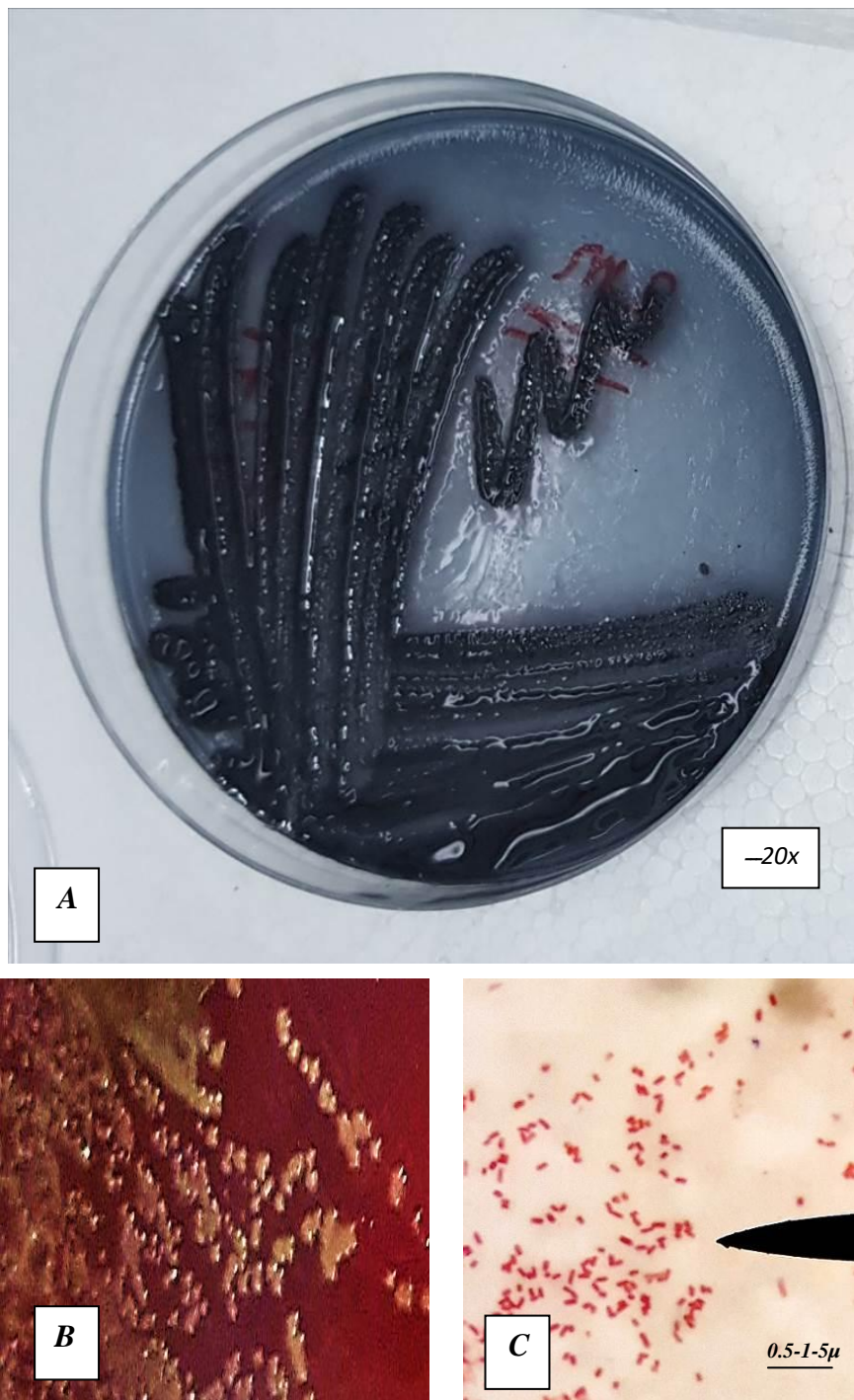
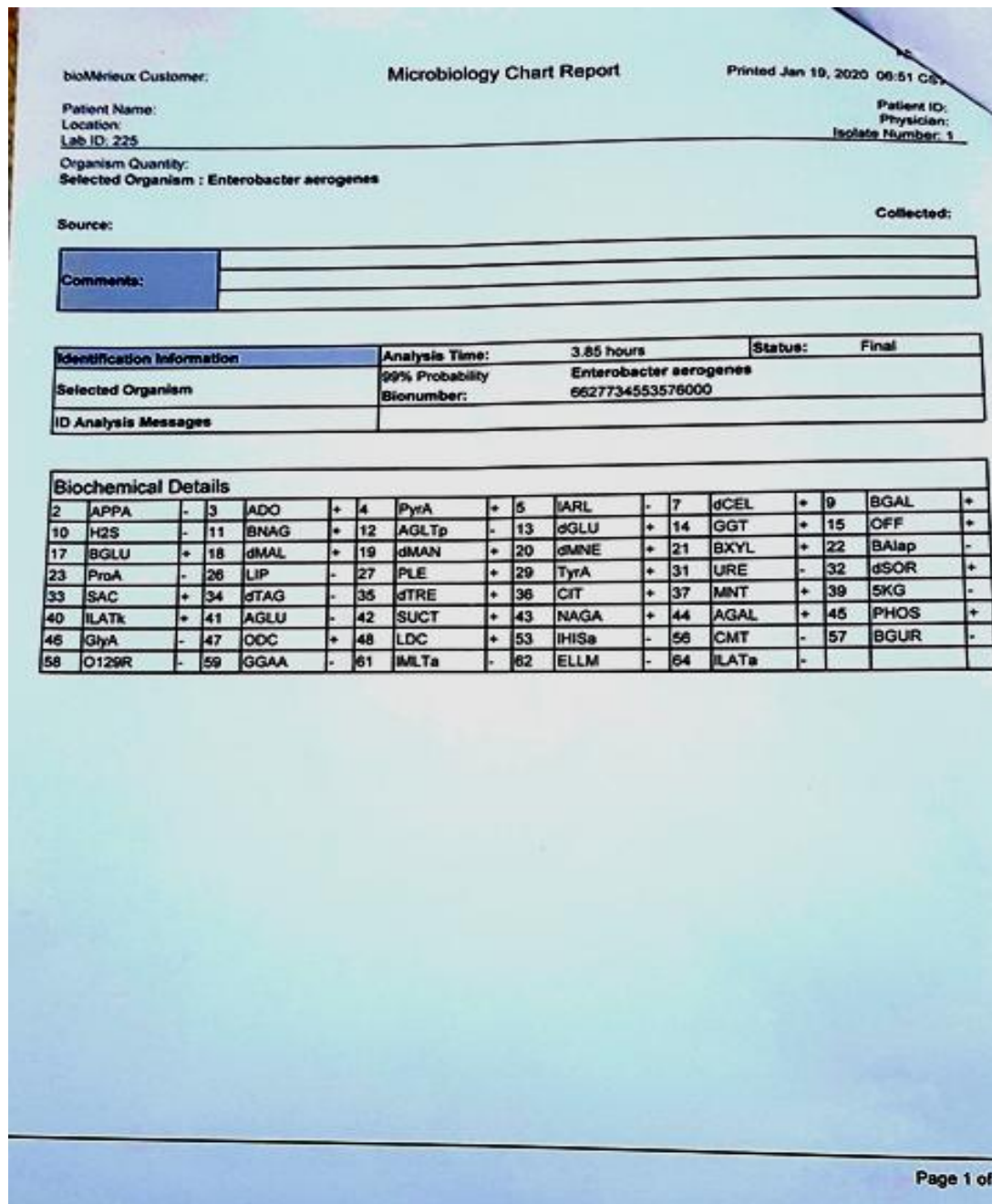


Figure (1): Shown bacterial strain of Enterobacter aerogenes cultured on (A) Mitis Salivorous Agar and(B) Blood Agar (C) Gram stain of bacterial isolate of Enterobacter aerogenes shown by microscopy 1250x

Figure (2): Shown Vitek2 Chart Report Microbiology for Identification of *Enterobacter aerogenes*



3.2 Molecular Identification

Characterization of AgNPs by bacterial strain.

Identification was done in addition to biochemical test, isolates was matching through 16SrRNA gene sequencing , molecular identification by extraction and amplification of gene

[Figure 3]. One strain selected (S2) from three strains(S1 , S2 , S3) belonging to *Enterobacter aerogenes* bacterial isolated strains , were identified by 16S rDNA sequenced of strain IHB B 6843 was 1290bp , extracted and DNA amplified using PCR, On the basis of analysis of the 16S rRNA strain IHB B 6843 showed the highest sequence similarity with NR_117547.1 *Enterobacter soli* ATCC BAA.2102 strain LF7.[Figure 4,Table1] sequenced ,and aligned with other identified strains in Gene bank database by using BLAST tool an online to determine the similarity score .

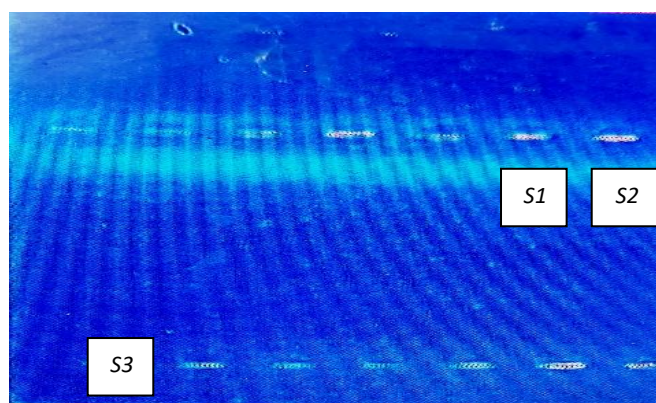


Figure (3) : Agarose gel electrophoresis patterns show of genomic DNA extracted from the bacterial strains (S1,S2and S3) For *Enterobacter aerogenes* on 1% Agarose gel at 5vol.cm for 1.15 hour. redsafe stained and documented UV gel viewer.

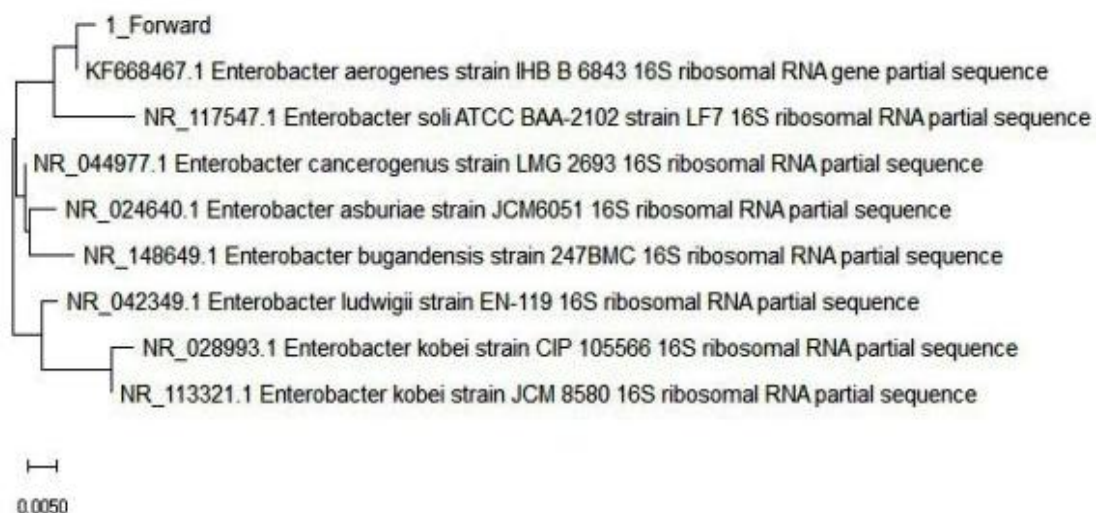


Figure (4): The phylogenetic tree relationships of isolated strain IHB B 6843 with related type strains. The scale par shows 0.005 substitutions per nucleotide position.

Table (1): Shown identical bacteria by 16SrRNA gene sequences, source and identical strains(%).

Sequence description	Sequence	identification	Percent identity
1_Forward	ATGGATGCGGCAGCTACACATG CAAGTCGAGCGGTAACACAGAG AGCTTGCTCTCGGGTGACGAGC GGCGGACGGGTGAGTAATGTCT GGGAACTGCCTGATGGAGGGG GATAACTACTGGAAACGGTAGC TAATACCGCATAACGTCGCAAG ACCAAAGTGGGGGACCTTCGGG CCTCATGCCATCAGATGTGCCCA GATGGGATTAGCTAGTAGGTGG GGTAATGGCTCACCTAGGCGAC GATCCCTAGCTGGTCTGAGAGG ATGACCAGCCACACTGGAAGT AGACACGGTCCAGACTCCTACG GGAGGCAGCAGTGGGGAATATT GCACAATGGGCGCAAGCCTGAT GCAGCCATGCCGCGTGTATGAA GAAGGCCTTCGGGTTGTAAAGT ACTTTCAGCGAGGAGGAAGGCG TTAAGGTTAATAACCTTGGTGAT TGACGTTACTCGCAGAAGAAGC ACCGGCTAACTCCGTGCCAGCA GCCGCGGTAATACGGAGGGTGC AAGCGTTAATCGGAATTACTGG GCGTAAAGCGCACGCAGGCGGT CTGTCAAGTCGGATGTGAAATC CCCGGGCTCAACCTGGGAACTG CATTCGAACTGGCAGGCTAGA GTCTTGTAGAGGGGGGTAGAAT TCCAGGTGTAGCGGTGAAATGC GTAGAGATCTGGAGGAATACCG GTGGCGAAAGCGGCCCCCTGGA CAAAGACTGACGCTCAGGTGCG AAAGCGTGGGGAGCAAACAGGA TTAGATACCCTGGTAGTCCACGC CGTAAACGATGTCGACTTGGAG GTTGTGCCCTTGAGGCGTGGCTT CCGGAGCTAACGCGTTAAGTCG ACCGCCTGGGGAGTACGGCCGC AGGGTTAAACTCAAATGAATT GACGGGGGCCCGCACAAAGCGGT GGAGCATGTGGTTTAATTCGATG CAACGCGAAGAACCTTACCTAC TCTTGACATCCAGAGAACTTAGC	Enterobacter aerogenes strain IHB B 6843 16S ribosomal RNA gene, partial sequence	99.79

	AGAGATGCTTTGGTGCCTTCGGG AACTCTTAGAACAGGTGCTGCA TGGCTGTCGTCCACCTCCGGTTG TGAAAAGTTGGGTAAATTCCCGC ACCCAGCGCCACCCCTTATTCCT TTGTGTGCCGCCGATTTTGTGCG GGAACCCCAAAGGGAAAATGCC CCTTTAAAAAACGGAAGAAAAG GGGGGGGGGACGCTCCACTTT CCCTCGGCC		
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3.3 Characterization of AgNPs

Harvesting the cell free medium

Cell free supernatant was harvested by centrifugation of the broth culture pellet cells at bottom of the tubes leaving cell free supernatant. Supernatant then collected in a sterile flask for use in the production of silver nanoparticles (Singh *et al.*, 2018). cellular synthesis of silver Nanoparticles was accomplished by mixing 250ml of cell free supernatant from 48 hour liquid culture of *Enterobacter aerogenes* strain (filtered through 0.2µm pore dimension) with 250ml of 1mM silver nitrate (AgNo₃) solution. Then supernatant mixture incubated in orbital shaker (200rpm) at 37°C in dark for 5 days. A flask with cell free supernatant without (AgNo₃) utilized as control (Saifudin *et al.*2009).

Because of increasing multi drug resistance (MDR) threat, we scope on search the efficient alternative sources to replace resisted antibiotics .Nanoparticles considered now as alternative to the traditional antibiotics .Recently , nanoparticles (AgNPs) were considered as attractive target for the fabrication of a new antibiotics generation (Rai *et al.*,2012 ;Singh *et al.*,2017) . The AgNPs of bacterial strain (S2) *Enterobacter aerogenes* showed effective antimicrobial activity against multi drug resistant bacterial isolate *Streptococcus mitis* [Figure 5 and 6],

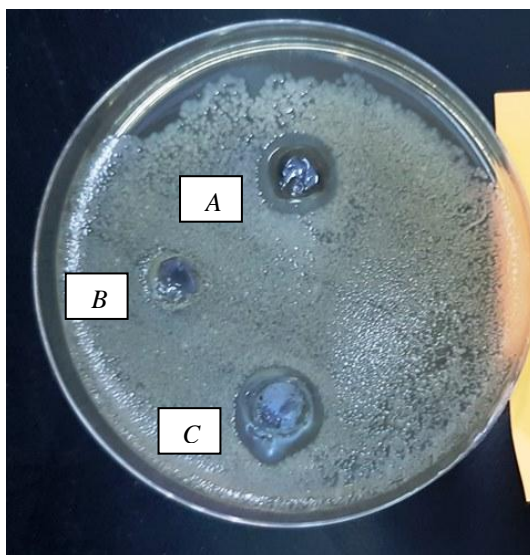
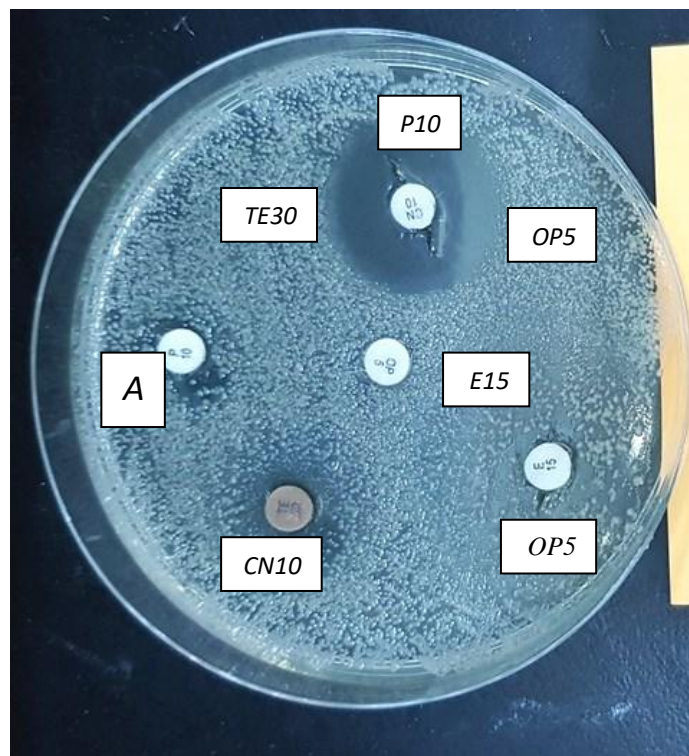


Figure (5): A. Effect of Nanoparticles on MDR *Streptococcus mitis* isolated from dental caries patients B. Control DDW C. Antibacterial activity of AgNo₃.

3.4 Antimicrobial activity:

Antimicrobial activity was done by disk diffusion method was used to evaluate Optochin OP , Penicillin P , Tetracycline TE ,Cefalexin CN , Streptomycin S antibiotics alone and in combination with Biogenic AgNPs synthesized by *Enterobacter aerogenes* have been evaluated for their antimicrobial activity against some pathogenic bacteria isolated from dental carries patients in misan province / Iraq , using agar well and disk diffusion method [Figure7 and 8] ,[Table1]. According to CLSI Standard , selective concentration of antibiotics .Selective concentration of AgNPs was 100µg. Its found that, at concentration of 100µg.The antibacterial activity of combined formulation of AgNPs through different discs of antibiotic , including optochin ,penicillin , tetracycline ,erythromycin ,cephalexin and streptomycin [Figure 8]. all of the clinical bacterial strains isolates from oral cavity of dental disease patients were obtained from center of Misan City-Iraq, exhibited resistance against conventional antibiotics such as Optochin OP ,Penicillin P, Erythromycin E , were bacterial isolate *Streptococcus mitis* showed more sensitively against Tetracycline TE ,Cefalexin CN , Streptomycin S. The results in present study showed only (Erythromycin E 30.7mm) as a synergistic effect of antibiotics , the combination with AgNPs mentioned increasing efficacies (percentage) of antibiotics. In similar with Fayaz et al (2010) the antimicrobial activities of synthesized of AgNPs were estimated with available antibiotics against Multi drug resistant *Streptococcus mitis* strains. However, maximum of synergistic

effect was done only with Erythromycin E 30.7mm for the AgNPs(0.1mg/ ml) .When in contrast Streptomycin S (Streptomycin 40mm were Streptomycin with AgNPs 6.0 mm) and Tetracycline TE (Tetracycline 15.6mm were were Tetracycline with AgNPs 6mm inhibition zone recorded) Antagonistic effects. Where Optochin OP , Penicillin P and Cefalexin CN antibiotics Indifferent effects [Figure 6 , Table2 and 3] .These results are in a similar with Birla *et al* (2009) the biologically synthesized AgNPs assessed with available antibiotics against both gram negative and gram positive bacteria. The antimicrobial activities of Erythromycin, ampicillin , kanamycin ,and chloramphenicol were extend in the presence of nanoparticles AgNPs. Finally, The minimum concentration of inhibition zone recorded with the concentration of the following (0.1 , 0.05 , 0.025 , 0.0125 and 0.0062 mg / ml) of the biosynthesized AgNPs detection for *Enterobacter aeruginose* [Figure 6 ,Table 4] .we findings that the action of mode of silver nanoparticles is same as, to that of silver ions, which interaction with groups of electron donor containing oxygen ,sulfur or nitrogen atoms that are usually present as phosphates or thiols on amino acids and nucleic acids (McDonnell,2007).The(Kvittek et al.,2008) explain that AgNPs interaction with the surface of membrane of the bacterial cell by interacting with proteins containing sulfur , that disrupting of the respiratory functions of the permeability of the cell membrane leading to cell death.



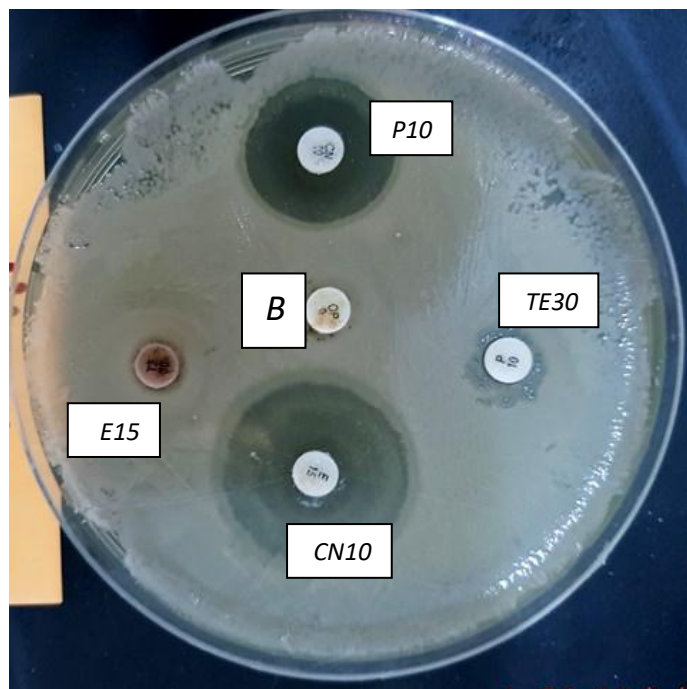


Table (2) : Percentage of inhibition zone by antibiotics alone and nanoparticles AgNPs combination with antibiotics against *Streptococcus mitis*

No	Antibiotics	Concentration µg/disk	Antibiotics Effects		Increase in fold area
			A	B	
1	Optochin OP	5	6	7.3	21.66
2	Penicillin P	10	6.6	9.7	46.96
3	Tetracycline TE	30	15.6	6.0	-61.53
4	Erythromycin E	15	7.3	30.7	320
5	Cefalexin CN	30	26.3	20.6	-21.67
6	Streptomycin S	10	40.6	6.0	-85.22

+Mean of the inhibition zone (mm) for each antibiotic tested of the inhibition zone was done by mean diameter fold increases for diverse antibiotics studied as $((b-a) / a) * 100$ (Zarina & Nanda, 2014). * Three replicates for each value.

No	AgNPs+Antibiotics	Interaction
1	Optochin + AgNPs	Indifferent
2	Penicillin + AgNPs	Synergistic
3	Tetracycline+ AgNPs	Antagonistic
4	Erythromycin+ AgNPs	Synergistic
5	Cefalexin+ AgNPs	Indifferent
6	Streptomycin+ AgNPs	Antagonistic

Table(3): Index of FIC to establishes the interaction between antibiotics agents

[FIC index to use for identify of th interaction between the two antibacterial combinations . The index is interpreted as follows : $1 \geq$ synergistic , $0.2 \leq$ antagonistic , $1 =$ additive ; $1.1-2.0$ indifferent (non-interactive) Habiba *et al.*,(2015).

Table (4): Shown minimum of inhibition zone concentration (MIC) of the biosynthesized AgNPs alone and with combination of six Antibiotics against Multi drug *Streptococcus mitis* bacterial isolated from oral cavity patients in misan city.

No	Multi drug <i>Streptococcus miti</i>	
	MIC (mg/ml)	
1	NPs alone	0.009
2	Optochin + AgNPs	-
3	Penicillin + AgNPs	0.037
4	Tetracycline+ AgNPs	0.018
5	Erythromycin+ AgNPs	0.018
6	Cefalexin+ AgNPs	-
7	Streptomycin+ AgNPs	0.037

* Three replicates for each value

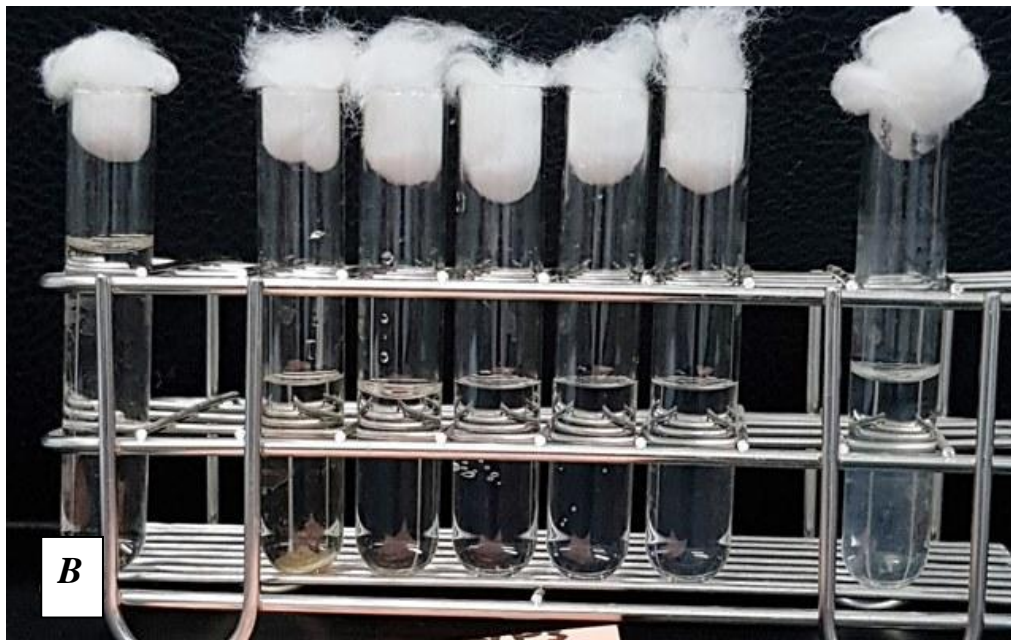
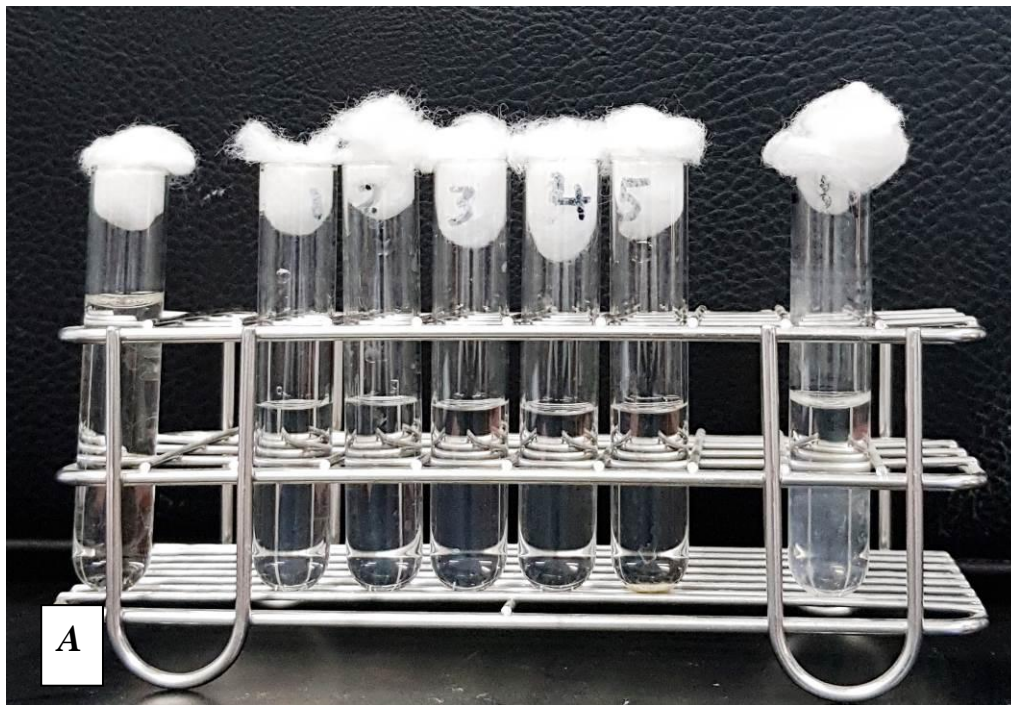


Figure (7) : Shown Minimum inhibitory concentration (MIC $\mu\text{g}/\text{ml}$) of NPs alone (A) ,
Minimum inhibitory concentration (MIC $\mu\text{g}/\text{ml}$) of NPs with antibiotic (B).

3.5 Physical characterization of synthesized nanoparticles AgNPs

Nanoparticle AgNPs is composed by (S2 strain) *Enterobacter aerogenes* was characterized visually by UV-Visible spectra analysis [Figure 14], FT-IR

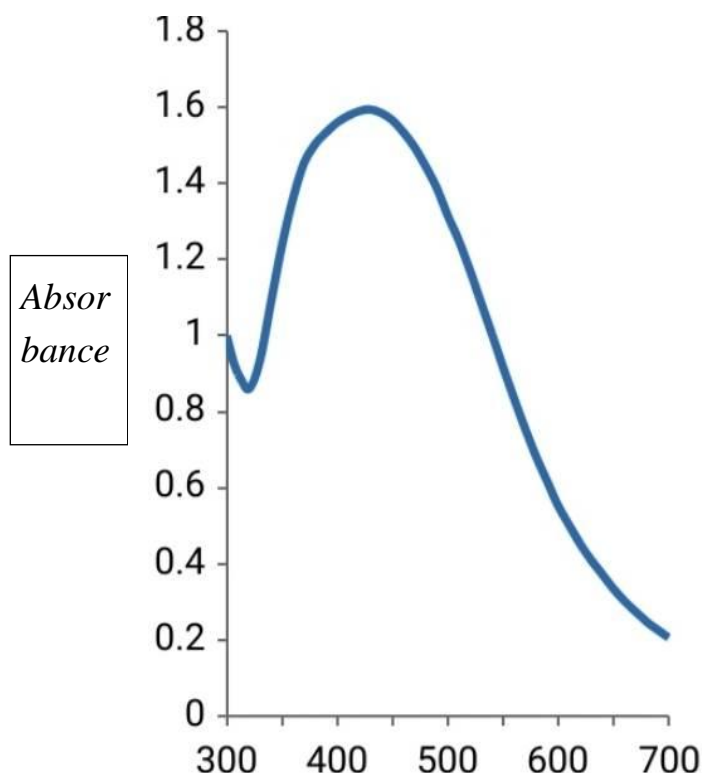
[Figure 15] and XRD assays [Figure 16, Table 7]. The initial formation of the silver nanoparticles AgNPs is mediated by the observation the colour change take place in the reaction solution from light yellowish brown to dark brown due to the excitation vibrations of surface plasmon of the incorporated AgNPs. The colour change of solution can be considered clearly indication of silver nanoparticles formation (Kalimuthu *et al.*, 2008; Luo *et al.*, 2018).



Figure (8) : Extracellular of AgNPs synthesis at start of the reaction and after 72 hours (end of reaction). A. Positive control filtrate without AgNO₃ B. Culture filtrate *Enterobacter aerogenes* with AgNO₃ solution (0.1mM). C. After 96 hours extracellular of AgNPs synthesis, dark brown solution that's detected the completion of reaction and D. shown Nanoparticles AgNPs after 24 h in petridish.

This ranged in color also confirmed strength absorbance also by UV-Vis spectroscopy. The optical peaks absorption of the reaction mixture were nearly 420nm[figure6],this result supported by the researcher Basavaraja *et al.*,(2008) they were reported that the reduction of Ag^+ to atomic Ag^0 in agreement with absorption at 420nm [Figure.14].

Figure (9) : UV- Vis absorption spectrum of AgNPs synthesized by Enterobacter aerogenes isolated from dental caries patient .



The FT-IR measurements were carried out to reveal the possible biomolecules that responsible for the stabilization of the synthesized AgNPs (Ananthi ,2018).The FT-IR spectrum [Figure 15] illustrated peaks at 3448 , 3426, 2928, 2881 ,1651 ,1392 and1111,1045 cm^{-1} , the absorbance band at 3475-3383 cm^{-1} assigned to the stretching vibrations of hydroxyl groups and amine groups, where N-H was recognized as stretching vibrations in primary and secondary amines of proteins , peptides and amino acids (Cheng *et al.*,2014).The peak of 2928 ,2958 cm^{-1} indicated for secondary amine (Vigneshwaran *et al.*, 2007) , while peak at 1647 cm^{-1} shown to be responsible for the capping of silver ion and forming consequently covering protein around AgNPs to thereby stabilize medium with prevent agglomeration (Shanmuganathan *et al.*,2017).The peaks positioned at 1539,1384 and 1053 cm^{-1} assigned to either function groups of sulfur or phosphorus , which attributed to

possibly of stabilizing and capping process of AgNPs (Castro *et al.*,2013).The proteins can interactions with nanoparticles through cysteine or free amine groups residues in the proteins

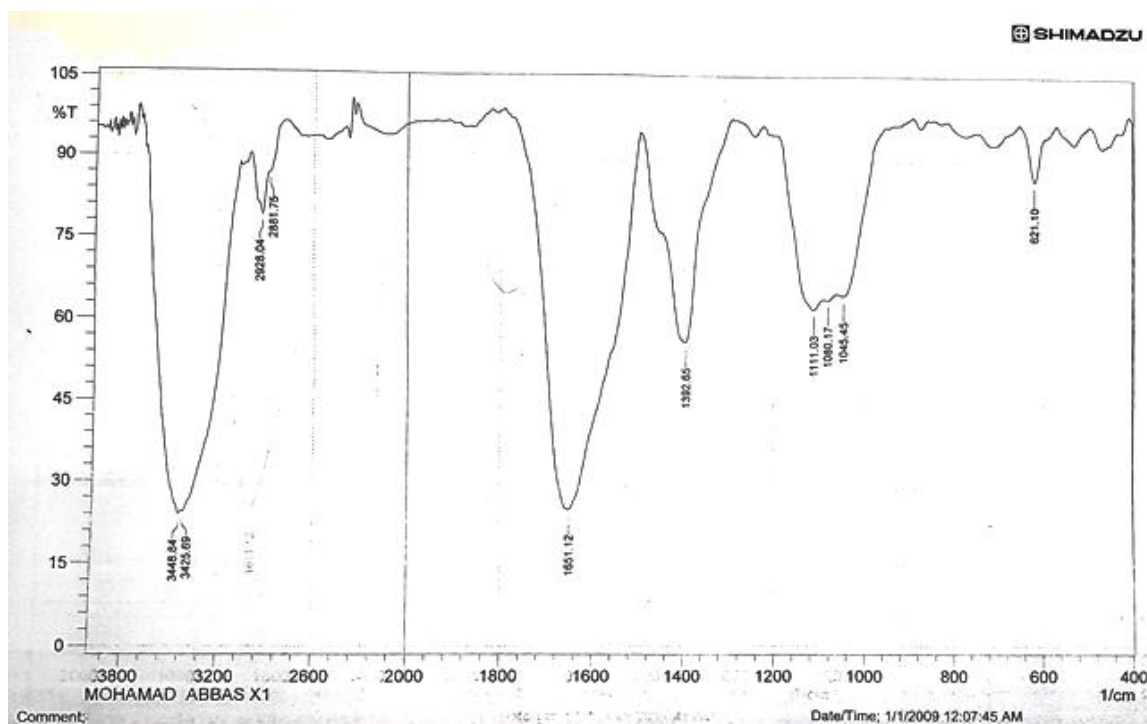


Figure (10) : FT- IR spectrum of AgNPs synthesized by *Enterobacter aerogenes* isolated from dental caries patient .

(Mandal *et al.*,2013).

The Deby – Scherrer formula can be using the mean size of the AgNPs crystallized particles (Allafchian and Jalali,2015). XRD patterns refer to four major peaks in the entire spectrum of (2θ) 38.42 , 46,51, 67,76 and 77.09 . These values refer to the (111) , (200) , (220) and (311) planes of face –centered –cubic(FCC) silver , with a network parameter of 4.08 °A, which is consistent with the Joint Committee Powder Diffraction Standards (JCPDS) Card No-087-0720 (Vanaja et al.,2014). Scanning electron microscope image showed various shapes of nanoparticles ; however , the shapes were spherical with ranged size 71.56 nm[Figure16 and Table 6 with7].

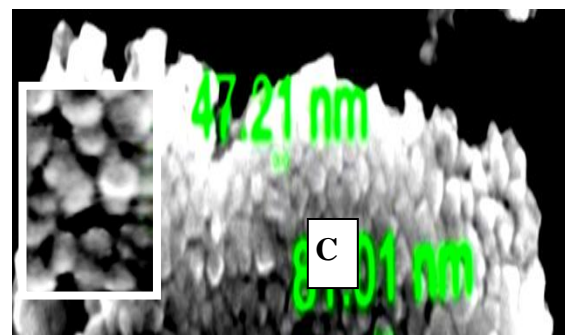
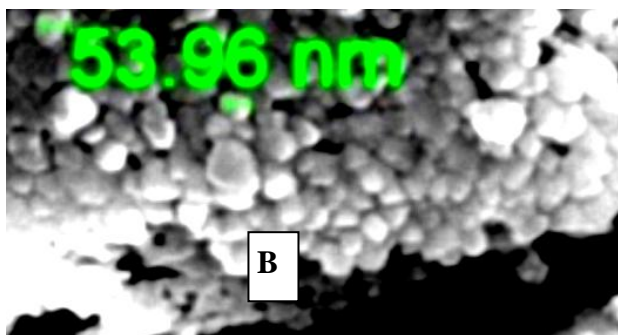
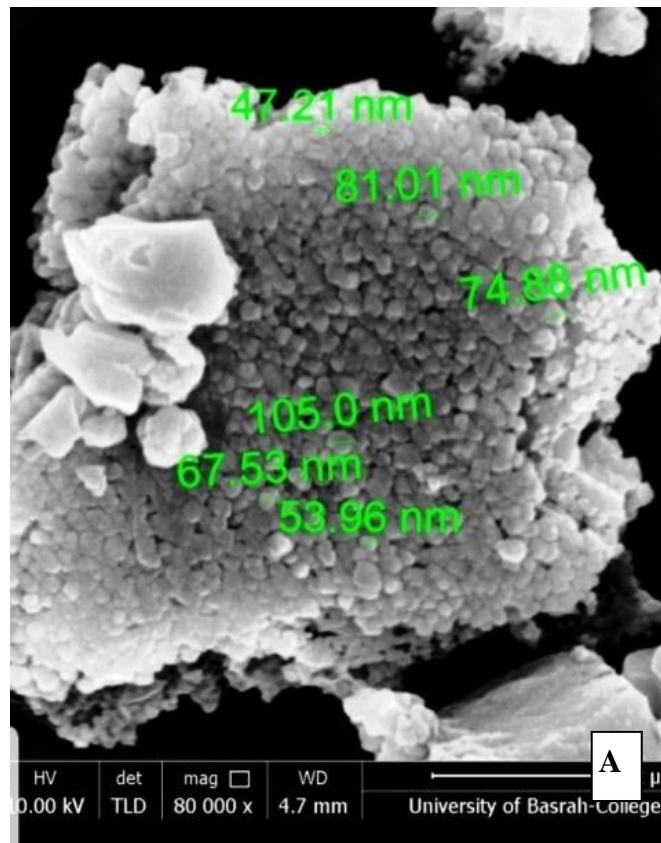


Figure (11): Chacterization of the biogenic AgNPs synthesized by *Enterobacter aerogenes* (A ,B,and C) Scanning electron microscopy .

Table (5):Lattice plane resulted AgNPs source by *Enterobacter aerogenes* as indexed to JCPDS data .

Source of AgNPs	h kl index			
	111	200	220	311

<i>Enterobacter aerogenes</i>	38.4213	46.5167	67.7625	77.0906
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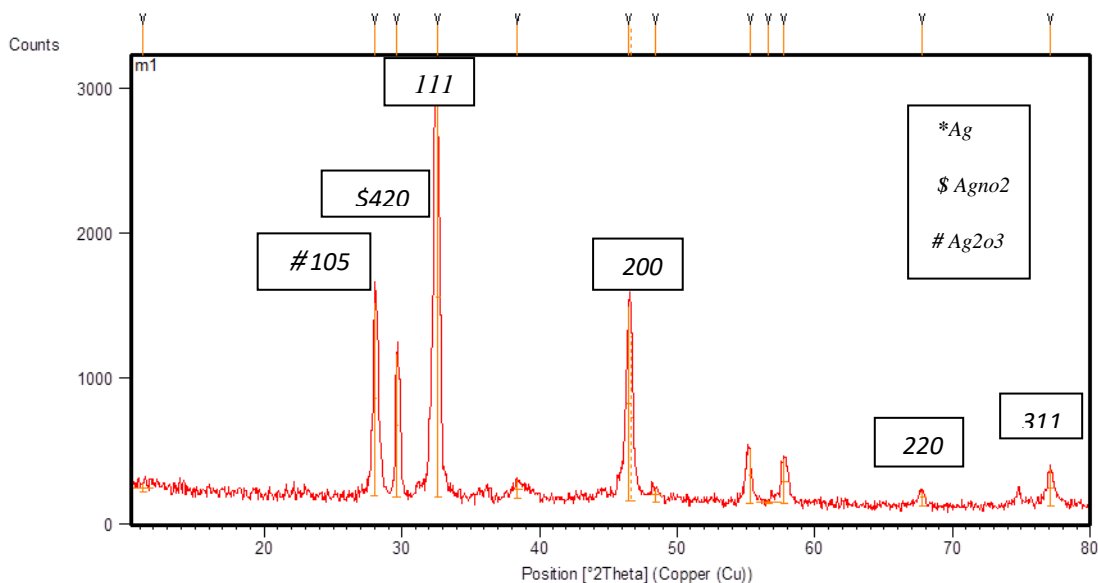


Figure (12) : XRD report ,resulted by AgNPs fabricated on copper surface obtained from *Enterobacter aerogenes* isolated from oral cavity of dental caries patients in center of misan province .

Table (6).XRD results of silver nanoparticles

Pos. [°2Th.]	Height [cts]	FWHM Left [°2Th.]	d-spacing [Å]	Rel. Int. [%]	Tip Width
38.4213	132.40	0.5904	2.34297	4.80	0.7085
46.5167	1340.05	0.3000	1.95073	48.56	0.3600
67.7625	108.62	0.4800	1.38177	3.94	0.5760
77.0906	255.07	0.4800	1.23617	9.24	0.5760

Results of this study suggest that the nanoparticles AgNPs of *Enterobacter aerogenes* may be useful either alone or when combined with antibiotics against oral pathogens MDR *Streptococcus mitis* bacterial isolates , which can be used as a wide application to treat many of oral cavity diseases in dental field.

5.Conclusion

The results of present study showed synergistic effect of combination of Erythromycin antibiotic with AgNPs nanoparticles against *Streptococcus mitis* isolates from some dental

caries patients . Some hypotheses were explain of these combinations mechanism between AgNPs nanoparticles with antibiotics. Finally, detailed is needed to evidence clarify mechanism of synergistic effect.

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