

# Body Fat Distribution and the Levels of Some Biochemical, Hormonal and Inflammatory Biomarkers in Obese Healthy Women

Sanaa Jameel Thamer\*

University of Basrah, College of Sciences, Department of Biology, Iraq

Corresponding author email: [sanaa.thamer@uobasrah.edu.iq](mailto:sanaa.thamer@uobasrah.edu.iq).

## ABSTRACT

The metabolic alternations related to obesity are a serious risk factors that leads to many health problems. The aim of the study is to estimate some biochemical and hormonal parameters in the blood of obese females with different distribution of body fat. Methods: premenopausal healthy females (n=148) that have various body weight and aged (39-43 years), which were assessed their obesity by BMI and the regional body fat by the WC, WHR and WHtR parameters. The Navy seal formula were used to detect body composition. Blood biochemical, insulin, cortisol, thyroid hormones, total testosterone, visfatin, IL8 and CRP were tested in the serum. Results: according to WC and WHR, the obese females (n=83) revealed two types of obesity, the central and peripheral obesity. The Navy formula showed high significant fat ratio and fat mass particularly in central obesity females. The serum of obese category showed significant increasing in fasting glucose, insulin, HOMIR with dyslipidemia. The serum cortisol, total testosterone, visfatin, IL8 and CRP were also elevated in obese females with significant differences in central obesity group than the peripheral obesity group. Compared to normal weight females, the obese females revealed significant increasing in TSH and FT3 levels while the FT4 was insignificant. Conclusion: the obesity in healthy females especially the central obesity type was associated with alternations in serum profiles of some hormonal and inflammatory biomarkers.

**Keywords** Body Fat, metabolism, CRP.

## Introduction

Obesity is the most common metabolic disorders in the world, especially in developing countries [1]. The overweight or body fat depots consider the first marker of health problems including hypertension, diabetes and cardiovascular disease [2]. The body fat is classified into central (android shape) and peripheral fat (gynoid shape) [3]. The central fat is results from fat deposit in abdominal region, while the deposit around the hips and limbs results in peripheral adiposity [4]. The abdominal fat is distributing into two lines, the subcutaneous and visceral fats, the visceral obesity plays important role as an endocrine gland that releasing many adipocytokines, those effecting on glucose and lipid metabolism and contributed to several metabolic dysfunctions, involving hyperinsulinemia, dyslipidemia [5]. The obesity in both women and men was observed to associate with many hormonal abnormalities, in women, the central (abdominal) obesity is correlated directly with diabetes and insulin resistance [6]. The ratio of urinary cortisone/ cortisol is increased in women with abdominal obesity [7] and the high cortisol level was correlated with central body fat [8]. The abnormalities in sex hormones are associated with obesity in female by decreasing in estradiol and follicle stimulating hormones [9]. The functions of thyroid gland in obesity is affecting on weight gain and on development of metabolic risk factors [10]. The obesity can estimated by body mass index (BMI) that widely used as anthropometric parameter for general adiposity but not identify the body composition [11]. The indirect measurement of visceral fat is using waist circumference WC and waist to hip ratio WHR [12], while the peripheral fat distribution measured by hip or thigh circumference [13]. The previous information can elicit that the BMI cannot expressed the body fat mass, also the endocrinal and metabolic alternations that associate or as resulted from obesity is a controversial matter, therefore the aim of the study is to estimate some blood biomarkers in obese healthy females with different body fat deposition, also to explore the correlation between the distribution type of body adiposity with the tested blood parameters.

## Materials and methods

In this study the participants consist of premenopausal females (n=148) with aged (39-43 years) and various body weight. The participants were dividing according to body mass index (BMI) to normal body weight (BMI =18.5-24.9 kg/m<sup>2</sup>, n=65) and obese (BMI ≥ 25 kg/m<sup>2</sup>, n=83). The obese females were classified to central obesity (n=55) and peripheral obesity (n=28) according to WC and WHR. The study protocol was following according to the ethical committal of Biology department - Science-collage. Basing on questioners, physical examination and medical history, the participants consider with good health. The questioners included (social status, healthy status, diseases, diet type, allergy, smoking, drugs, hormonal therapy, polycystic ovary syndrome, life style, physical activity). The study was carried from May 2019 to January 2021.

### Anthropometric measurements

The general obesity was measured using the common formula of body mass index BMI= body weight (kg) / height<sup>2</sup> (m) [14]. Waist circumference (WC) was measure at the umbilicus region (cm), the hip circumference (HC) was measure at iliac ridge over the buttocks (cm), the neck circumference at the neck place (cm). All measured by plastic strip [15]. The WHR ratio by WC/HC, the WHtR ratio by WC/height. The central obesity was determined according to WC ≥ 80 cm, WHR ≥ 0.85 and a WHtR > 0.50; the peripheral obesity when WHR < 0.85 [16]. The percentage of body fat according to formula of American Navy: females =  $163.205 \times \log_{10}(\text{waist} + \text{hip} - \text{neck}) - 97.684 \times \log_{10}(\text{height}) - 78.387$  [17]. Using digital scale to measure the weight. The blood pressure was taken in sitting position.

### Blood sample

The venous blood samples (5ml) of the participants were taken in the morning after fasting (12 hours, at 8-10 Am). Blood specimens were left at room temperature to clot and get serum after centrifugation (3000rpm, 10 minutes). Serum was stored at -70°C in deep freeze for biochemical and hormonal test [18].

### Serum assays

Serum glucose and lipid profiles (triglycerides TG, total cholesterol T-ch, high density lipoprotein HDL and very low density lipoprotein VLDL) were measured by a commercial kits (Biolab, France). LDL according to formula:  $Tch - HDL - TG/5$  [19]. The insulin, thyroid stimulating hormone TSH, total testosterone and cortisol were measured by Elisa kits using COBAS 411e automated analyzer (Roche Diagnostics, Germany). Free triiodothyronine FT3 (Calbiotech, USA Elisa Kit), free thyroxine FT4 (Calbiotech, USA Elisa Kit), visfatin (abcam, USA Elisa Kit), interleukin 8 (Bio-Vender, UK Elisa Kit), C-reactive protein (Enzo, USA Elisa Kit). The homeostatic index of insulin resistance (HOM-IR) by the formula:  $[\text{glucose (mmol/l)} \times \text{insulin (pmol/l)}] \div 155$  [20].

### Statistical Analysis

The data were analyzed by ANOVA using t-test. The correlation by bivariate analysis (SPSS version 22). The least significant difference ( $p \leq 0.05$ ) was used to compare the means. The data expressed by means ± standard deviation SD.

### Results

According to BMI (table 1), the females participants were dividing into obese group and normal weight group. The using of anthropometric parameters WC, WHR and WHtR showed significant ( $p \leq 0.05$ ) increasing in these parameters in obese group 105.257, 0.858, 0.641 respectively compared to normal levels in normal weight group 74.360, 0.739, 0.452 respectively.

The Navy formula revealed that obese group showed higher body fat percentage (%BF) 50.550 % and higher fat mass 47.5 kg with significant ( $p \leq 0.05$ ) than the normal weight group 29.23%, 18.7 kg respectively (table 1). The lean mass between the two groups showed insignificant differences. The results of Navy formula showed that the normal body weight group had an average body fat.

The obese group display a significant elevation in blood pressure 130.119/85.268 mmHg compared to blood pressure in normal body weight group 118.5/78.901 mmHg.

The obese group showed two type of regional obesity, the central and peripheral obesity (table 2). The central obesity group characterized by significant ( $p \leq 0.05$ ) increasing in WHR and WHtR 0.921, 0.673 respectively, while in peripheral obesity group the values of WHR and WHtR were 0.795, 0.609 respectively. According to Navy formula the central obesity group have higher fat percentage 51.6% and fat mass 49.5 kg with significant ( $p \leq 0.05$ ) than the peripheral obesity group 49.5 % and 45.5 kg respectively. The lean body mass showed insignificant between the two groups 46.5 kg.

There was insignificant differences in systolic and diastolic pressure between the central and peripheral obesity.

The increasing of body weight in obese female group showed alternations in serum biochemical and hormonal parameters compared to normal weight female group (table 3). In obese group, there were significant ( $p \leq 0.05$ ) elevation in serum glucose 9.375 mmol/l and lipid profile TG 134.421 mg/dl, T-ch 196.47 mg/dl, LDL 128 mg/dl, VLDL 37.153 mg/dl, while there was a significant decreasing in HDL level 41.244 mg/dl compared to normal weight group.

Comparing to normal weight category, the obese females display significant ( $p \leq 0.05$ ) increasing in serum level of TSH 2.231  $\mu$ IU/ml and FT3 3.090 pg/ml, while showed insignificant differences in serum level of TF4 (1.807 ng/dl).

The obese group showed significant ( $p \leq 0.05$ ) elevation in serum cortisol, insulin and total testosterone hormones 883.740 nmol/l, 189.257 pmol/l, 0.578 ng/ml respectively than the normal weight female group. The mean values of HOMIR, visfatin, IL8 and CRP were significantly higher in obese group when compared to normal weight group (obese: 11.451, 35.721 ng/ml, 4.100 pg/ml, and 180.501 ng/ml respectively).

In comparison between the central and peripheral obesity groups (table 4), the central obesity group revealed significant ( $p \leq 0.05$ ) elevation in serum levels of glucose, lipid profile compared to the peripheral obesity group.

The female with central obesity showed significant ( $p \leq 0.05$ ) increasing in the levels of the tested hormones compared to females with peripheral obesity. The levels of FT3, cortisol and insulin were significantly high in central obesity (3.367 pg/ml, 947.192 nmol/l, 192.412 pmol/l respectively).

The concentrations of visfatin, IL8 and CRP were increased significantly ( $p \leq 0.05$ ) in central obesity (41.959 ng/ml, 4.768 pg/ml, 197.773 ng/ml respectively) compared to peripheral obesity (29.483 ng/ml, 3.432 pg/ml, 163.229 ng/ml respectively).

The elevation in serum glucose and insulin in central obesity group reflected in high level of HOMIR (11.855) with significantly ( $p \leq 0.05$ ) than the peripheral obesity group (11.047).

The serum level of TSH was elevated in both central and peripheral obesity with significant differences ( $p \leq 0.05$ ) between them (2.446  $\mu$ IU/ml, 2.016  $\mu$ IU/ml respectively). There were insignificant differences in serum levels of HDL, FT4 and total testosterone between the two groups.

The bivariate correlation analysis (table 5) display the significant ( $p \leq 0.05$ ) correlations among the tested variables. In central obesity group, the BMI correlated positively with VLDL and HOMIR. The WC have positive correlation with serum glucose level, while the WHR correlated positively with serum level of IL8. The serum level of HDL revealed negative correlation with cortisol and CRP, while it showed positive correlation

with total testosterone level. There was a positive correlation between TSH and FT3. The serum visfatin level correlated with the level of CRP in positive correlation.

In peripheral obesity group (table 5), there was a positive correlation between WC and serum glucose level. The WHR correlated negatively with serum TSH and positively with HOMIR. There was a negative correlation between level of HDL and level of IL8. The serum level of TSH correlated positively with both FT4 and HOMIR which they showed also a positive correlation between them. There was a positive correlation between serum total testosterone and CRP level.

**Table 1.** Anthropometric parameters in normal weight and obese females.

Parameters	Control group Normal weight (n=65) Mean± SD	Obese group (n=83) Mean± SD	P value Sig. p ≤ 0.05
Weight kg	64.962±12.890	94.359±18.321	0.000
Height cm	164.311±11.965	163.966±14.510	0.670
BMI kg/m <sup>2</sup>	23.75±2.890	35.169±1.582	0.000
WC cm	74.360±13.533	105.257±19.859	0.003
HC cm	100.523±8.561	122.802±11.303	0.025
WHR	0.739±0.011	0.858±0.051	0.012
WHtR	0.452±0.005	0.641±0.004	0.043
TC cm	54.392±4.011	91.272±6.783	0.001
NC cm	33.524±3.920	39.401±1.895	0.032
% BF	29.23±1.128	50.550±3.502	0.013
Body Fat category	average	obese	-
Fat mass kg	18.7±1.468	47.5±1.930	0.000
Lean mass kg	45.3±1.966	46.5±1.825	0.241
SBP mmHg.	118.500±6.713	130.119±9.545	0.010
DBP mmHg.	78.901±7.111	85.268±8.601	0.000

**Table 2.** Anthropometric parameters in obese females with central and peripheral obesity.

Parameters	Central obesity (n=55) Mean± SD	peripheral obesity (n=28) Mean± SD	P value Sig. p ≤ 0.05
Weight kg	96.703±4.098	92.015±2.560	0.033
Height cm	164.521±0.219	163.411±0.303	0.621
BMI kg/m <sup>2</sup>	35.701±.587	34.638±.733	0.372
WC cm	110.853±1.025	99.661±1.002	0.033
HC cm	120.252±11.511	125.351±7.701	0.048
WHR	0.921±.005	0.795±.008	0.022
WHtR	0.673±.009	0.609±.007	0.027
TC cm	82.024±1.515	100.520±1.616	0.034
NC cm	39.271±0.658	39.532±0.721	0.541
% BF	51.6±.622	49.5±.756	0.046
Body Fat category	obese	obese	-
Fat mass kg	49.5±0.128	45.5±0.321	0.0301
Lean mass kg	46.5±0.011	46.5±0.021	0.830
SBP mmHg.	130.298±0.199	129.940±0.138	0.590

DBP mmHg.	85.407±0.311	85.129±0.660	0.637
-----------	--------------	--------------	-------

**Table 3.** Serum biomarkers in normal weight and obese females.

Parameters	Control group (n=65) Normal weight Mean± SD	Obese group (n=83) Mean± SD	P value Sig. p ≤ 0.05
Glucose mmol/l	8.20±0.387	9.375±0.715	0.014
Triglycerides mg/dl	81.021±7.124	134.421±7.333	0.004
Total cholesterol mg/dl	174.311±10.238	196.47±9.131	0.011
HDL mg/dl	49.560±3.751	41.244±6.620	0.039
LDL mg/dl	109.0±5.140	128±7.894	0.021
VLDL mg/dl	26.590±0.330	37.153±1.822	0.005
TSH µIU/ml	1.679±0.065	2.231±0.061	0.044
FT3 pg/ml	3.010±0.007	3.090±0.008	0.048
FT4 ng/dl	1.810±0.011	1.807±0.007	0.101
Cortisol nmol/l	623.311±1.011	883.740±0.912	0.040
Insulin pmol/l	172.550±10.204	189.257±13.716	0.030
HOMI-IR	9.128±0.086	11.451±0.086	0.048
Total testosterone ng/ml	0.355±0.018	0.578±0.083	0.037
Visfatin ng/ml	14.906±0.036	35.721±0.187	0.028
IL8 pg/ml	1.874±0.015	4.100±0.516	0.034
CRP ng/ml	120.791±0.011	180.501±0.007	0.029

**Table 4.** Serum biomarkers in obese females with central and peripheral obesity.

Parameters	Central obesity (n=55) Mean± SD	Peripheral obesity (n=28) Mean± SD	P value Sig. p ≤ 0.05
Glucose mmol/l	9.550±0.040	9.201±0.228	0.010
Triglycerides mg/dl	149.321±14.756	119.521±11.166	0.044
Total cholesterol mg/dl	212.011±17.908	180.929±18.304	0.035
HDL mg/dl	40.172±4.906	42.317±5.422	0.257
LDL mg/dl	142.0±4.707	114.0±4.483	0.021
VLDL mg/dl	39.395±1.873	34.911±1.974	0.011
TSH µIU/ml	2.446±0.008	2.016±0.004	0.029
FT3 pg/ml	3.367±0.039	2.813±0.015	0.048
FT4 ng/dl	2.131±0.037	1.483±0.051	0.767
Cortisol nmol/l	947.192±0.015	820.288±0.444	0.035
Insulin pmol/l	192.412±9.017	186.102±10.273	0.019
HOMI-IR	11.855±0.020	11.047±0.022	0.009
Total testosterone ng/ml	0.606±0.037	0.550±0.356	0.452
Visfatin ng/ml	41.959±0.317	29.483±0.044	0.020
IL8 pg/ml	4.768±0.012	3.432±0.058	0.012
CRP ng/ml	197.773±0.002	163.229±0.008	0.025

**Table 5.** Observed significant correlations among variables in females with central and peripheral obesity.

The central obesity		The peripheral obesity	
Variables	Pearson Correlation Sig. $p \leq 0.05$	Variables	Pearson Correlation Sig. $p \leq 0.05$
BMI-VLDL	.909(*) .033	WC-Glucose	.963(**) .008
BMI-HOMIR	.918(*) .028	WHR-TSH	-.902(*) .037
WC-Glucose	.981(**) .003	WHR-HOMIR	.956(*) .011
WHR-IL8	.923(*) .025	HDL-IL8	-.880(*) .049
HDL-Cortisol	-.969(**) .006	TSH-FT4	.925(*) .024
HDL-Testosterone	.985(**) .002	TSH-HOMIR	.947(*) .014
HDL-CRP	-.913(*) .030	FT4-HOMIR	.903(*) .036
TSH-FT3	.967(**) .007	Testosterone-CRP	.883(*) .047
Visfatin-CRP	.973(**) .005	-	-

### Discussion

The identifying obesity in population is an important matter to estimate the metabolic syndrome associated with fat deposit. The indirect method to evaluate the total body fat is using different anthropometric parameters [21]. Some digital or radial techniques were developed to determine the body composition, but characterized with high cost and show limitations in availability and usage [22]. According to these restrictions, we used in this study various anthropometric parameters in addition to BMI to classify obesity in health females with aged 39-43 years. The general diagnosis of obesity in individual is using the BMI, this parameter is establish the popular obesity but it consider useless to reveal the body fat distribution ,therefore adding another parameters including WC, WHR and WHtR is useful to determine the regional obesity. In the current study, the group of obese participant showed a central obesity ( $WHR \geq 0.85$ ) that characterized by excessive fat depots in the abdominal area, while the other group of obese participants showed peripheral obesity ( $WHR < 0.85$ ) that distinguish by excessive fat accumulation around the hip and the thighs. The measuring of WC consider a good index for central obesity detection, while the WHR and WHtR are essential measurements with BMI [15]. The premenopausal women in middle age showed higher visceral fat deposition compared to young women [23]. The small muscle mass in the legs was associated with high ratio of WHR [24].

In this study we determined the percentage of body fat by using a formula of American Navy. This formula includes sex, age and neck circumference in addition to all other anthropometric parameters for measurement. In this study, the formula results show that the obese healthy females group have a high fat percentage, but their lean mass was the same to the lean mass of normal weight females, therefore this formula indicate that the obesity in obese group related to high percentage of fat and the weight again is result from fat deposit. In normal body weight group in spite of normal levels of all anthropometric measurement, the group showed mild percentage of body fat that classified under average fat. From these results we can give an impression of the importance of this formula to detect body fat percentage and body fitness degree even so in normal body weight category. The Navy formula is simple, suitable, certain and inexpensive method for measure body composition [25, 26] and used as one of prevention or treatment programs in obesity monitoring and its related

complications [27]. The formula can be use any time of the day regardless of immediate food intake and physical activity without fluctuation in results [28]. The Navy formula give similarities in results with the results of using bioelectrical analysis for body composition measurement in obese and normal adults for both gender and recommended to use as screening tools [29].

The elevation of blood pressure in obese females may related to structural alternations in kidney that effect on nephron activity by renal fat deposition [30] or related to alternations in aldosterone secretion and angiotensin II [31].

The study results showed that the obese females particularly with central obesity had dyslipidemia compared to normal weight females. The enlargement of abdominal fat cause disturbing in lipid profile that increased lipolytic activity of adipose tissue which in turn elevate TG production and transfer it to the liver. This will enhance VLDL synthesis in the liver and increase the circulating TG. In the same time, the inhibition of lipoprotein lipase in adipose tissue and skeletal muscles also cause hypertriglyceridemia. These events lead to exchange cholesteryl esters from VLDL to HDL and LDL, and hydrolyze TG by hepatic lipase result in producing a small dense of LDL and a low HDL concentration [32].

In the current study, the increasing in glucose and insulin levels were observed in both central and peripheral obesity females while the HOMI was higher in central obese females than peripheral, this suggested that the central obesity contributed to accelerate developing of insulin resistance by disturbing in lipid profiles and as shown from the positive correlation between the WC and the serum glucose level. The high circulating free fatty acids resulted from lipolytic activity will enhance gluconeogenesis that caused insulin resistance [33]. The metabolites of fatty acids like acyl- coenzyme, diacylglycerol and ceramides impair insulin signaling [34]. The high flux of free fatty acids to the liver during hyperglycemia will increased TG synthesis and decreased insulin clearance, that lead to hyperinsulinemia and insulin resistance [35].

The WHR is associated with increased visceral fat deposit that expressed by central obesity which is caused alternations in metabolic profiles [36]. The visceral fat is accompanied with insulin resistance, dyslipidemia and inflammation [37]. The lipolytic activity in adipocytes in obesity condition increased macrophage accumulation in adipose tissue that increased production of pro- inflammatory proteins like IL8, IL6 and visfatin [38, 39], and lowering the angiogenic ability and cause hypoxia which in turn stimulated metabolic alternations and diseases occurrence [40]. Also the risk factors of metabolic diseases is increased with accumulation of abdominal visceral fat and subcutaneous fat, independently of total body fat [41]. The metabolic alternations in obese females with central obesity is related to incidence of insulin resistance [15].

In this study the high level of thyroid stimulating hormone in obese females may effect on body composition by affecting on thermogenesis and energy expenditure [42, 43]. The inflammatory cytokines like IL6 and 8 is resulted from the fat accumulation in obesity caused morphological and functional changes in thyroid gland by varying the permeability of gland blood vessels [44, 45], impact on iodide uptake by thyroid cells, or inhibit the mRNA expression of symporter sodium/iodide [45]. In adult women and men there is a positive correlation between BMI, WC and serum TSH, FT3 and FT4 [46]. The overweight person have high level of TSH and low level of FT4, while obese person have high level of TSH and low ratio FT3/FT4, suggest the thyroid effect on body weight [10]. The hypothyroidism cause insulin resistance even if the thyroid stimulating hormone TSH is at the normal level range [47]. It found that the visceral body fat reduction enhance the thyroid parameters than the reduction in subcutaneous body fat [48].

The study results display that the obese females had higher cortisol level than the normal weight females, this may explained by alternations in glucocorticoids activities in abdominal obesity [49], or the elevation of serum cortisol in central obesity group of this study may explained by the increasing in blood cortisol lead to visceral fat accumulation and increased central obesity [50]. The high expression of glucocorticoids receptor in visceral fat made it hypersensitive to cortisol that cause fat accumulation [51]. The visceral adipose tissue contain 11 $\beta$ -hydroxy steroid dehydrogenase I that activate the cortisone (inactive) to cortisol (active) that may contribute to regulate cortisol level in blood circulation [52].

From the study results, the high level of total testosterone in obese females, may related to the obese females with abdominal obesity showed hyper-androgenism, as the adipose tissue act as intracrine site for androgen synthesis (androstenedione convert to testosterone)[53]. The hyperinsulinemia caused changes in levels of sex hormones include androgen, estrogen and sex hormone binding globin [54]. The premenopausal overweight women showed high total and free testosterone level [55]. The obese women with insulin resistance display low levels of estradiol and sex hormone binding globin [56].

From the study results, the increasing in visfatin level in obese females with central obesity may related that the visfatin level is associated with visceral fat not subcutaneous fat in obese subjects [57]. The obese women showed high serum of visfatin level compared to normal weight women [58]. Visfatin secretion increased with increased adiposity [59] and its level correlate positively with insulin resistance, WC and IL6 in obesity [60, 61]. The obese women with insulin resistance showed elevated level of hormone visfatin [56].

The elevation of inflammatory biomarkers including IL8 and CRP in obese females especially with central obesity possibly related that the excessive size of adipocytes caused decreased in anti-inflammatory adipokines secretion from adipose tissue and enhance inflammatory process in addition to hyperglycemia caused elevation in IL8 [62]. The WHR is associated with increased visceral fat depots that expressed by central obesity which is caused alternation in metabolic profiles[36], these association was shown in our results in which the WHR correlated positively with IL8. The free fatty acids intermediates activate the pro-inflammation pathway include serine kinase that stimulate the IL8 secretion that promote secretion of C-reactive protein [63]. The process of chronic-low grade inflammation in obesity play important role in metabolic syndrome development, central obesity, insulin resistance, dyslipidemia, and increased oxidative stress by production of reactive oxygen species [64, 65].

### Conclusion

Monitoring the metabolic changes associated with obesity increase demand to use reliable and low cost measurement tool, therefore, in this study the using of Navy formula was useful to detect differences in fat ratio and fat mass between the regional obesity groups. In obese healthy females, the alternations in biochemical profiles was associated with changes in some hormonal and inflammation biomarkers which were characterized in central obesity group.

### References

- [1] Goryakin, Y., Lobstein, T., James, W. P. T., & Suhrcke, M. (2015). The impact of economic, political and social globalization on overweight and obesity in the 56 low and middle income countries. *Social Science & Medicine*, 133, 67-76.
- [2] González-Muniesa P., Martínez-González M.A., Hu F.B., Després J.P., Matsuzawa Y., Loos R.J.F., Moreno L.A., Bray G.A., Martínez J.A. (2017). Obesity. *Nat Rev Dis Primers*, 3:17034.
- [3] Blouin, K., Boivin, A., & Tchernof, A. (2008). Androgens and body fat distribution. *The Journal of steroid biochemistry and molecular biology*, 108(3-5), 272-280.
- [4] James, W. P. T. (2008). The epidemiology of obesity: the size of the problem. *Journal of internal medicine*, 263(4), 336-352.
- [5] Demerath, E.W., Reed, D., Rogers, N., Sun, S.S., Lee, M., Choh, A.C., Couch, W., Czerwinski, S.A., Chumlea, W.C., Siervogel, R.M. and Towne, B. (2008). Visceral adiposity and its anatomical distribution as predictors of the metabolic syndrome and cardiometabolic risk factor levels. *The American journal of clinical nutrition*, 88(5), pp.1263-1271.
- [6] Garaulet, M., Perez-Llamas, F., Fuente, T., Zamora, S. and Tebar, F.J. (2000). Anthropometric, computed tomography and fat cell data in an obese population: relationship with insulin, leptin, tumor necrosis factor-alpha, sex hormone-binding globulin and sex hormones. *European Journal of Endocrinology*, 143(5), pp.657-666.



- [7] Duclos M., Marquez Pereira P., Barat P., Gatta B., Roger P. (2005). Increased cortisol bioavailability, abdominal obesity, and the metabolic syndrome in obese women. *Obes Res*, 13:1157–1166.
- [8] Ward A.M., Fall C.H., Stein C.E., Kumaran K., Veena S.R., Wood P.J., Syddall H.E., Phillips D.I. (2003). Cortisol and the metabolic syndrome in South Asians. *Clin Endocrinol (Oxf)*, 58:500–505.
- [9] Zhang, H., & Sairam, M. R. (2014). Sex hormone imbalances and adipose tissue dysfunction impacting on metabolic syndrome; a paradigm for the discovery of novel adipokines. *Hormone molecular biology and clinical investigation*, 17(2), 89-97.
- [10] Xu, R., Huang, F., Zhang, S., Lv, Y., & Liu, Q. (2019). Thyroid function, body mass index, and metabolic risk markers in euthyroid adults: a cohort study. *BMC endocrine disorders*, 19(1), 1-9.
- [11] Mastaglia, S. R., Solis, F., Bagur, A., Mautalen, C., & Oliveri, B. (2012). Increase in android fat mass with age in healthy women with normal body mass index. *Journal of Clinical Densitometry*, 15(2), 159-164.
- [12] Bray, G.A., Jablonski, K.A., Fujimoto, W.Y., Barrett-Connor, E., Haffner, S., Hanson, R.L., Hill, J.O., Hubbard, V., Kriska, A., Stamm, E. and Pi-Sunyer, F.X. (2008). Relation of central adiposity and body mass index to the development of diabetes in the Diabetes Prevention Program. *The American journal of clinical nutrition*, 87(5), pp.1212-1218.
- [13] Snijder, M.B., Dekker, J.M., Visser, M., Bouter, L.M., Stehouwer, C.D., Yudkin, J.S., Heine, R.J., Nijpels, G. and Seidell, J.C. (2004). Trunk fat and leg fat have independent and opposite associations with fasting and postload glucose levels: the Hoorn study. *Diabetes care*, 27(2), pp.372-377.
- [14] WHO: World Health Organization (2006). BMI classification. [http://www.who.int/bmi/index.jsp?intro\\_page=intro\\_3.html](http://www.who.int/bmi/index.jsp?intro_page=intro_3.html).
- [15] Aras, Ş., Üstünsoy, S., & Armutçu, F. (2015). Indices of central and peripheral obesity; anthropometric measurements and laboratory parameters of metabolic syndrome and thyroid function. *Balkan medical journal*, 32(4), 414.
- [16] World Health Organization (2011). Waist circumference and waist-hip ratio report of a WHO expert consultation, Geneva. Switzerland.
- [17] Peterson, D. D. (2015). History of the US Navy body composition program. *Military medicine*, 180(1), 91-96.
- [18] Bain, B. J., Bates I. & Laffan M.A. (2017). Dacie and Lewis Practical Hematology (12th edition), Elsevier, London. Neha Bhatnagar.585 p.
- [19] Friedewald, W.T.; Levy, R.I. and Fredrickson, D.S. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma; without the use of the preparative ultracentrifuge. *Clin. Chem.*, 18; 499-502.
- [20] Matthews, D. R., Hosker, J. P., Rudenski, A. S., Naylor, B. A., Treacher, D. F., & Turner, R. C. (1985). Homeostasis model assessment: insulin resistance and  $\beta$ -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*, 28(7), 412-419.
- [21] Canoy, D., Wareham, N., Luben, R., Welch, A., Bingham, S., Day, N., & Khaw, K. T. (2006). Serum lipid concentration in relation to anthropometric indices of central and peripheral fat distribution in 20,021 British men and women: results from the EPIC-Norfolk population-based cohort study. *Atherosclerosis*, 189(2), 420-427.
- [22] Glickman SG, Marn CS, Supiano A, Denget DR. (2004). Validity and reliability of dual-energy X-ray absorptiometry for the assessment of abdominal adiposity. *J Appl Physiol*, 97:509-514.
- [23] Pascot A., Lemieux I., Prud'homme D., Tremblay A., Nadeau A., Couillard C., Bergeron J., Lamarche B., Després J.P. Reduced HDL (2001). particle size as an additional feature of the atherogenic dyslipidemia of abdominal obesity. *J Lipid Res* 42: 2007–2014.

- [24] Yusuf, S., Hawken, S., Ounpuu, S., Bautista, L., Franzosi, M.G., Commerford, P., Lang, C.C., Rumboldt, Z., Onen, C.L., Lisheng, L. and Tanomsup, S. (2005). Obesity and the risk of myocardial infarction in 27 000 participants from 52 countries: a case-control study. *The Lancet*, 366(9497), pp.1640-1649.
- [25] Macias, N., Alemán-Mateo, H., Esparza-Romero, J., & Valencia, M. E. (2007). Body fat measurement by bioelectrical impedance and air displacement plethysmography: a cross-validation study to design bioelectrical impedance equations in Mexican adults. *Nutrition Journal*, 6(1), 1-7.
- [26] Wang, L., Hui, S. S. C., & Wong, S. H. S. (2014). Validity of bioelectrical impedance measurement in predicting fat-free mass of Chinese children and adolescents. *Medical science monitor: international medical journal of experimental and clinical research*, 20, 2298.
- [27] Muralidhara, D. V., & Pui, C. (2015). An evaluation of overweight/obesity and body fat status in medical students. *Integr Obes Diabetes*, 1(1), 7-10.
- [28] Androutsos, O., Gerasimidis, K., Karanikolou, A., Reilly, J. J., & Edwards, C. A. (2015). Impact of eating and drinking on body composition measurements by bioelectrical impedance. *Journal of human nutrition and dietetics*, 28(2), 165-171.
- [29] Shaheen, A., Javed, N., Azam, F., Liaquat, A., Khan, M., Alam, S. M., & Mumtaz, S. (2019). Comparison of bioelectrical impedance and navy seal formula to measure body composition in medical students. *Cureus*, 11(5).
- [30] Hall, J.E. (2003). The kidney, hypertension, and obesity. *Hypertension*, 41: 625–633.
- [31] Nagase, M., Yoshida, S., Shibata, S., Nagase, T., Gotoda, T., Ando, K., & Fujita, T. (2006). Enhanced aldosterone signaling in the early nephropathy of rats with metabolic syndrome: possible contribution of fat-derived factors. *Journal of the American Society of Nephrology*, 17(12), 3438-3446.
- [32] Berndt, J., Klötting, N., Kralisch, S., Kovacs, P., Fasshauer, M., Schön, M. R., ... & Blüher, M. (2005). Plasma visfatin concentrations and fat depot-specific mRNA expression in humans. *Diabetes*, 54(10), 2911-2916.
- [33] Ritchie SA., Connell J.M. (2007). The link between abdominal obesity, metabolic syndrome and cardiovascular disease. *Nutr Metab Cardiovasc Dis*, 17:319–326.
- [34] Schenk, S., Saberi, M., & Olefsky, J. M. (2008). Insulin sensitivity: modulation by nutrients and inflammation. *The Journal of clinical investigation*, 118(9), 2992-3002.
- [35] Tourniaire, F., Romier-Crouzet, B., Lee, J. H., Marcotorchino, J., Gouranton, E., Salles, J., ... & Landrier, J. F. (2013). Chemokine expression in inflamed adipose tissue is mainly mediated by NF- $\kappa$ B. *PLoS one*, 8(6), e66515.
- [36] Van Gaal, L. F., Mertens, I. L., & Christophe, E. (2006). Mechanisms linking obesity with cardiovascular disease. *Nature*, 444(7121), 875-880.
- [37] Despres, J. P. (2006). Intra-abdominal obesity: an untreated risk factor for Type 2 diabetes and cardiovascular disease. *Journal of endocrinological investigation*, 29(3), 77.
- [38] Fain, J. N. (2010). Release of inflammatory mediators by human adipose tissue is enhanced in obesity and primarily by the nonfat cells: a review. *Mediators of inflammation*, 2010.
- [39] Ouchi, N., Parker, J. L., Lugus, J. J., & Walsh, K. (2011). Adipokines in inflammation and metabolic disease. *Nature reviews immunology*, 11(2), 85-97.
- [40] Hocking, S., Samocho-Bonet, D., Milner, K. L., Greenfield, J. R., & Chisholm, D. J. (2013). Adiposity and insulin resistance in humans: the role of the different tissue and cellular lipid depots. *Endocrine reviews*, 34(4), 463-500.
- [41] Tchernof, A., & Després, J. P. (2013). Pathophysiology of human visceral obesity: an update. *Physiological reviews*.
- [42] Topsakal, S., Yerlikaya, E., Akin, F., Kaptanoglu, B., & Erürker, T. (2012). Relation with HOMA-IR and thyroid hormones in obese Turkish women with metabolic syndrome. *Eating and Weight Disorders-Studies on Anorexia, Bulimia and Obesity*, 17(1), e57-e61.

- [43] Tarcin, O., Abanonu, G. B., Yazici, D., & Tarcin, O. (2012). Association of metabolic syndrome parameters with TT3 and FT3/FT4 ratio in obese Turkish population. *Metabolic syndrome and related disorders*, 10(2), 137-142.
- [44] Longhi, S., & Radetti, G. (2013). Thyroid function and obesity. *Journal of clinical research in pediatric endocrinology*, 5(Suppl 1), 40.
- [45] Fontenelle, L.C., Feitosa, M.M., Severo, J.S., Freitas, T.E.C., Morais, J.B.S., Torres-Leal, F.L., Henriques, G.S. and do Nascimento Marreiro, D., 2016. Thyroid function in human obesity: underlying mechanisms. *Hormone and Metabolic Research*, 48(12), pp.787-794.
- [46] Kitahara, C. M., Platz, E. A., Ladenson, P. W., Mondul, A. M., Menke, A., & de González, A. B. (2012). Body fatness and markers of thyroid function among US men and women. *PloS one*, 7(4), e34979.
- [47] Baeza, I., Vigo, L., de la Serna, E., Calvo-Escalona, R., Merchán-Naranjo, J., Rodríguez-Latorre, P., ... & Castro-Fornieles, J. (2017). The effects of antipsychotics on weight gain, weight-related hormones and homocysteine in children and adolescents: a 1-year follow-up study. *European child & adolescent psychiatry*, 26(1), 35-46.
- [48] [Mukherjee, S., Deoghuria, D., & Mukhopadhyay, D. (2019). Relationship of thyroid function with body fat, waist circumference, sub-cutaneous and visceral adipose tissue in eu-thyroid female subjects. *International Journal of Research in Medical Sciences*, 7(3), 822.
- [49] Vicennati, V., & Pasquali, R. (2000). Abnormalities of the hypothalamic-pituitary-adrenal axis in nondepressed women with abdominal obesity and relations with insulin resistance: evidence for a central and a peripheral alteration. *The Journal of Clinical Endocrinology & Metabolism*, 85(11), 4093-4098.
- [50] Kyrou, I., & Tsigos, C. (2009). Stress hormones: physiological stress and regulation of metabolism. *Current opinion in pharmacology*, 9(6), 787-793.
- [51] Hirata, A., Maeda, N., Nakatsuji, H., Hiuge-Shimizu, A., Okada, T., Funahashi, T., & Shimomura, I. (2012). Contribution of glucocorticoid–mineralocorticoid receptor pathway on the obesity-related adipocyte dysfunction. *Biochemical and biophysical research communications*, 419(2), 182-187.
- [52] Schorr, M., Lawson, E. A., Dichtel, L. E., Klibanski, A., & Miller, K. K. (2015). Cortisol measures across the weight spectrum. *The Journal of Clinical Endocrinology & Metabolism*, 100(9), 3313-3321.
- [53] Quinkler, M., Sinha, B., Tomlinson, J. W., Bujalska, I. J., Stewart, P. M., & Arlt, W. (2004). Androgen generation in adipose tissue in women with simple obesity—a site-specific role for 17 $\beta$ -hydroxysteroid dehydrogenase type 5. *Journal of Endocrinology*, 183(2), 331-342.
- [54] Wang, X., Xian, T., Jia, X., Zhang, L., Liu, L., Man, F., Zhang, X., Zhang, J., Pan, Q. and Guo, L. (2017). A cross-sectional study on the associations of insulin resistance with sex hormone, abnormal lipid metabolism in T2DM and IGT patients. *Medicine*, 96(26).
- [55] Stanikova, D., Zsido, R.G., Luck, T., Pabst, A., Enzenbach, C., Bae, Y.J., Thiery, J., Ceglarek, U., Engel, C., Wirkner, K. and Stanik, J. (2019). Testosterone imbalance may link depression and increased body weight in premenopausal women. *Translational psychiatry*, 9(1), pp.1-12.
- [56] Alnowihi, S. M., Al Doghathier, H. A., & Osman, N. N. (2020). Serum visfatin concentration and its relationship with sex hormones in obese Saudi women. *International Journal of Health Sciences*, 14(3), 9.
- [57] Sandeep, S., Velmurugan, K., Deepa, R., & Mohan, V. (2007). Serum visfatin in relation to visceral fat, obesity, and type 2 diabetes mellitus in Asian Indians. *Metabolism*, 56(4), 565-570.
- [58] Zahorska-Markiewicz, B., Olszanecka-Glinianowicz, M., Janowska, J., Kocelak, P., Semik-Grabarczyk, E., Holecik, M., Dąbrowski, P. and Skorupa, A. (2007). Serum concentration of visfatin in obese women. *Metabolism*, 56(8), pp.1131-1134.
- [59] Taşkesen, D., Kirel, B., & Us, T. (2012). Serum visfatin levels, adiposity and glucose metabolism in obese adolescents. *Journal of Clinical Research in Pediatric Endocrinology*, 4(2), 76.

- [60] Olszanecka-Glinianowicz, M., Kocęlak, P., Nylec, M., Chudek, J., & Zahorska-Markiewicz, B. (2012). Circulating visfatin level and visfatin/insulin ratio in obese women with metabolic syndrome. *Archives of medical science: AMS*, 8(2), 214.
- [61] Li, R. Z., Ma, X. Y., Hu, X. F., Kang, S. X., Chen, S. K., Cianflone, K., & Lu, H. L. (2013). Elevated visfatin levels in obese children are related to proinflammatory factors. *Journal of Pediatric Endocrinology and Metabolism*, 26(1-2), 111-118.
- [62] Esposito, K., Pontillo, A., Ciotola, M., Di Palo, C., Grella, E., Nicoletti, G., & Giugliano, D. (2002). Weight loss reduces interleukin-18 levels in obese women. *The journal of clinical endocrinology & metabolism*, 87(8), 3864-3866.
- [63] Rocha, V. Z., & Libby, P. (2009). Obesity, inflammation, and atherosclerosis. *Nature Reviews Cardiology*, 6(6), 399.
- [64] Shaker, O., El-Shehaby, A., Zakaria, A., Mostafa, N., Talaat, S., Katsiki, N. and Mikhailidis, D.P. (2011). Plasma visfatin and retinol binding protein-4 levels in patients with type 2 diabetes mellitus and their relationship to adiposity and fatty liver. *Clinical biochemistry*, 44(17-18), pp.1457-1463.
- [65] Tiganis, T. (2011). Reactive oxygen species and insulin resistance: the good, the bad and the ugly. *Trends in pharmacological sciences*, 32(2), 82-89.