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First record of ciliophora (Orchitophryidae) isolated from CSF samples of patients with meningitis in Thi-Qar province south of Iraq

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ABSTRACT

In September 2020 when we were investigating about opportunistic free living amoeba in patients with meningitis in Thi –Qar province south of Iraq, throughout examination the CSF which was cultured on NN Agar medium. It was surprisingly we found the ciliates protozoa (Orchitophryidae) in both stages, vege-tative and cystic, after that the ciliates were isolated and cultured in vitro and then morphological, molecular and virulence characterizations were studies. The results of morphological study revealed that these ciliates were similar to *Mesanophrys* sp. whereas the sequence of rDNA of studied ciliate with accession numbers being given. in NCBI LC621170.1 was show (86.04%) homology identity to *Mesanophrys* sp. 18S rRNA gene (accession number MN260367.1). but show (98.56%) and (97.92%) homology identity to Orchitophrydiae 18S rRNA gene (accession number EF023919.1 and EF023549.1), that may be this ciliates in our study is a new species for *Mesanophrys*. In addition experimental infection confirmed that *Mesanophrys* like ciliates was the pathogen that infected human. In summary this study represent the first record of the ciliophora Orchitophryidae (*Mesanophrys* like ciliates) as a human pathogen from Thi-Qar province / Iraq.

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1. Introduction

Orchitophryidae ciliates are belonging to the class : oligohymenophorea, subclass : scuticociliatia and order : philasterida, they are tiny ciliates and widely distributed in marine environments . This family have four genera including : *Metanophrya, Paranophrys, Orchitophrya and Mesanophrys* [1]. The morphology of *Orchitophrya* ciliates was similar to *Mesanophrys* ciliates in some cases , which often causes difficulty in classification [2]) . *Mesanophrys* ciliates can live freely in sea water or sediment invade the haemolymph of crustaceans and cause the death of blue crab [3].

Systemic infection of ciliates parasites have been reported from many organisms such as fish (Munday *et al.* 1997; Iglesias *et al.*, 2001),sea stars[4,5], bivalve molluscs[6,7,8]) and farmed swimming crab [9]).

The belief prevailed that *Balantidium coli* is the only member of the ciliate phylum known to be pathogenic to human but recently many cases of infections in human with ciliates were recorded, in

* Corresponding author. *E-mail address:* bassadalaboody_bio@sci.utq.edu.iq (B.A. AL-Aboody). the urine of patient with chronic prostatitis, renal microlithiasis and acute cystitis were found the ciliate protozoa Colpoda spp. [10],(] B. and first recorded of the ciliophoran *Trichophrya piscium* as a human pathogen from Basrah, Iraq [11],(B. Al Hayani 2018).

The present finding documents the first record of ciliophora (Orchitophryidae) from CSF samples of meningitis patients, a systemic parasitic ciliate infection was discovered in two individual human with meningitis during investigations into opportunistic free living amoebae in Thi-Qar province ,one case from a female patient 5 months old and the second case from a male patient 5 years old),(B.

2. Material and methods

2.1. Sample collection

One ml of CSF collected from eight patients there age ranged between 5 month to 10 years they were suspected to be infected with meningitis but showed a negative bacterial cultured in laboratory of bacteriology in Bint AL-Huda teaching hospital and Ibn AL-Btar private laboratories in Thi-Qar province south of Iraq,

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the CSF was cultured on non– nutrient agar medium(NN-agar) and incubated in 26 C^0 and 37 C^0 and followed up daily examination of wet a mount slide for four week . After four days ciliates were found in the culture with the same shape in two samples only.

2.2. Ciliate isolation and cultivation

The ciliates with the same shape were discovered in to CSF culture by microscopic examination , in order to establish a monoclonal culture the ciliate isolated from CSF culture and cultured in NN-agar medium added to it Page amoeba saline (PAS) incubated in $4C^0$, $20C^0$ and $37C^0$ to determined the optimum temperature for growth),(B. .

2.3. Morphological observation

The cultured ciliates were observed and photographed using imaging microscope .

2.4. Experimental infection

3. Preparation of ciliate for injection

Ciliate trophozoites were scraped from agar surface by a cotton swab after 2–3 days and then the liquid phase of the culture was

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transported to sterile plain tubes and centrifuged at 5000 rpm for one minute . The supernatant was discarded and the ciliates were washed twice with 1 ml of sterile PBS and finally resuspended in 1 ml of sterile PBS, the number of ciliate trophozoites were calculated by haemocytometer),(] B. .

4. Infection of animals

4.1. Animals

12 male of white mice (Mus musculus /Bulb / c) were divided in to three groups, four mice in each group weighting approximately 25–35 gm , the animals were bred in special cages in the animals house of the college of science / Thi- Qar university),(B. .

The animals injected as following :

1-First group : A wound on the left femur was made using a sterile blade , the wound was contamination with ciliate trophozotes (10^5 per animal) by a sterile syringe .

2- Second group : It was injected intraperitoneal with ciliate trophozotes (10⁵ per animal).

3- Third group : Injected with normal saline as negative control

After 27 days of infection , mice were killed by ether ,part of brain cultivation on NN-agar medium at $20 C^0$. The remaining part of mice brain and liver were fixed in 10% formalin for histological study .





(B)



(C)

Fig. 1. Images of a live ciliate from an in vitro culture (unstained)100X : (A & B) trophozoite stage note the contractile vacuole (red arrow) and the granular appearance of the cytoplasm : (C) encystations (blue arrow) and cyst stage(whit arrow). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

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4.2. Tissue preparation and histology :

Tissue preparation was made according to Drury et al. [12] : A small piece of the targeting organs (brain and liver) of the infected mice was kept in 70% ethanol till tissue preparation.

5. DNA isolation and sequencing

Genomic DNA (gDNA) of ciliates in CSF culture , brain and liver tissues of infected mice was extracted by using gSYNC TM DNA Extraction kit , Geneaid . Korea , and done according to company instructions . Conventional PCR was done for 18S rRNA using a set specific two primers designed by Dopheide et al. [13] : 384F (YTB GAT GGT AGT GTA TTG GA) 1147R (GAC GGT ATC TRA TCG TCT TT) of common ciliophora. according to the following protocol :

Initial denaturation $95C^0$ for 10 min and 35 cycle of 35 sec at $95C^0$, 35 sec at $55 C^0$ and 40 sec at 72 C^0 followed by 10 min final extension at 72 C^0 , PCR product was electrophoresed on 1.5% agarose gel and visualized by UV, the PCR products (750 bps) were sent to Macrogene company (South Korea) for sequencing. The sequences were analyzed by BLAST comparison with previously



Fig. 2. Agarose gel electrophoresis image that show the PCR product(750 bps)t analysis of 18S ribosomal RNA gene from genomic DNA of ciliophora from clinical samples : Where M: Marker (2000–100 bp) lance (1,2) positive samples of CSF culture and lance (4,5,) positive samples of infected brain tissue (6) positive sample of infected liver tissue .

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published sequences which were derived from the GenBank database.

6. Results

6.1. Ciliate morphology

Two phases (trophozoite and cyst) of ciliates were observed in the culture. .Trophozoite was melon – shaped with a tapered anterior end and a rounded posterior ,it has long cilia , live ciliates were very active and flexible continually moving forward and stopping to change direction . They have a contractile vacuole at the posterior end and an oral apparatus at the anterior end , ciliate cell were between 40 and 55 μ m in length and 12.5–15 μ m in width with one macronucleus and one micronucleus ciliate had a densely ciliated oral area . The cyst of the ciliate was spherical with thick cyst wall , the mean diameter of cyst was 22.5–25 μ m . Fig. 1.

6.2. PCR and sequences analysis

The conventional PCR using the common ciliophora two primers (384F & 1147R) that amplified a portion of 18S rRNA gene yield a 750 bp product was positive for ciliophora in CSF culture , brain tissue and liver tissue of infected mice Fig. 2. Following sequencing and analysis of the CSF culture and brain tissue PCR products Blast results indicated that the sequence of rDNA of ciliate with accession numbers being given in NCBI LC621170.1 was show (86.04%) homology identity to *Mesanophrys* sp. 18S rRNA gene (accession number MN260367.1). but show (98.56%) and (97.92%) homology identity to Orchitophrydiae 18S rRNA gene (accession number EF023919.1 and EF023549.1). Fig. 3

6.3. Experimental ecology of temperature

The results showed that ciliate . could grow normally in the temperature of 20 C^0 and 37 C^0 , ciliates entered the exponential growth period on the 3rd day reached the maximum density on the 7th day and it can remain active on the NN-agar until the 20th day after cultivation.

Score		Expect	Identities Gaps Strand 0 613/622(99%) 0/622(0%) Plus/Plus CGGAGAGGGAGCCTGAGAAATGGCTACCACATCTAAGGAAGG		
1099 b	its(59	5) 0.0	613/622(99%)	0/622(0%)	Plus/Plus
uerv	1	GGTTCGATTCCGGA	GAGGGAGCCTGAGAAA	GGCTACCACATCTAAG	GAAGGCAGCAGGC
bjct	365				
uery	61	GCGTAAATTACCCA	ATCCTAATTCAGGGAGG	TAGTGACAAGAAATAA	CAACTCGGACCTC
bjct	425				
uery	121	ACACGAGGTTACGA	GATTGCAATGAGAACAA	TTTAAACCTCTTATCG	AGTAACAATTGGA
bjct	485				
uery	181	GGGCAAGTCTGGTG	CCAGCAGCCGCGGTAAT	TCCAGCTCCAATAGCG	TATATTAAAGTTG
bjct	545				
uery	241	TTGCAGTTAAAAAG	CTCGTAGTTGAAAATCT	GGCCGGTGCTACTCTC	GGCTCCCTGAGTC
bjet	605		T	GTT	
uery	301	GGTGTAGCGCCTGG	TCATCCGTACGGGAAAA	CTAGCTCGACCTTCACT	GGTCGGCTAGTGG
bjet	665	CTC	·····T·····		
uery	361	ATCGTACACTTTAC	TTTGAAAAAATTAGAGI	GTTTCAGGCAGGCAAT	TGCTTGGATACTG
bjet	725	A			
uery	421	TAGCATGGAATAAT	GGAATAGGACTTTGAC	CTATTTGTTGGTTTCTC	GAGGTCAAAGTAA
bjct	785		••••••	• • • • • • • • • • • • • • • • • • • •	
uery	481	TGATTAATAGGGAC	AGTTGGGGGGCATTCGT	ATTTAATTGTCAGAGGT	GAAATTCTTGGAT
bjct	845				
uery	541	TTTTTAAAGACGAA	CTTATGCGAAAGCATTI	GCCAAGGATGTTTTCA	TTAATCAAGAACG
bjct	905				
uery	601	AAAGTTAGGGGATC	AAAGACGa 622		
bjct	965				

Fig. 3. NCBI BLAST Multiple sequence alignment analysis that show single nucleotide in Orchitophrydiae.

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Fig. 4. Organs of infected mice (A) swelled Brain (whit arrow), (B) hepatomegaly (yellow arrow). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



Fig. 5. Its showing ciliates in different tissues within infected mice (whit arrow) (A) Brain tissue (B) Liver tissue . H& E staining 100X.

6.4. Experimental infection

Clinical signs : The affected mice showed abnormal behavior compare with non affected mice such as fear ,imbalance and lack of movement were noted .



Fig. 6. Histopathological changes in the brain tissue of infected mice showing vacuolated parenchymal in cell cerebral cortex (yellow arrow) and dilation of smal blood (whit arrow.) H&E staining 10X . (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

After dissection of infected mice appeared obvious swelling within brain as well as hepatomegaly ,were only noted in first group Fig. 4. Histopathological examination confirmed that infected mice had systemically infection with ciliates where present within examined tissues (brain & liver) Fig. 5.



Fig. 7. The arachnoid matters as a part of brain tissue in infected mice appeared with congestion(whit arrow) and great dilation in arteries(red arrow). H&E staining 4X. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

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Fig. 8. This microphoto of liver section which is infected with ciliates (A) dilatation of blood vessels(red arrow). H&E staining : 10X (B) congestion of blood vessels(whit arrow) 4X. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

€Histological structure of infected brain showing parenchymal cell in cerebral cortex underwent from vacuolation especially in pyramidal neurons and pyknosis in glial cells as well as it could be noted dilation of small blood vessels within cortical tissue Fig. 6, in addition, the congestion and great dilation in arteries within arachnoid matters were observed moreover the hemorrhage within arachnoid parenchyma was obviously recognized Fig. 7.

entral hepatic . Furthermore disorganization of liver paranchymal tissue Fig. 8.

7. Discussion

In this study *Mesanophrys* sp. like ciliates was iolated for the first time from CSF samples from patients with meningitis from Thi-Qar province / Iraq. We were investigating about opportunistic free living amoeba, throughout examination the cultured CSF on NN Agar medium. It was surprisingly we found the ciliates protozoa (Orchitophryidae) in both stages, vegetative and cystic ,. in two samples among eight samples of CSF , it observed through microscopic examination . In this study that the ciliates were isolated and cultured in vitro and then morphological ,molecular and virulence characterizations were studies .

In this study microscopic examination showed that the morphological characteristics of live the ciliates from cultured CSF was similar to genus *Mesanophrys* belong to family orchitophryidae. The ciliates had a pointed anterior and a rounded posterior with a granular structure of the cytoplasm and one macronucleus and one micronucleus, it's length was between 40 and 55 μ m and 12.5–15 μ m in width , similar to these finding, previous studies also identified *Orchitophrya or Mesanophrys* like ciliates [17,18;9].

Indicating the limitation of distinguishing and identifying ciliates by morphological analysis the ciliates can be accurately classified by molecular studies[16]). In this study the rDNA sequences of the ciliates were amplified and had a high percent identity with Orchitophrydiae family 98.56% and 97.92% but not highly similar to *Mesanophrys*, may be this ciliates in our study is a new species for *Mesanophrys*.

From the experimental infection the ciliates could enter mice through the wound made by artificial cutting. In this study histopathological results showed that *Mesanophrys* like ciliates appeared in the brain and liver tissues of infected mice and causing a sever tissue damage . our study indicate the ciliates opportunistically invade human body by wound and can cause systemic infection .

Few studies have shown that these ciliates may be opportunistic parasites ,Small et al., [18] showed a histophagous ciliates infection was discovered in a number of Nroway lobsters from the clyde sea area, the cultured ciliates and structural features that are morophologically similar to scuticociliate in the genus *Mesanophrys*.

The experimental infection confirmed that *Mesanophrys* sp. was the pathogen that infected farmed crabs in China [9,17]).

In summary the ciliates parasites isolated from human CSF is morphologically very similar to members of the genus *Mesanophrys* but rDNA sequences of the ciliates were amplified and had a high percent identity with Orchitophrydiae family but not highly similar to *Mesanophrys*, may be this ciliates in our study is a new species for *Mesanophrys*. It is suggested that these ciliates can be a great threat to human.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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