

Available online at http://bjas.bajas.edu.iq https://doi.org/10.37077/25200860.2021.34.2.03 College of Agriculture, University of Basrah Basrah Journal of Agricultural Sciences

ISSN 1814 - 5868

Basrah J. Agric. Sci. 34(2): 29-41, 2021

E-ISSN: 2520-0860

Effect of Supplementing Different Levels of Okra Seed (*Abelmoschus esculentus* L.) Powder on Growth Performance, Carcass Characteristics, Blood Parameters and Gut Microbial Populations in Broiler Chickens

Rabia J. Abbas

Animal Production Department, Collage of Agriculture, University of Basrah, Iraq

*Corresponding author: rj.abbas@yahoo.com Received 25th October 2020; Accepted 13th March 2021; Available online 1 November 2021

Abstract: A study was conducted to determine the effect of different levels of okra (Abelmoschus esculentus L.) seed powder (OSP) in diets on performance, carcass characteristics, some blood parameters, and gut microbial populations in broiler chickens. For the present experiment, 216 day old chicks were randomly divided into four groups, each group consisting three replicates of 18 chicks in a completely randomized design. Four diets were formulated with diet 1 as the basal diet (control), while 2, 3, and 4 were supplemented with OSP at 1, 2 and 4 %, respectively. The results revealed that the highest final live body weight, accumulative weight gain, and better feed conversion ratio was achieved in birds fed with 1 or 2 % of OSP. Feed intake and carcass characteristics were similar among groups. The highest relative weight of the spleen and cecum was observed in the control group, while the lowest value was observed in birds fed with 4% and 2% OSP, respectively. The longest length of the gastrointestinal tract was seen in 2% OSP compared to other groups. Serum cholesterol and ALT activity were decreased as compared to control diets. The group fed with 2 % OSP showed higher albumin levels compared to those fed with 1% of OSP. The population of total bacteria and Escherichia coli in jejunum digesta of OSP supplemented broiler chickens was reduced, meanwhile, there was an increased in lactic acid bacteria counts as compared to control. Therefore, 1 and 2 % okra seed powder in the diet of broiler chickens was able to improve the growth performance.

Keywords: Broiler, Okra seed powder, Productive performance, Biochemical traits.

Introduction

Okra, *Abelmoschus esculentus* L. (Moench) commonly known as ladies finger, besides in several other vernacular names. It was cultivated as an important vegetable crop in tropical, subtropical and warm temperate regions around the world (Roy *et al.*, 2014). It has been called "a perfect villager's vegetable" because of its robust nature, dietary fibers and distinct seed protein balanced in both lysine and tryptophan amino acids (Kumar *et al.*, 2010). *A. esculentus* L. seeds were founded to contain high levels of crude protein 24.85% (Ndangui *et al.*, 2010). Previous studies revealed that seed protein is rich in tryptophan

(94 mg.g⁻¹ nitrogen) and as well contains suitable amounts of sulfur-containing amino seeds exceptionally useful in reducing human malnutrition (NAP, 2006). Besides that, the composition of okra seed protein from amino acids is comparable to that of soybean, and the protein efficiency ratio (PER) is higher than that of soybean, as well as the pattern of amino acids in the protein, renders it an adequate supplement to legume or cereal basic diets according to Gemede et al. (2015). On the other hands, okra seed protein with good PER and net protein utilization (NPU) values is comparable to many cereals (excepting wheat) and its oil yield is similar to the most oil seed crops except oil palm and soybean oil (Kumar, et al., 2010). Also, a recent study confirmed that the great of essential amino acids of okra seeds powder were leucine (6.71%) followed by lysine (5.22%) (Abouel-Yazeed, 2019).

In terms of biomolecular characters in okra seeds, Dhruve *et al.* (2020) indicated that the highest amount of free amino acid, reducing sugars and calcium was recorded in genotype AOL 13-75 whereas, lysine was higher present in Pusasawani and methionine the highest in genotype AOL 13-90. According to studies by Benchasri (2012) and MEF (2013).

Abelmoschus esculentus L. seeds are a potential sources of oil, with levels varying from 20- 40 (%), which consists of linoleic acid up to 47.4%. Okra seed oil seed was found to contain high levels of unsaturated fatty acids, mainly oleic (up to 24.89%) and linoleic (up to 42.78%), for that, the oil can be classified in the oleic-linoleic acid group, whereas, dominant saturated acid was palmitic (up to 25.79%) (Ndangui *et al.*, 2010). Furthermore, okra seed oil has potential hypocholesterolemic effect

(Hassan & Ali, 2015). As well, it can be considered the oil of okra seeds for the prevention and treatment of human diseases and its complication as a potent antioxidant (Al-Kanani et al, 2019). Seed mucilage, of okra is may be responsible for washing away toxic substances and bad cholesterol, which loads the liver (Dhruve et al., 2015). Kumar et al. (2013) have revealed that okra contains special fiber, which takes sugar levels in blood under control, providing sugar quantity, acceptable for the bowels. According to Gemede et al. (2015), okra vegetable crop is a powerhouse of valuable nutrients, about half of which is soluble fiber in the form of gums and pectin's which help to hypocholesterolemic, lowering the risk of heart diseases.

Arapitsas (2008) reported that the main component of okra seeds was oligomeric catechins (2.5 mg.g⁻¹ of seeds) and flavonol derivative (3.4 mg.g⁻¹ of seeds), however the pod skin mostly a component of hydroxycinnamic, and quercetin derivatives $(0.2 \text{ and } 0.3 \text{ mg.g}^{-1} \text{ of skins})$, also, the author pointed out that okra seeds and pods are rich in phenolic compounds with important biological properties like quartering derivatives, catechin oligomers and hydroxycinnamic derivatives. Some studies have indicated that different parts of okra plant (flower, fruit, leaf, and seed) had high antioxidant activity (Shui & Peng, 2004; Atawodi et al., 2009; Liao et al., 2012). Hassan & Ali (2015) mentioned that the intake of okra seeds will provide the necessary energy to body, and important phenolics and flavonoids that could support immune body system and prevention of diseases. Additionally, okra fruit extracts showed antimicrobial activity against seven pathogenic strains that belong to Bacillus subtilis, Streptococcus pyogens, Klebsiella

pneumoniae, Staphylococcus aureus, Escherichia coli, Proteus mirabillis and Pseudomonas aeruginosa (Chaudhari et al., 2011). Olorunnipa et al. (2013) showed that the methanol extracts of Abelmoschus esculentus L. Moench (Okra) dried fruit had bactericidal activity against Helicobacter strains (H. pylori strains coded BAA009 and H. pylori BAA026) in vitro study.

Kalarani et al. (2017) stated that acetonic extract of various parts of okra plant (peel, seed, the combinations of peel and seeds) has been shown antibacterial activity that was higher than the activity of the positive control (Ofloxacin). Many studies have focused to describe the medicinal characteristics of okra (Amin, 2011; Nwachukwu, et al., 2014; Roy et al., 2014; Singha et al., 2014; Gemede et al., 2015). There is limited research available on the effect of okra seeds on poultry performance. supplementation of okra meal in broiler diets was not changed among groups on daily gains and feed conversion ratio, whereas there was a significant improvement in broiler skin pigmentation and belly fat considering that xanthophyll's - rich okra meal may be used as a natural coloring source in poultry feed (Liu et al., 2008). Machebe et al. (2013) reported that feeding okra seeds extracts (50 ml.l⁻¹) breeder Turkey hens significantly improved the number of eggs laid, fertility and hatchability of the eggs. Given this background, the objective of this study was to determine the effect of dietary supplementation with okra seeds meal on the productive performance in broiler chickens. As well as, to examine whether okra seeds supplementation affected carcass characteristics, internal organ properties and blood parameters.

Materials & Methods

Preparation of okra seed powder

Okra seeds were obtained from a local market of Basrah city, Iraq. The seeds were grinded to a fine powder in an electrical mixer and then mixed manually with the basal broiler diet (starter or grower diets) at the levels 1, 2 and 4 %.

Animal husbandry and treatments

Two hundred and sixteen day-old broiler chicks (Ross 308) were distributed randomly into four groups, each including three replicate battery cages (18 birds. cage⁻¹). Four diets were formulated with diet 1 as the control diet, while 2, 3, and 4 were supplemented with okra seed powder (OSP) at 1, 2, and 4 %, respectively. All the birds under the experiment were provided standard feeding that formulated to meet the nutrient requirements. The parts composition of broiler starter and grower diets has been presented in table (1). Nutrient analysis of OSP was determined as described in AOAC methods (2016) (Table 2).

Carcass traits

Six birds from each treatment (two birds per replicate) were selected based on the average weight of the group and sacrificed for carcass study at the end of 4th week, after the birds were manually eviscerated and dressed. Internal organs were carefully removed, weighed and expressed as a percentage of the live weights. The weight of carcass cuts (thigh, breast, back, wings and neck) was calculated. Dressing percent was calculated according to the equation of Al-Fayadh *et al.* (2011):

Dressing percentag = $\frac{\text{Dressed weight (gm)}}{\text{Live weight (gm)}} \times 100$

Ingredients (%)	Starter	Grower	
Ingreatents (70)	(1-20 days)	(21-28 days)	
Maize	29.80	33.80	
Wheat	15.00	15.00	
Broken rice	14.00	14.00	
Soybean meal	34.00	29.00	
Vegetable oil	1.00	2.00	
¹ Broiler protein Concentrate	5.00	5.00	
Calcium carbonate (CaCo ₃)	0.70	0.70	
² Mineral mixture	0.30	0.30	
Common salt	0.20	0.20	
Total	100	100	
³ Calculated chemical analysis			
Energy ME (Kcal. Kg ⁻¹)	3004	3106	
Crude protein (%)	23.07	21.23	
Calorie: protein ratio	130.21	146.30	
Ether extract (%)	2.30	3.35	
Crude fiber (%)	3.90	3.65	
Calcium (%)	0.81	0.81	
Available P (%)	0.34	0.33	
Lys (%)	1.25	1.13	
Met + Cys(%)	0.74	0.69	

Table (1): The composition and nutrient content of experimental rations.

¹Protein concentrate used from Al-Hayat Company, Jordanian Origin, provided per kilogram of diet: 44% Protein, 2800 kcal/kg ME, 12% Fat, 25% Ash, 5% Calcium, 2.9% Phosphorus, 2.55% Methionine + cysteine , 2.8% Lysine. ²Content/kg: Manganese 80 g; Iron 80 g; Zinc 50 g; Copper 10 g; Cobalt 2 g; Iodine 1 g; Excipient Q.S. - 1,000 g. ³Was calculated according to the feed composition Tables given in NRC (1994).

Nutrients (%)	Okra seed powder
Moisture	9.73
Crude protein	22.01
Ether Extract	14.99
Total ash	5.21
Crude fiber	17.63
Nitrogen free extract	30.43
Organic matter	94.79
¹ Metabolized Energy (Kcal.Kg ⁻¹)	2810

Table (2): Chemical composition (% on a dry weight basis) of the of okra seed powder.

¹ME was calculated by the equation described by Lodhi *et al.* (1976).

Blood parameters

At the end of the experiment, six chicks per treatment were slaughtered, and blood samples were collected in tubes without heparin for biochemical assays. The blood sample was drawn in the early morning from the wing vein of birds and allowed to clot at room temperature $(25^{\circ}C)$ for one hour prior to serum collection. The blood sample was collected in heparinized test tubes and centrifuged at 3000 rpm, 15 min, and 25°C to obtain serum. The serum preserved at -20 °C until the time of analysis. The following variables were

measured: Total proteins and albumin by a Colorimetric method using available Commercial Kits (Biolab SAS, France); sera globulin was calculated as the difference between total protein and albumin; cholesterol, triglycerides, and glucose were measured according to the methods described by Tietz (1999) by using available Commercial kits (Biolabo SAS); aspartate aminotransferase (AST) and alanine aminotransferase (ALT), were measured by using diagnostic kits (QCA, Amposta, Spain). AST: ALT ratio was calculated.

Micro-bacterial count

The number of bacteria was estimated according to pour plate method mentioned by Harrigan & McCance (1976). Samples of the contents from the jejunum were immediately collected per chick (six chicks per treatment) into glass containers, later it was transferred to the microbial population laboratory. One gram of the jejunum contents were mixed and placed in sterile glass containing 9 ml of sterile peptone solution. The dilution process was then carried out to dilute the decomposition to 10^{-2} dilution, then 1 ml of the dilute was added to 9 ml of peptone solution to reached dilution 10^{-3} . This process was re-reduced to 10⁻⁵. 10⁻⁸ dilution. In the microbial cultivation, transfer 1 ml of each dilution to two sterile plates and add 20 ml of sterilized agar culture to estimate total bacterial, MRS (De Man, Rogosa, and Sharpe agar) agar (facultative anaerobes Lactobacillus spp.) and McConkey agar to estimate colonic bacteria (Escherichia coli). For bacterial growing, all the plates were incubated at 37°C, MRS agar plates were incubated anaerobically for 48 hours and other plates were incubated aerobically for 24 hours. The bacteria counts

were estimated by multiplying the total number of bacterial colonies at each incubation period in a dilution inverted.

Statistical analysis

The data were subjected to analysis of variance (One –way ANOVA) in accordance to Completely Randomized Design (CRD) using SPSS software (2015) to analyze the results. Least Significant Difference (L.S.D.) test was applied to the separated means at significant level of 0.05 (SPSS, 2015).

Results

Effect of OSP supplementation on productive performance of broilers

The effect of dietary supplementation with okra (Abelmoschus esculentus) seed powder (OSP) on the productive performance of broiler chickens is presented in Table 3. The result revealed that the groups supplemented with 1 and 2 % OSP caused significantly ($P \le 0.05$) higher body weight and weight gain when compared with 4% treated and control group at 28 days of age, meanwhile all OSP supplementations did not show any effect on total feed intake. As table (3) shows, there were a significant improvement for feed conversion ratio (FCR) at levels 1 and 2% during experimental period as compared with control and higher levels of OSP (4%).

Carcass and internal organ characteristics

No significant differences were found with regard the carcass characteristics to measurements (Table 4). Table (5) shows that, with the exception of the relative weight of the spleen, gastrointestinal tract length and caecum weight, all other organs measured were not significantly influenced by the dietary treatments.

Deverators	Treatment of okra seeds powder (%)					
Parameters	0	1	2	4		
Initial live weight (g)	$35.4^{a} \pm 0.23$	$35.5^{a}\pm 0.70$	$34.2^{a} \pm 0.23$	$35.0^{a} \pm 0.62$		
Final body weight (g)	1473 ^{bc} ±17.89	$1540^{a}\pm0.88$	1524 ^{ab} ±23.09	1468°±3.28		
Total weight gain(g)	1437.6 ^b ±17.67	1504.5 ^a ±1.63	1489.8 ^a ±22.86	1433.0 ^b ±12.82		
Total feed intake (g.bird ⁻¹)	2351 ^a ±33.26	2366 ^a ±25.58	2311 ^a ±0.87	2303 ^a ±4.30		
Feed conversion ratio(g:g)	$1.64^{b} \pm 0.014$	$1.57^{a}\pm0.019$	$1.55^{a}\pm0.008$	1.61 ^{ab} ±0.017		

Table (3): Effect of okra seeds powder on the overall (0-28 d) performance of broiler chicks.

^{a-c} Means within main effects with no common superscripts are different significantly $p \le 0.05$)

Result for relative spleen weight showed that birds fed the control and 1 and 2 % okra seed diets had the highest significant values that were similar (P \leq 0.05) but higher than the value recorded by those fed the 4% okra seed diet. Birds fed a 2% okra seed diet had the longest length of the gastrointestinal tract as compared to other groups. Caecum relative weight was higher (P \leq 0.05) in control (1.004%) as compared to 2% okra seed group.

Parameters	Okra seeds (%)					
	0	1	2	4		
Carcass weight (g)	1191ª± 40.90	1224ª± 34.05	$1110^{a} \pm 44.97$	$1145^{a} \pm 53.24$		
Carcass yield (%)	$71.03^{a}\pm 0.84$	$71.66^{a} \pm 0.48$	$71.28^{a} \pm 0.56$	$71.49^{a} \pm 0.29$		
Breast yield (%)	$37.74^{a}\pm 0.86$	$38.56^{a} \pm 2.07$	$38.85^{a} \pm 2.08$	$36.35^{a} \pm 0.67$		
Thigh yield (%)	27.41 ^a ± 0.61	27.84 ^a ± 1.69	30.28ª± 3.20	28.33ª± 1.14		
Back yield (%)	$17.66^{a} \pm 0.80$	$18.18^{a} \pm 1.05$	$17.26^{a} \pm 0.96$	$18.30^{a} \pm 0.19$		
Wing yield (%)	$10.19^{a}\pm 0.42$	9.53 ^a ± 0.33	$10.46^{a} \pm 0.36$	9.69ª± 0.39		
Neck yield (%)	$4.18^{a}\pm 0.38$	4.07 ^a ± 0.35	4.35 ^a ± 0.20	$4.86^{a} \pm 0.47$		
Total giblets (%)	$6.04^{a}\pm 0.23$	$5.96^{a} \pm 0.16$	6.14 ^a ± 0.13	5.88ª± 0.21		
Liver yield (%)	$2.83^{\mathrm{a}}\pm0.07$	$2.56^{a} \pm 0.14$	$2.78^{a} \pm 0.13$	$2.56^{a} \pm 0.09$		
Heart yield (%)	$0.52^{a}\pm 0.03$	$0.61^{a}\pm 0.01$	$0.56^{a} \pm 0.02$	$0.56^{a} \pm 0.03$		
Gizzard yield (%)	$2.64^{a}\pm 0.24$	$2.79^{a} \pm 0.12$	$2.80^{\mathrm{a}} \pm 0.22$	$2.76^{a} \pm 0.16$		

 Table (4): Effect of okra seeds powder on carcass traits in broiler chicks.

^aMeans in the same row with a common superscript are insignificant differences ($P \ge 0.05$).

Serum parameters

Table (6) presents the serum metabolites to the dietary groups. Total serum protein, globulins, Albumin to globulins ratio, glucose, triglyceride, AST activity, and AST to ALT ratio did not vary significantly due to dietary groups. A significant difference ($P \le 0.05$) was recorded on serum albumin, cholesterol and ALT activity of broilers fed various diets. Serum cholesterol and ALT activity were

declined ($P \le 0.05$) in all experimental groups as compared with the control one, on other hands, the group of 2 % okra seed powder showed higher ($P \le 0.05$) albumin levels than those fed 1% okra seed powder.

Micro-bacterial count

The results in table (7) showed that there was a significant ($p \le 0.05$) reduction in total bacterial and *E. coli*, whereas the *Lactobacilli* population

Treatment of okra seeds powder (%)					
0	1	2	4		
$0.16^{a} \pm 0.007$	0.13 ^{ab} ±0.005	$0.15^{a} \pm 0.02$	$0.11^{b} \pm 0.008$		
$0.32^{a} \pm 0.05$	0.32 ^a ±0.03	$0.36^{a} \pm 0.02$	0.31 ^a ±0.03		
$0.51^{a} \pm 0.02$	0.51 ^a ±0.07	$0.54^{a} \pm 0.08$	$0.59^{a} \pm 0.06$		
$4.87^{a} \pm 0.18$	5.43 ^a ±0.49	$5.32^{a} \pm 0.24$	5.23 ^a ±0.43		
$11.70^{b} \pm 0.47$	12.02 ^b ±0.75	13.97 ^a ±0.34	$12.13^{b} \pm 0.44$		
0.21 ^a ±0.14	$0.22^{a} \pm 0.02$	$0.24^{a} \pm 0.03$	$0.23^{a} \pm 0.01$		
$1.00^{a} \pm 0.00$	$1.07^{a} \pm 0.16$	$1.16^{a} \pm 0.15$	$1.14^{a} \pm 0.10$		
1.00 ^a ±0.23	$0.69^{ab} \pm 0.07$	$0.56^{b} \pm 0.05$	$0.64^{ab} \pm 0.10$		
$1.46^{a} \pm 0.12$	1.47 ^a ±0.15	1.31 ^a ±0.13	1.19 ^a ±0.15		
	$\begin{array}{c} 0\\ \hline 0.16^{a}\pm 0.007\\ \hline 0.32^{a}\pm 0.05\\ \hline 0.51^{a}\pm 0.02\\ \hline 4.87^{a}\pm 0.18\\ \hline 11.70^{b}\pm 0.47\\ \hline 0.21^{a}\pm 0.14\\ \hline 1.00^{a}\pm 0.00\\ \hline 1.00^{a}\pm 0.23\\ \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $		

 Table (5): Effect of okra seeds powder on some gut measurements and relative organ weights in broiler chicks.

^{a-b}Means in the same row with a common superscript are insignificant differences ($P \ge 0.05$).

Table (6): Some blood parameters at 28 days of age of chick fed okra seeds powder.

Parameters	Treatment of okra seeds powder (%)				
	0	1	2	4	
Total Protein (g.dl ⁻¹)	3.63 ^a ±0.14	$3.66^{a} \pm 0.23$	4.28 ^a ±0.33	$4.16^{a} \pm 0.51$	
Albumin (g.dl ⁻¹)	2.46 ^{ab} ±0.13	$2.22^{b}\pm0.07$	2.52 ^a ±0.05	2.43 ^{ab} ±0.08	
Globin (g.dl ⁻¹)	1.17 ^a ±0.12	$1.44^{a} \pm 0.21$	$1.76^{a} \pm 0.28$	$1.73^{a} \pm 0.46$	
Albumin to Globulin ratio	2.19 ^a ±0.35	$1.63^{a} \pm 0.22$	$1.52^{a} \pm 0.17$	$1.64^{a} \pm 0.29$	
Glucose (mg/dl)	$229.18^{a} \pm 10.81$	$317.25^{a} \pm 51.54$	250.35 ^a ±10.88	298.47 ^a ±60.83	
Cholesterol (mg.dl ⁻¹)	120.08 ^a ±9.26	105.91 ^b ±4.56	87.79 ^b ±2.49	104.76 ^b ±2.06	
Triglyceride (mg.dl ⁻¹)	$86.25^{a} \pm 9.57$	$125.87^{a} \pm 10.34$	119.07 ^a ±9.99	126.75 ^a ±24.42	
1 AST (U.1 ⁻¹)	392.0 ^a ±16.56	379.12 ^a ±13.81	$395.42^{a} \pm 8.14$	367.81 ^a ±13.49	
2 ALT ((U.1 ⁻¹)	254.41 ^a ±2.24	242.86 ^b ±1.09	247.09 ^b ±3.03	242.55 ^b ±1.24	
AST to ALT ratio	$1.54^{a} \pm 0.05$	$1.56^{a} \pm 0.06$	1.60 ^a ±0.02	$1.52^{a} \pm 0.05$	

^{a,b}Means bearing different superscripts within a row differ significantly (P≤0.05)

 1 AST = Aspartate aminotransferase; 2 ALT= Alanine aminotransferase.

Table (7): Micro-bacterial a count (log	cfu.g ⁻¹) as affected by supplemented with okra seed powder.

Bacteria types	Treatment of okra seeds powder (%)				
	0	1	2	4	
Total bacterial count (TBC) ($\times 10^8$)	$3.64^{a}\pm0.41$	2.63 ^b ±0.42	$2.05^{b}\pm0.32$	2.47 ^b ±0.34	
lactic acid bacteria (LAB) ($\times 10^7$)	$2.99^{b}\pm0.50$	5.47 ^a ±0.54	$4.80^{a}\pm0.66$	5.19 ^a ±0.47	
<i>Escherichia coli</i> (E. coli) ($\times 10^{6}$)	$6.68^{a}\pm0.40$	$3.48^{b} \pm 1.08$	$6.00^{b} \pm 0.48$	$5.87^{b}\pm0.83$	

^{a,b}Means within main effects with no common superscripts are different significantly $p \le 0.05$).

was increased significantly in dietary okra seeds powder (*Abelmoschus esculentus* L.) supplemented diets as compared to control at the jejunum.

Discussion

Growth performance of broiler

The indicated results herein revealed that the supplementation of okra seed powder (OSP)

enhanced the growth performance of broilers (Table 3). However, Liu *et al.* (2008), noticed that supplementation of okra seed powder on broiler diets did not significantly affect the broiler's daily gain and feed efficiency. The positive effects of supplementing okra seed powder on broilers performance may be due to the antimicrobial activity and antioxidants activity of the components of the OSP, which resulted in better intestinal health and improved digestion and absorption (Chaudhari *et al.*, 2011; Olorunnipa *et al.*, 2013; Kalarani *et al.*, 2017).

Among the factors that positively affect the growth rate of birds are the nutritional components present in okra seeds which rich in unsaturated fatty acids especially linoleic acid, with adequate amounts of sulfur-containing amino acid, that could be considered as good sources of protein with good PER and NPU values, additionally rich in important minerals such as phosphorus, magnesium, calcium and potassium (Ndangui et al., 2010; Gemede et al., 2015), which may enhancing weight and feed efficiency in chicks. On the other hands, study on nutritional analysis of okra seeds, Dhruve et al. (2015), suggested that the consumption of OSP, will provide the necessary energy to the body and important antioxidants that could boast immune body system and prevent diseases. Similar results on the feed intake (Table 3) were reported by Liu et al. (2008), when okra stems and leaf powder were used in broiler chicks' diets.

Carcass characteristics and internal organs

The supplementation with okra seed powder did not cause any significant differences in carcass traits (Table4) compared with the control. Similarly, no significant differences between relative organ weights (liver, heart, kidney, lungs and brain) were observed in mice ingested with 200 mg. kg⁻¹ aqueous and methanolic seed extracts of A. esculentus (Doreddula et al., 2014). In this study, the spleen, caecum weights and gastrointestinal tract length differed between various okra seed powder levels in diets (Table 5). The significant increase in gastrointestinal tract length obtained in okra seed supplemented diets may cause better digest feed efficiently, which reflected to improve feed efficiency ratio that observed in experimental groups (Table 3). Mabelebele et al. (2014) stated that lengthy digestive tracts in broiler chickens indicated a higher surface area for nutrient digestion and absorption. On the other hand, the information presented by Gemede et al. (2015) showed that the potential nutritional importance of okra and its role in improved nourishment and health. In addition, it can use it to apply in manufacturing for dietary improvement, due to its several health benefits (Abouel-Yazeed, 2019).

Blood biochemistry

Serum cholesterol and ALT activity were significantly (P \leq 0.05) decreased in all supplemented groups with okra seed powder as compared with the control group (Table 6). The reduction of serum total cholesterol may be due to okra content such as polysaccharide, fiber or antioxidant. In this respect, Kahlon *et al.* (2007) were pointed that okra polysaccharide reduces the cholesterol concentration in the blood due to its ability to bind bile acids. In this connection, Ngoc et al. (2008), detected that the extracts from the total plant of A. esculentus by dichloromethane (AE1) or methanol (AE2) and extracts from the fruit by dichloromethane (AE3) or methanol (AE4) in mice, may be useful in lowering cholesterol and triglyceride

levels in hyperlipidemia. Doreddula et al. (2014) demonstrated that the pretreatment of mice with aqueous and methanolic seed extracts of Abelmoschus esculentus (200 m.kg⁻¹ p. o.) for seven days significantly reduced the blood glucose, cholesterol and triglyceride levels elevated by acute restraint stress, their results were similar to our finding in respect of cholesterol, but differ with glucose and triglyceride levels which did not vary significantly. In the sense, the fibers in ladies finger (okra, Abelmoschus esculentus L.) lead to stabilize blood sugar by regulating the rate at which sugar is absorbed from the intestinal tract (Doreddula et al., 2014). The result from other researchers revealed that okra contains special fiber which takes sugar levels in blood under control, providing sugar quantity, acceptable for the bowels (Kumar et al., 2013).

According to the report of Hu et al. (2014) the ladies finger polysaccharide has been possesses hepatoprotective, besides antidiabetic (Aligita et al., 2019). Additionally, antidiabetic and antihyperlipidemic activities were reported in rats. According to previous study published by Sabitha et al. (2011, 2012) the diabetic rats that fed okra peel and seed powder at 100 and 200 mg.kg-1 dose, exhibited a significant decrease in sera glucose levels and a boost in body weight as the comparison with untreated, diabetic rats. Furthermore, the antidiabetic activity of the okra fruit extract with a dose of 50 mg. kg⁻¹ BW, with the mechanism of action by increasing insulin secretion and rising insulin sensitivity, as well as inhibited carbohydrate absorption in the intestine (Aligita et al., 2019).

Micro-bacterial count

The results showed that there was a significant $(p \le 0.05)$ reduction in total bacterial and Echerichiae coli count (Table 7), whereas the Lactobacilli population was increased significantly in dietary okra (Abelmoschus esculentus L.) supplemented diets as compared to control in jejunum. The reduction in count of E. coli was in accordance with the study of Chaudhari et al. (2011), De Carvalho et al. (2011) and Kalarani et al. (2017), who showed that okra extracts (A. esculentus L.) had bactericidal activity against E. coli, besides other pathogenic bacteria. In addition, Olorunnipa et al. (2013) suggested that anti-Helicobacter pylori activities exhibited by A. esculentus L. Moench its local use in the treatment of gastrointestinal diseases associated with the H. pylori species. Moreover, the effectiveness of A. esculentus (okra) in treating gastric irritations and inflammatory diseases was due to polysaccharides that inhibit the adhesion of H. pylori to stomach tissue (Messing et al., 2014). According to the report of Jones (2017) antimicrobial impact of purified okra seed proteins (POP) against pathogens in water treatment marked an excellent inactivation of approximately 100% of fecal coliform and E-coli count in raw water was achieved and zero re-growth of bacteria after 72-hour post-treatment, in this sense, the use of POP in water treatment may improve access to clean water and could help in reducing the import of water treatment chemicals in developing countries. The inactivation of total bacteria and E-coli count by the okra seed may be due to the presence of various phytochemical compounds such as tannin, saponin, and alkaloids in the seeds, (Jones, 2017).

Conclusion

It was concluded that supplementation okra seed powder (up to 4% of diet) can reduce the levels of serum total cholesterol and ALT activities, Moreover, the supplementation of OSP led to the reduction of harmful bacteria (*E. coli*) and increased numbers of beneficial (lactic acid) in the jejunum which can help to improve intestinal health. In this trial, OSP at levels 1 and 2 % had the ability to enhance the productive performance of broilers.

Acknowledgments

The author sincerely thanks Dr. Kutaiba J. Al-Kafaji and Dr. Alfred S. Karomy from Department of Animal Production, College of Agriculture, University of Basrah for their assistance in this study.

Conflict of interest

The author doesn't have any probable conflict of interest regarding the publisher's policy requirements.

Ethical approval

In this study, all ethical standards issued by national and international institutions related to poultry breeding and care was applied.

ORCID:

Abbas: https://orcid.org/0000-0003-1330-9894

References

- Abouel-Yazeed, A. M. (2019). Incorporation of okra seeds powder to employ in some foodstuffs based on its physical, chemical and sensorial evaluation. *Journal of Food and Dairy Sciences*, 10, 231-236. http:// 10.21608/jfds.2019.53498
- Al-Fayadh, H. A. A., Naji, S. A. H., & Al-Hajo, N. N. (2011). *Poultry Meat Technology*. 2nd part. Higher

Education Press, University of Baghdad, 292pp. (In Arabic).

- Aligita, W., Muhsinin, S., Susilawati, E., DahliaPratiwi,
 D.S., Aprilliani, D., Artarini, A., & Adnyana, I.K.
 (2019. Antidiabetic activity of okra (*Abelmoschus* esculentus L.) fruit extract. Rasayan Journal of Chemistry, 12, 157-167. http://dx.doi.org/10.31788/RJC.2019.1215059
- Al-Kanani, I.A. S., Al-Hilifi, S. A. H., & Abd-Al-Kareem, A. H. (2019). Extraction of Iraqi okra seeds oil and study of its properties during different periods. *Basrah Journal of Agricultural Sciences*, 32(Special Issue 2), 63-74. https://doi.org/10.37077/25200860.2019.257
- Amin, I. M. (2011). Nutritional properties of Abelmoschus Esculentus as remedy to manage diabetes mellitus: A literature review. International Conference on Biomedical Engineering and Technology, IPCBEE vol. 11 (2011) © (2011) IACSIT Press, Singapore.
- A.O.A.C. (2016). *Official Methods of Analysis of AOAC International*. George W., & Latimer, Jr. (Eds.), 20th edition. Rockville, Maryland 20850-3250, 3172pp.
- Arapitsas, P. (2008). Identification and quantification of polyphenolic compounds from okra seeds and skins. *Food Chemistry*, 110, 1041-1045. http://doi. 10.1016/j.foodchem.2008.03.014
- Atawodi, S. E., Atawodi, J. C., Idakwo, G. A., Pfundstein, B., Haubner, R., Wurtele, G., Spiegelhalder, B., Bartsch, H., & Owen, R.W. (2009). Polyphenol composition and antioxidant potential of *Hibiscus esculentus* L. fruit cultivated in Nigeria. *Journal of Medicinal Food*, *12*, 1316–1320. http://doi.<u>10.1089/jmf.2008.0211</u>
- Benchasri, S. (2012). Okra (Abelmoschus esculentus (L.) Moench) as a valuable vegetable of the world. *Ratarstvoi Povrtarstvo.* 49, 105-112. https://doi.10.5937/ratpov49-1172
- Chaudhari, Y, Kumar, E. P., Badhe, M., Mody, H, R., & Acharya, V. B. (2011). An evaluation of antibacterial activity of *Abelmoschus esculentus* on clinically isolated infectious disease causing bacterial pathogen from hospital. *International Journal of Pharmaceutical and*

Abbas, / Basrah J. Agric. Sci., 34(2): 29-41, 2021

Phytopharmacological Research, *1*, 107-111. Available on line https://eijppr.com/GDOCQ5P

- De Carvalho, C. C. C. R., Cruz, P.A., da Fonseca, M. M. R., & Xavier-Filho, L. (2011). Antibacterial properties of the extract of *Abelmoschus esculentus*. *Biotechnology and Bioprocess Engineering*, 16, 971-977. http://doi. 10.1007/s12257-011-0050-6
- Dhruve, J. J., Shukla, Y., Shah, R., Patel, J., & Talati, J. G. (2015). Contribution of okra (*Abelmoschus esculentus* L.) seeds towards the nutritional characterization. World Journal of Pharmaceutical Sciences, 4, 1009-1023.
- Dhruve, J. J., Shukla, M. Y., Shah, R., Patel, J., & Talati, J. G., (2020). Contribution of okra (Abelmoschus esculentus L.) seeds towards the nutritional characterization. World Journal of Pharmacy and Pharmaceutical Sciences, (Abs.). https://www.wjpps.com/Wjpps_controller/abstract_i d/3406
- Doreddula, K. S., Bonam, R. S., Gaddam, P. D., Desu, R.S.R., Ramarao, N., & Pandy, V. (2014).
 Phytochemical analysis, antioxidant, antistress, and nootropic activities of aqueous and methanolic seed extracts of ladies finger (*Abelmoschus esculentus* L.) in mice. *The Scientific World Journal, 2014*, 1-14. https://doi. 10.1155/2014/519848
- Gemede, H. F., Ratta, N., Haki, G. D., Woldegiorgis, A. Z., & Beyene, F. (2015). Nutritional quality and health benefits of okra (*Abelmoschus esculentus*): A review. *Journal of Food Processing & Technology*, 6, 1-6. http://doi:10.4172/2157-7110.1000458
- Harrigan W. F., & McCance M. E. (1976). Laboratory Methods in Microbiology. 1st edition, Academic Press, London and New York. 374pp.
- Hassan, M. A. M., & Ali, H. M. (2015). The nutritional composition of three cultivars of okra (*Abelmoschus esculentus* L.) seeds flour. *World Journal of Dairy & Food Sciences*, 10, 122-131. https://doi. 10.5829/idosi.wjdfs.2015.10.2.1150
- Hu, L., Yu, W., Li, Y., Prasad, N., & Tang, Z. (2014). Antioxidant activity of extract and its major constituents from okra seed on rat hepatocytes injured by carbon tetrachloride. *BioMed Research International*, 2014, 1-9. https://doi.org/10.1155/2014/341291

- Jones, N. A. (2017). Investigation the bacterial inactivation potential of purified okra (*Hibiscus esculentus*) seed proteins in water Purification. *Arid Zone Journal of Engineering, Technology and Environment, 13*, 6-14.
- Kahlon, T. S., Chapman, M. H., & Smith, G. E., (2007). *In vitro* binding of bile acids by okra, beets, asparagus, eggplant, turnips, green beans, carrot and cauliflower. *Food Chemistry*, 103, 676–680. https://doi.org/10.1016/j.foodchem.2006.07.056
- Kalarani, G., Govindharajan, M., Leoney, A., Prabhu, A., Krishnakumar, R., & Dhurikaliyamorthy, S. (2017).
 Antibacterial effect of *Abelmoschus esculentus* (okra) extracts on dental caries derived *Streptococcus mutans. Journal of Dental and Medical Sciences*, 16, 63-66. https://doi.10.9790/0853-1604026366
- Kumar, D. S., Tony, D. E., Kumar, A. P., Kumar, K. A., Rao, D. B. S., & Nadendla, R. (2013). A review on: *Abelmoschus esculentus* (Okra). *International Research Journal of Pharmaceutical and Applied Sciences*, 3, 129-132. https://scienztech.org/irjpas/article/view/586
- Kumar, S., Dagnoko, S., Haougui, A., Ratnadass, A., Pasternak, D., & Kouame, C. (2010). Okra (Abelmoschus spp.) in West and Central Africa: Potential and progress on its improvement. *African Journal of Agricultural Research*, 5, 3590-3598. http://oar.icrisat.org/168/1/nset4.pdf
- Liao, H., W. Dong, X. Shi, H.Liu, & K. Yuan. (2012). Analysis and comparison of the active components and antioxidant activities of extracts from *Abelmoschus esculentus* L. *Pharmagnosy Magazine*, 8, 156–161. http://dx.doi.org/10.4103/0973-1296.96570
- Liu, G. –D., Hou, G.-Y., Wang, D.-J., Lv, S.-J., Zhang, X.-Y., Sun, W.-P., & Yang, Y. (2008). Skin pigmentation evaluation in broilers fed different levels of natural okra and synthetic pigments. *Journal of Applied Poultry Research*, 17, 498–504. https://doi.org/10.3382/japr.2008-00058
- Lodhi, G. N., Singh, D., & Ichhponani, J. S., (1976). Variation in nutrient content of feeding stuffs rich in protein and reassessment of the chemical method for metabolizable energy estimation for poultry. *The*

Journal of Agricultural Science, 86, 293-303. https://doi.org/10.1017/S0021859600054757

- Mabelebele, M., Alabi, O. J., Ng ambi, J. W., Norris, D., & Ginindza, M. M., (2014). Compaction of gastrointestinal tract and pH values of digestive organs of Ross 308 broiler and indigenous Venda chickens fed the same diet. *Asian Journal of Animal* and Veterinary Advances, 9, 71-76. https://doi. 10.3923/ajava.2014.71.7.
- Machebe, N. S., Ugwu, S. O, Atu, C. S., & Mbunwen, N-F H., (2013). Intake of some biological seeds and root extracts of plants improves fertility and hatchability of Turkey eggs. *Journal of Basic and Applied Sciences 9*, 538-542. https://doi. 10.6000/1927-5129.2013.09.69
- MEF. (Ministry of Environment and Forest) (2013). *Biology of Okra. Series of crop specific biology document*. Ministry of Environmental and Forest Government of India. 26pp. http://bangladeshbiosafety.org/wpcontent/uploads/2017/06/Biology_of_Okra_In.pdf
- Messing, J., Thöle, C., Niehues, M., Shevtsova, A., Glocker, E., Borén, T., & Hensel, A. (2014). Antiadhesive properties of *Abelmoschus esculentus* (Okra) immature fruit extract against *Helicobacter pylori* adhesion. *PLoS ONE*, 9, e84836. https://doi.org/10.1371/journal.pone.0084836
- NAP. (2006). National Academies Press. Lost crops of Africa volume II: Vegetables. Washington, D.C., The National Academies Press, 378pp. www.nap.edu/catalog/11763.html: pp. 287-301
- Ndangui, C. B., Kimbonguila, A., Nzikou, J. M., Matos, L., Pambou-Tobi., N. P. G, Abena, A. A., Silou, Th., Scher, J., & Desobry, S. (2010). Nutritive composition and properties physic -chemical of *Research Journal of Environmental and Earth Sciences*, 2, 49-54.
- Ngoc, T. H., Ngo, Q. N., Van, A.T. T., & Vo Phung, N. (2008). Hypolipidemic effect of extracts from *Abelmoschus esculentus* L. (Malvaceae) on Tyloxapol-induced hyperlipidemia in mice. Mahidol University Journal of Pharmaceutical Sciences, 35, 42-46.
- Nwachukwu, E. C., Nulit, R., & Go, R. (2014). Nutritional and biochemical properties of Malaysian

okra variety. Advancement in Medicinal Plant Research, 2, 16–19.

- Olorunnipa, T. A., Igbokwe, C. C., Lawal, T. O., Adeniyi, B. A., & Mahady, G. B. (2013). Anti-Helicobacter pylori activity of Abelmoschus esculentus L. Moench (okra): An in vitro study. Clinical Microbiology, 2, 132. https://doi.10.4172/2327-5073.1000132
- Roy, A., Shrivastava, S. L., & Mandal, S. M. (2014). Functional properties of okra *Abelmoschus esculentus* L. (Moench): traditional claims and scientific evidences. *Plant Science Today*, 1, 121-130. https://doi. 10.14719/pst.2014.1.3.63
- Sabitha, V., Ramachandran, S., Naveen, K. R., & Panneerselvam, K. (2011). Antidiabetic and antihyperlipidemic potential of *Abelmoschus* esculentus (L.) Moench in streptozotocin-induced diabetic rats. Journal of Pharmacy and Bioallied Sciences, 3, 397–402. https://doi.10.4103/0975-7406.84447
- Sabitha, V., Ramachandran, S., Naveen, K. R., & Panneerselvam, K. (2012). Investigation of *in vivo* antioxidant property of *Abelmoschus esculentus* (L) Moench fruit seed and peel powders in streptozotocin-induced diabetic rats. *Journal of Ayurveda and Integrative Medicine*, *3*, 188-193. http://dx.doi.org/10.4103/0975-9476.104432
- Shui, G., & Peng, L. L. (2004). An improved method for the analysis of major antioxidants of *Hibiscus* esculentus Linn. Journal of Chromatography A, 1048, 17–24. https://doi.10.1016/j.chroma.2004.07.032
- Singha, P., Chauhana, V., Tiwaria, B. K., Chauhanb, S. S., Simonb, S., Bilalc, S., & Abidia, A. B. (2014). An overview on okra (*Abelmoschus esculentus*) and its importance as a nutritive vegetable in the world. *International Journal of Pharma and Bio Sciences*, 4, 227-233. www.ijpbs.com
- SPSS, Statistical Package for the Social Sciences. (2015). Quantitative Data Analysis with IBM SPSS version 23: A Guide for Social Scientists. New York: Routledge.
- Tietz, N. W. (1999). Textbook of Clinical Chemistry. 3rded. Burtis, E R Ash Wood, W.B. SaundersCompany,Philadelphia,616pp.

Abbas, / Basrah J. Agric. Sci., 34(2): 29-41, 2021

تأثير إضافة مستويات مختلفة من مسحوق بذور الباميا (Abelmoschus esculentus L.) في أداء النمو، خصائص الذبيحة، المعايير الدمية والمجتمع الميكروبي لأمعاء فروج اللحم

> **ربيعة جدوع عباس** قسم الإنتاج الحيواني، كلية الزراعة، جامعة البصرة، العراق

المستخلص: اجريت الدراسة الحالية لمعرفة تأثير اضافة مسحوق بذور الباميا (.Abelmoschus esculentus L) الى العليقة في الأداء الانتاجي, صفات الذبائح, بعض معايير الدم واعداد البكتريا في الامعاء لفروج اللحم. تم توزيع 216 فرخاً بعمر يوم واحد من فروج اللحم عشوائياً على اربع مجاميع وبواقع ثلاث مكررات للمجموعة الواحدة وبواقع 18 فرخا لكل مكرر وفقا للتصميم العشوائي فروج اللحم عشوائياً على اربع مجاميع وبواقع ثلاث مكررات للمجموعة الواحدة وبواقع 18 فرخا لكل مكرر وفقا للتصميم العشوائي الكامل. كانت الاولى معاملة السيطرة (العليقة الاسامية) وأضيف مسحوق بذور الباميا الى العليقة الأساسية بالمستويات 1، 2 و 4 في المحاميع الثانية والثالثة والرابعة على التوالي أظهرت النتائج ان أعلى معدل لوزن الجسم النهائي، الزيادة الوزنية, وافضل معدل % في المجاميع الثانية والثالثة والرابعة على التوالي أظهرت النتائج ان أعلى معدل لوزن الجسم النهائي، الزيادة الوزنية, وافضل معدل للتحويل الغذائي تحقق في المجموعة على اعطيت 1 و 2% من مسحوق بذور الباميا, في حين لم يتأثر معدل استهلاك العلف وصفات الدبائح بمعاملات الاضافة. لوحظ ان أعلى وزن نسبي للطحال والأعورين تحقق في مجموعة السيطرة ، بينما ظهرت أقل وصفات الذبائح بمعاملات الاضافة. لوحظ ان أعلى وزن نسبي للطحال والأعورين تحقق في مجموعة السيطرة ، بينما ظهرت أقل بعمة عند المستوى 4% و 2% من مسحوق البذور على التوالي, وسجلت مجموعة 2% فضل الاطوال للقناة الهضمية مقارنة قيمة عند المستوى 4% و 2% من مسحوق البذور على التوالي، وسجلت مجموعة 2% فضل الاطوال للقناة المضمي 2% بالمجاميع الأخرى. وحصل انخفاض معنوي في تركيز الكوليسترول وفي فعالية انزيم ALT مقارنة بالمستوى 2% وحصل انخفاض معنوي في تركيز الكوليسترول وفي فعالية انزيم معدل الميورة. وأظهر المستوى 2% المجاميع الأخرى. وحصل انخفاض معنوي في تركيز الكوليسترول وفي فعالية انزيم معلون الكلية وبكتريا في حموي في يربي المالية المور للميورة بينما ظهرت أقل بالمجاميع الأخرى. وحصل انخفاض معنوي في عادية الزيم 2% فصل الاطوال للقناة الهضمية كزي بالمجاميع الأخرى. وحصل انخفاض معنوي في عداد الميتريا الاليومين مقارنة بالمستوى 1%. وحصل انخفاض معنوي في اعداد الكتريا المالي ليتزي الأدا مامي معانية. يومني قاده المورة فضلاً عن تحسن معنوي في اعداد بكتريا الالماليي العزي أي مام مليمة في مردي الكومي. توم

الكلمات المفتاحية: فروج اللحم, مسحوق بذور الباميا، الاداء الانتاجي، الصفات الكيموحياتية للدم.