

Indo – Asian Journal of Multidisciplinary Research (IAJMR) ISSN: 2454-1370

# EFFECT OF ACRYLAMIDE ON THYROID AND LIVER FUNCTIONS IN ADULT MALE RATS

Nawras A. Alwan\*, Jassim M. A. Alkalby and Eman Aboud Al-Masoudi,

Department of Physiology, Pharmacology and Chemistry, College of Veterinary Medicine, University of Basrah, Basrah, Iraq.

#### Abstract

The present study was carried out to investigate the effects of acrylamide (ACR) on thyroid and liver functions and histopathological changes in adult male rats. Twenty adult male rats were used in the present study. The animals were acclimatized for two weeks before beginning of the experiment. The animals were divided randomly into two equal groups. The 1<sup>st</sup> group (control) administrated distal water orally by gavage, the 2<sup>nd</sup> was given ACR at a dose of 5 mg/kg BW/day for 45 days. The results revealed a significant increase in serum thyroxin (T<sub>4</sub>), triiodothyronine (T<sub>3</sub>) concentrations and thyroid stimulating hormone (TSH) was observed in ACR group. Moreover, a significant increase in serum levels of AST, ALT and ALP enzymes in ACR-group. Histopathological study showed the irregular size and shape of thyroid follicles and vacuolation of colloid and liver section appeared congestion of central vein, flattening and vacuolation of hepatocytes in ACR treated group.

Key words: Acrylamide, Thyroid, Liver and Male rats.

#### **1. Introduction**

Acrylamide is an odorless, white crystalline solid at room temperature and has chemical formula:  $C_3H_5NO$  and structure  $H_2C=CHCONH_2$ , and it is a water soluble substance (Asha et al., 2008; Schwend et al., 2009). The orally consumed acrylamide is absorbed into the general circulation, then distributed to different organs and reacts with DNA, neurons, essential enzymes and hemoglobin and causes different toxic effect (Rayburn and Friedman, 2010). Acrylamide and polyacrylamide used in many applications such as: plastic, aesthetic surgeries, cosmetic industries, opthalmic operations, laboratory processes and oil recovery processes (Schwend et al., 2009).

\**Corresponding author*: **Nawras A. Alwan** *Received*: 20.05.2016; *Revised*: 03.06.2016; *Accepted*: 26.06.2016. Acrylamide in the small amounts used in sugar beet juice clarification, foundry sand, printing ink emulsion stabilizers, thickening agents (Magdy *et al.*, 2013), scientific application of acrylamide as: water purification, gel electrophoresis and sewage treatment for agricultural sprays, textile printing paste and water retention aids, adhesive and binders for seed coating, monomer acrylonitrite used as a raw material in many manufacturing of acrylic fiber, plastics and synthetic rubbers (Woutersen, 1998).

Thyroid gland secretes thyroid hormones ( $T_4$  and  $T_3$ ). Thes hormones remain fused with thuroglobulin which was stored in the colloid of the thyroid follicles until release into the circulation in response of the thyroid gland to thyroid stimulating hormone (TSH). More than 90



% of  $T_4$  and  $T_3$  are bound to the protein and small amount of them still free in the circulation and a small amount of them still free in the circulation which are physiologically active form and responsible for regulation of TSH secretion from pituitary through the feedback mechanism (Tietz, 1987). In adult thyroid hormones are playing important role in protein, lipid and carbohydrate metabolism and in heat generation (Kim, 2008), many endogenous or exogenous agents affected thyroid gland function resulted in various subclinical effects (Surks et al., 2004) or clinical manifestation (Dallaire *et al.*, 2009). Liver is a organ for detoxifying chemicals vital of the body and play important role in the regulation of the concentration of most metabolites, mainly glucose and amino acids, it has an ability in synthesizing, souring, secreting, regulating. transforming and breaking down of several materials in the body (Akharaiyi et al., 2012). Serum AST and ALT activity are used as most sensitive biomarkers in diagnosis of liver diseases (Pari and Kumar, 2002). The present study was aimed to investigate the effect of acrylamide administration on thyroid and liver functions and histopathological changes in adult male rats.

### 2. Materials and Methods

Acrylamide of 99.9 purity from sigma chemical Co. was used in the present study.

### **Experimental Animals**

Twenty adult healthy rats weighting  $(205.00\pm19.00)$  gram was used in this study. Rats were kept in the animal house for two weeks before beginning of the experiment for adaptation.

## **Experimental Design**

Twenty adult male rats randomly divided into two groups as the following:

**Group I (control):** Ten adult male rats orally administered detailed water (0.4 ml/animal) by Cavage daily for 45 days. **Group II:** Ten adult male rats were orally administered acrylamide (5 mg/Kg. B.W.) dissolved in 0.4 ml distilled water

by garbage daily for 45 days. At the end of the experiment ten animals of each group were anaestheized by chloroform and sacrificed. The blood samples were collected between 9:00 to 11:00 A.M. in order to minimize the diurnal variation of hormone levels by heart puncture of each rat and serum were separated for hormonal and biochemical tests and the following organs (thyroid and liver) were excised and maintained at buffered formalin 10 % neutral for histopathological study. Blood samples were collected from the heart via cardiac puncture of sacrificed animals using 5cc sterile syringe dropped in plain without anticoagulant tubes and serum samples were isolated from blood by centrifugation at 3000rpm for 15 min. and seperated in eppendroff tubes and stored at -20°C until used for hormonal and biochemical analysis.

674

## **Biochemical analysis**

The basic principle of an Enzyme linked immunosorbent is to use an enzyme to detect the binding of antigen (Ag) antibody (Ab). The enzyme converts a colorless substrate to a colored product, indicating the presence of Ag: Ab binding, the estimation of thyroid stimulating hormone (TSH), total thyroxine (tT4) and tetraiodthyronine (tT3) Concentrations described by Ma *et al.* (2006). Measurement of serum tetraiodthyronine concentration is generally regarded as a valuable tool in the diagnosis of thyroid dysfunction; kit was used (Monobind Inc. lake forest CA 92630, USA).

## Histopathological Study

Specimens from thyroid and liver were collected and preserved in 10% neutral buffered formalin and processed to obtain paraffin sections of 6 micron thickness, then stained with hematoxylin and eosin stain (Bancroft et al., 1996). Then the slide is uses for histolopathological examination under light microscope.

## **Statistical Analyses Method**



Statistical analyses were performed by the standard method. All the results were expressed as mean  $\pm$  standard deviation (S.D.). The mean of the all groups compared using independent t-test. 'P' value of less than 0.05 was considered to represent statistically significant change (Abo-Allam, 2003).

### 3. Results

The results presented in Table - 1 revealed a significant (P<0.05) decrease in serum concentrations of TSH, T4 and T3 in ACR-treated group compared with the control group.

### Table - 1: Effect ACR on serum concentrations of TSH, T4 and T3 of adult male rats (M±SD) (n=10)

Groups	TSH uIU/ml	T <sub>4</sub> ug/dl	T <sub>3</sub> ng/dl
G1 (Control)	$18.85 \pm 1.45^{a}$	9.38±0.86 <sup>a</sup>	2.13±1.37 <sup>a</sup>
G2 (ACR)	8.72±1.74 <sup>b</sup>	$4.63 \pm 0.91^{b}$	$1.03 \pm 0.55^{b}$
LSD	10.13	4.74	1.01

The values expressed in small letters mean significant differences in (P<0.05) levels.

The effect of ACR-treatment for 45 days on serum ALT, AST and ALP concentrations are presented in Table - 2. The results indicated that there were a significant (P<0.05) increased in serum ALT, AST and ALP concentrations in ACR-treated group compared with control.

### Table – 2: Effect of ACR on serum ALT, AST and ALP in adult male rats (M±SD) (n=10)

Groups	ALT (U/l)	AST (U/l)	ALP (U/l)
G1(Control)	$17.80 \pm 4.00^{b}$	22.00±3.41 <sup>b</sup>	$11.77 \pm 1.40^{b}$
G2 (ACR)	$26.96 \pm 5.10^{a}$	$38.58 \pm 4.69^{a}$	$18.63 \pm 1.40^{a}$
LSD	9.16	16.58	6.85

The values expressed in small letters mean significant differences at (P<0.05) levels

**Hisopathological examination: Thyroid gland -**Thyroid gland of control male rats showed the normal architecture composed of thyroid follicles of different sizes lined by single layer of cuboidal thyrocytes and parafollicular cells (c-cells) as shown in Figure - 1. While histopathological section in thyroid gland of adult male rats treated with ACR for 45 days revealed that thyroid follicles of irregular shapes and sizes with vacuolated non-homogenized colloid and non clear of thyrocytes lining the follicles and parafolilicular cells. **Liver** - The section of liver in control group showing normal hypatocytes, sinusoids emptied into a clear central veins hepatocytes radiate as hepatic plates from central vein (Figure - 3) while in Figure – 4. Liver of ACR group showed clear blood accumulation in the central vein and clear flatting of hepatocytes with obvious vacuolation and absence of sinusoids imported into the central vein.



Fig. (1): histopathological section in the thyroid gland of control male rats showed normal thyroid follicles (Tf), filled with homogenized colloid (C), lined by single layer of cuboidal thyrocytes parafollicular cells (cc) (H&E stain 400X)



Fig. (2): Thyroid gland of ACR-treated group showing irregular size and shape of thyroid follicles(Tf) with no



676

clear lining thyrocytes (tc), vacuolation of colloid (c) and prafollicular cells (cc) (H&E stain 400X).



Fig.(3): Liver of control group showing normal hypatocytes(HC), sinsoids (S) emptied into a clear central vein (CV) hepatocytes radiate as hepatic plates from central vein (H& E stain 400X).



Fig. (4): Liver of ACR group showed clear blood accumulation in the central vein (CV) and clear flatting of hepatocytes (HC) with obvious vaculation and absence of sinusoids embeted into the central vein (H&E stain 400X).

### 4. Discussion

The present results are consistent with El-Elaimy (2006) and Manna *et al.* (2006) who all indicated that adult male rats treated with ACR resulted in a significant decrease in serum  $T_3$  and  $T_4$  levels compared with control. While Bower *et al.* (2008) demonstrated that  $T_4$  level decreased only with high doses of ACR-treatment. Similarly Hamdy *et al.* (2012) showed that adult male rats treated with ACR in doses of 5, 10 and 15 mg/kg BW/day for 8 weeks cause an in serum  $T_3$  and  $T_4$ levels compared with controls. In contrast of the present study Khan *et al.* (1999) reported that meanling rats treated orally with ACR in doses of 2 mg and 15 mg/kg Bw/day for 2 or 7days showed a slight dose-dependent increase in plasma T<sub>4</sub> level and slight dose-dependent decrease in plasma TSH. Moreover Abd El-Mottaleb and Rashed (2008) reported that adult male treated with 75 and 150 mg/kg bw/day ACR for 28 days showed a significant decrease of serum  $T_3$  and  $T_4$  were recorded in group intoxicated with 150 mg/kg/day ACR compared with control. The reduction in thyroid hormone in the present study may be attributed to the iodine deficiency and then the thyroid gland fails to synthesize thyroid hormones and the animal suffered from hypothyroidism (Kaneko et al., 1997) or may be occurred as a secondary resulted due to pituitary insufficiency. The reduction of thyroid hormone levels may be resulted from increased deiodination and biliary excretion and eventually increase the rate of elimination of thyroid hormones from circulation as that occurred in pesticide toxicity (Farokhi and Taravati, 2014).

The data illustrated in Table - 2 represent the effect of ACR treatment for 45 days on serum aminotransferasr, aspartate alanine aminotranstferase and alkaline phosphatase levels. The present results are in accordance with Sawka et al. (2007); Sharma and Jain (2008); Tedor et al. (2011) and Gabr et al. (2010) who all found that adult male rats treated with ACR showed a significant increase in serum AST, activity compared with the ALT and ALP control group. The results in line with those of El-Bohi et al. (2011) who demonstrated that administration of ACR resulted in a significant increase in the activities of serum AST and ALT enzymes in adult male rats compared with the control group. The elevation of AST and ALT enzymes may result from the destruction of cellular lysosome which in turn lead to increase autolysis process. The present results are in the study made in adult albino mice by Sharma and Jain (2008) who found adult male mice treated with different doses of ACR (5, 15 and 25 mg/kg BW/dy) for two months cause a significant increase in serum AST, ALT and ALP activities



in all ACR- treated groups compared with control. The present results for the effect of ACR upon the serum liver enzymes activity also agreed with Siahkoohi et al. (2014) and Osman et al. (2016) who reported a significant increase in serum AST, ALT and ALP levels in ACR-treated adult male rats compared with control. The elevation in serum AST, ALT and ALP activities may be resulted from the increase permeability of hepatocyte cell membrane which may occur due to the toxic effect of ACR which cause cellular damage and in turn leakage of the enzymes outside the cellular structure to extracellular fluid. The elevation of serum AST, ALT and ALP activities may occurred due to the bipolar nature of ACR the CH<sub>2</sub>=CH-part may undergo hydrophobic interaction and the -CONH<sub>2</sub> part can form hydrogen bonds with the cell component, this property may enhance its ability to alter the structure of cell membarne structure and make the paranchymal cell membrane of the liver more permeable, ther by causing the active retention of enzymes and making them appear first in the extracellular space and then in the blood (Chinoy and Memon, 2001).

Histopathological studies, Thyroid gland -The microscopic examination of the thyroid gland ACR-treated rats showed avariable of pathological changes which in consistent with that obtained by Khan (1999) who showed a significant reduction in the colloid area and a significant increase in the hight of thyrocytes in ACR-treated rats.with (2 and 5 mg/kg BW/day) for 2 and 7 days. Similarly polychlorinated biphenyls treatment produces striking hypertrophy and hyperplasia of thyroid folliclar cells, along with a significant reduction in serum T<sub>4</sub> level in rats (Capen & Martin, 1989). However Osman et al. (2016) reported that some follicles were dilated and lined with flattened epithelium with colloid in their lumen and other follicles were reduced in size with few colloidal materials. This may explain the decrease in  $T_3$  and  $T_4$  in ACR treated male rats. In contrast of the results of present study Imai et al. (2008) did not observe any hisopathological changes in thyroid of males and females treated with ACR in

drinking water at (1.0, 2.1 and 4.4 mg/kg BW/ day) in males and 1.2, 2.5 or 4.9 mg/kg BW/day) in females for 12 weeks.

Liver - A microscopic examination of the liver section of male treated with ACR showed different pathological changes as show in figure (4) these results are accorded with Abd El-Mottaleb and Rashed (2008) and Allam et al. (2010) who reported that the liver of rats treated with low dose of ACR showed vacuolar degenerative changes of hepatocyes and blood vessel congestion. The significant increase in the levels of serum liver enzymes resulted from these pathological changes in the liver. Rahangadale et al. (2012) also revealed the liver of ACR-treated group appeared congestion and Kupper cell proliferation. While Khan et al. (2011) showed disruption of hepatic cords, infiltration of inflammatory cells in the central vein. vacuolation of hepatocytes and necrosis. On other hand liver rats treated with ACR showed frequent necrosis and bleeding indicated hypertrophy of nuclei, pyknotic nuclei, proliferation of sinusoidal bile duct and hemorrhage (Vasundhara, 2005; Nagao et al, 2007). The vacuilation of the hepatocytes cytoplasm may be occurring due to progressive ischemia, hypoxia and accumulation of lipid in the hepatocytes (Abdul-Hamid and Moustafa, 2005 and Moustafa and Abdul-Hamid, 2007).

### **5. References**

- Abd El-Mottaleb, E. M and Rashed, A. Y. 2008. Studies on Acrylamide Intoxication In Male Albino Rats. *Egyptian Journal of Competitive & Clinical Pathology*, 21 (4): 222 - 245.
- Abdul-Hamid, M and Moustafa, N. A. 2005. Effect of ethanol administration on the liver and kidney of rat newborns at different conditions. *Journal of German Society of Zoology*, 24: 267 - 294.
- Abo-Allam, R. M. 2003. Data statistical analysis using SPSS Program. 1<sup>st</sup> Ed. Publication for the Universities, Cairo.



- Akharaiyi, F. C., Boboye, B and Adetuyi, F. C. 2012. Antibacterial, phytochemical and antioxidant activities of the leaf extracts of *Gliricidia sepium* and *Spathodea campanulata. World Applied Sciences Journal*, 16 (4): 523 - 530.
- Allam, A. A., El-Ghareeb, A. W., Abdul-Hamid, M., Bakery, A. E., Gad, M and Sabri, M. 2010. Effects of prenatal and perinatal acrylamide on the biochemical and morphological changes in liver of developing albino rats. *Archives of Toxicology*, 84: 129 -141.
- 6) Asha, S., Renu, S. and Jyotsna, J. 2008. Biochemical changes in the liver of swiss albino mice orally exposed to acrylamide. *International Journal of Science and Technology*, 2 (03): 542 - 550.
- Bancroft, J. D and Stevens, A. 1996. The haematoxylin and eosin. Theory and practice of histological techniques. 4<sup>th</sup> ed, Ch 6, pp. 99 – 112. Churchill Livingstone, London, New York and Tokyo.
- Bower, K. S., Price, K. L., Sturdee, L. E., Dayrell, M., Dougherty, D. A and Lummis, S. C. 2008. 5-Fluorotryptamine is a partial agonist at 5-HT3 receptors, and reveals that size and electronegativity at the 5 position of tryptamine are critical for efficient receptor function. *European Journal of Pharmacology*, 4 (3): 291 - 297.
- 9) Capen, C. C and Martin, S. L. 1989. The effects of xenobiotics on the structure and function of thyroid follicular and C-cells. *Toxicology and Pathology*, 17: 266 293.
- Chinoy, N. J. and Memon, M. R. 2001. Beneficial effects of some vitamins and calcium on fluoride and aluminium toxicity of gastrocnemius muscle and liver of male mice. *Fluoride*, 34: 21 - 33.
- 11) Dallaire, F., Dewailly, E., Vezina, C., Muckle, G., Weber, J. P., Bruneau, S. and Ayotte, P. 2006. Effect of prenatal exposure to polychlorinated biphenyls on incidence of acute respiratory infections in preschool Inuit children. *Environmental Health Prospects*, 114: 1301 - 1305.

- 12) El-Bohi, K. M., Moustafa, G. G., El-Sharkawi, N. I and Sabik, L. M. 2011. Genotoxic effects of acrylamide in adult male albino rats liver. *Journal of American Science*, 7 (1): 1097 -1108.
- 13) EL-Elaimy, W. S. M. 2006. Biochemical studies on the toxic effect of dome insecticides on experimental animals. MD in Agricultural Science, Department of Agricultural Biochemistry, Faculty of Agriculture, Ain Shams University.
- 14) Farokhi, F. and Taravati, A. 2014. Pesticide exposure and thyroid function in adult male sprayers. *International Journal of Medical Investigation*, 3 (4): 127 - 132.
- 15) Gabr, S. A., Kamal, A. M and Taha, M. S. 2010. Modulation of acrylamide induced biochemical alterations in albino rats. *Isotope and Radiation Research*, 42 (2): 239 247.
- 16) Hamdy, S. M., Bakeer, H. M., Eskander, E. F and Sayed, O. N. 2012. Effect of acrylamide on some hormones and endocrine tissues in male rats. *Human Experimental Toxicology*, 31: 483 – 491.
- 17) Imai, T., Takami, S., Cho, Y. M., Hirose, M. and Nishikawa, A. 2008. A 12 week toxicological study 40 of orally administered acr in juvenile rats. The Toxicologist. Poster 480. Annual meeting of the Society of Toxicology. Seattle, Washington.
- 18) Kaneko, J. L., Harvery, T. W and Michael, L.
  B. 1997. Clinical biochemistry of domestic animals. 5<sup>th</sup> ed. Academic Press, Inc. San Diego, London, Boston, New York. Pp.: 54 65.
- 19) Khan, M. A. 1999. Changes in thyroid gland morphology after acute acrylamide exposure. *Toxicological Sciences*, 47: 151 – 157.
- 20) Khan, S. H., Butt, M. S., Sharif, M. K; Sameen, A., Mumtaz, S and Sultan, M. T. 2011. Functional properties of protein isolates extracted from stabilized rice bran by microwave, dry heat, and parboiling. *Journal* of Agriculture and Food Chemistry, 59: 2416– 2420.
- 21) Kim, K. 2005. Effect of subchronic acrylamide exposure on the expression of neuronal and



inducible nitric oxide synthase in rat brain. Journal of Biochemistry and Molecular Toxicology, 19: 162 - 168.

- 22) Ma, Q., Tipping, R. H and Boulet, C. 2006. Irreducible correlation functions of the matrix in the coordinate representation: Application in calculating Lorentzian half widths and shifts. *Journal of Chemistry and Physics*, 124: 014 - 019.
- 23) Magdy, F., Ali, H. A. and Kotb, A. 2013. Toxicity of bisphenol A and acrylamide and their combination in mature rats. M.V.SC. Zagazig University.
- 24) Manna, F., Abdel Wahhab, M., Ahmed, H. H. and Park, M. H. 2006. Protective role of paua and ginseng extract standardized with ginsenoside Rg3 against avrylamide induced neurotoxicity in rats. *Journal of Applied Toxicology*, 76 (3): 198 - 206.
- 25) Moustafa, N. A and Abdul Hamid, M. 2007. Protective effect of phytic acid against the microscopical changes induced by aflatoxin B1 in the liver and kidney of white rats. *Journal of German Society of Zoology*, 53: 1 -27.
- 26) Nagao, T.; Yuko, O. and Yoshio, F. 2007. Effects of trace acrylamide intake in wistar rats. *Journal of Ocean Science*, 56 (9): 501 -506.
- 27) Osman, M. A., Romeilah, R. M., Elgammal, M. H., Ramis, E. S. and Hasan, R. S. 2016. Sub-chronic toxicity of acrylamide in fried rice and preventive effect of grape leaves. *Asian Journal of Biochemistry*, 11: 68 81.
- 28) Pari, L. and Kumar, N. A. 2002. Hepatoprotective activity of *Moringa oleifera* on antitubercular drug induced liver damage in rats. *Journal of Medicine and Food*, 5: 171 – 177.
- 29) Rahangadale, S., Jangir, B. L., Patil, M., Verma, T., Bhandarkar, A., Sonkusale, P and Kurkure, N. 2012. Evaluation of protective effect of vitamin E on acrylamide induced testicular toxicity in Wistar rats. *Toxicology International*, 19 (2): 158 - 161.
- 30) Rayburn, J. R. and Friedman, M. 2010. Icysteine, N-acetyl-I-cysteine and glutathione

protect xanopus leavis embryos against acrylamide induced malformations and mortality in the frog embryo teratogenesis assay. *Journal of Agriculture and Food Chemistry*, 58 (20): 11172 - 111785.

- 31) Sawka, M. N., Burke, L. M., Eichner, E. R., Maughan, R. J., Montain, S. J and Stachenfeld, N. S. 2007. American College of Sports Medicine position stand. Exercise and fluid replacement. *Medicine and Science in Sports* and Exercise, 39 (2): 377 – 390.
- 32) Schwend, T., Moller, M., Schabacker, J., Ruppert, T. and Wink, M. 2009. Alkylation of adenosine deaminase and thioredoxin by acrylamide in human cell cultures. *Journal of Nature and Forest*, 64 (5 - 6): 447 - 453.
- 33) Sharma, A. and Jain, J. 2008. Effects of oral exposure of acrylamide on plasma levels of thyroid hormones and haematological parameters in the swiss albino mice. *Asian Journal of Experimental Science*, 22: 317 324.
- 34) Siahkoohi1, S., Anvari, M., Tafti M. A. and Hosseini Sharifabad, M. 2014. The effects of Vitamin E on the Liver Integrity of Mice Fed with Acrylamide Diet. *Iranian Journal of Pathology*, 9 (2): 89 - 98.
- 35) Surks, M. I., Ortiz, E., Daniels, G. H., Sawin, C. T., Col, N. F., Cobin, R. H., Franklyn, J. A. Hershman, J. M., Burman, K. D., Denke, M. A., Gorman, C., Cooper, R. S and Weissman, N. J. 2004. Subclinical thyroid disease: scientific review and guidelines for diagnosis and management. *JAMA*, 291 (2): 228 238.
- 36) Teodor, V., Cuciureanu, M., Slencu, B. G., Zamosteanu, N. and Cuciureanu, R. 2011.
  Potential protective role of selenium in ACR intoxication. A Biochemical Study. Standia Universitatis "Vasite Goldis"; 21 (2): 263 -268.
- 37) Tietz, N. M. 1987. Fundamentals of Clinical Chemistry. 3<sup>rd</sup> ed., W. B. Sanders Co.; Pp.: 584 - 595.
- 38) Totani, N., Yawata, M., Ojiri, Y and Fujioka, Y. 2007. Effect of trace acrylamide intake in Wister rats. *Journal of Ocean Science*, 56: 501 - 506.



- 39) Vasundhara, K. 2005. Characterization of rat glutathione S-transferases under the influence of methyl cholanthrene. Ph.D thesis, S. V. University, Tirupati, India.
- 40) Woutersen, R. A. 1998. Toxicological profile of acrylonitrite. *Toxicological Science*, 47 (2): 151 - 157.

