Effect of Irrigation Water Types and Spraying With Seaweeds on Vegetative Growth, Biochemical Attributes and Minerals Contents

of Ziziphus mauritiana and Citrus sinensis Seedlings

Abbas, Karima F.¹; Khawla H. Mohamad² and Salwa J. Fakher³

¹Kerbala University, College of Applied medical Science, Department of Environmental health.

²Basrah University, College of Agriculture, Department of Horticulture and Landscape. ³Basrah University, College of Agriculture, Department of Soil Sciences and Water Resources.

Received on: 18/9/2016	Accepted for publication on:	27/9/2016
-------------------------------	------------------------------	-----------

Abstract

This experiment aimed to investigate the effect of the different irrigation water sources and spraving with seaweeds on the seedlings of two plants one is saltsensitive (Citrus sinensis) and the other is salt-tolerant (Ziziphus mauritiana) through studying growth parameters, as well as biochemical attributes and minerals in leaves of the studied plants. Four treatments were used for irrigation water as follow: (W1)The Tap Water (control) $E.c = 1.2 \text{ dS.m}^{-1}$, (W2) Shatt al-Arab river (sub river) $Ec = 5.2 \text{ dS.m}^{-1}$, (W3) Shatt al-arab river (main) 6.2 dS.m⁻¹, (W4) The well water E.c = 8.1 dS.m^{-1} with 3 levels (A1, A2, A3) of seaweed treatments prepared from (Agrosign marine) in the concentration of (0, 2, 4 ml /L). The results explained that there is a decrease in all parameters in this study with increasing the salinity of irrigation water and this is represented in well water irrigation as compared with another water types, while the interaction withseaweeds treatments caused significant increases in the growth parameters, biochemical attributes and minerals contents. This promotion was associated with increasing the concentrations to (4 ml/L), Which gave 42.94 cm in plant height, 2.00 number of shoots, 24.61 in the number of leaves, 5.34 in stem diameter, 101.2 g in fresh weight and 19.02 g dry weight. On the other hand the interaction with seaweed extract increased significantly growth parameters, attributes biochemical and mineral content, it gave the highest value A3W1 which was 3.48 mg/g in the total chlorophyll and 73.82 mg/g in total carbohydrates, while the lowest at A1W4 which was 1.38 mg/g in the total chlorophyll and 42.92 mg / g in the total carbohydrates. The activities of the enzymes superoxide dismutase (SOD), peroxidase (POD) and catalase (CAT) have shown a marked increase when increasing the level of salinity in irrigation water as well as, the seaweeds extraction caused increasing in the content of these enzymes. The treatment (A1W4) gave the highest enzymes activities CAT, SOD, BOD which was 24.10, 21.83 and 7.93 u / g / f.w respectively.

Keywords: Irrigation water, salinity, seaweeds, minerals contents, antioxidant enzymes, Citrus sinensis, Ziziphus mauritiana

Introduction

Water is very necessary for growth and development of the plants. In fact, the global infrequency of water resource and increasing the salinity of soil and water which considered as one of the most important factors of irrigation water (Beltran,

1999) that appeared on the last years and it can be seen obvious in Iraq .It is known that the Downfall of Sumer civilization about 4000 years ago was caused by unsuitable agricultural techniques, that's led to soil salinization and descent in the agricultural yield in the area (Pitman and Lauchli, 2002). Currently, the decrease of water levels in Tigris, Euphrates and shatt-alarab rivers in Iraq create a big problem that's lead to use other sources like wells water. Water quality can vary from source to source. Three sources of water are commonly used in the region of this study: municipal water, rivers and well water. Well water frequently contains high levels of dissolved elements and classified as a saline water while thewater obtained from rivers or lakes commonly has a lower level of dissolved chemicals than well water. In general, salinity is considered as a one of the major severe abiotic stress factors that effects on plant metabolism by inducing decreases in soil water potential ,which leads to adisturbance in water uptake by roots. Overall, Plants vary greatly in their ability to tolerate salt in water and classified into the sensitive plants to the salinity in spite of the difference among the species for the range of salinity tolerance degree like citrus (Mass, 1996; Murkute et al., 2005) and halophytes, which are able to grow at high concentrations of salt, like Ziziphus (Flowers and Colmer, 2008).

Ziziphus mauritiana Lam (Rhamnaceae) has marked ability to tolerate salt stress and drought (Bhatt *et al.*, 2008), while the citrus (Rutacea), are especially more sensitive to salinity than other plant (Boman,

1993; Maas, 1993). Current studies have focused on finding substitute natural solutions that will work to prevent and reduce the risk of salinity on plant, one of these processors are adding the seaweeds (Agrosign marine), that stimulate plant growth (Zodape, 2001; Strick et. al., 2003). It contains all major and minor nutrients including macro and micro elements amino acids and fatty acids and plant growth regulator like auxin, gibberllins and cytokinin. Seaweed extracts have been reported to stimulate the growth and yield of plants, develop tolerance to environment stress. increase nutrient uptake from soil and enhance antioxidant properties (Rathore, 2009). This work aimed to investigate the effect of various kinds of irrigation water on tolerant plant Ziziphus mauritiana and sensitive plant Citrus sinensis and increasing the tolerance to salinity by spraying the seaweeds.

Materials and Methods

This study was conducted in the Horticulture Department, Agriculture College, Basrah University, Basrah, Iraq from 17 November to 17 April, 1916 .The experiment was designed according to completely randomized Block design (RCBD), as a factorial experiment with three factors (Irrigation water, Seaweed Liquid extraction and plant types) four time replication, The experiment consisted of 96 plastic pots (29cm diameter), containing one year -old seedlings of same size and length taken for both experimental plants previously characterized as salt-tolerant Ziziphus mauritiana cv. Tuffahi (P1) and salt-sensitive Citrus sinensis cv. local (P2), (an experimental unit was one plant in one pot). Irrigation water salinity treatment were applied as:

1. The tap water (control) (W1) Ec = 1.2 dS.m^{-1}

2. Shatt al-arab river (sub river) (W2) $Ec = 5.2 \text{ dS.m}^{-1}$

3. Shatt al-arab river (main) (W3) = 6.2 dS.m^{-1}

4. The well water (W4) Ec = 8.1 dS.m⁻¹

Seaweed liquid fertilizer was prepared from (Agrosign marine, it consist of organic matter extracted from seaweeds 12%, fatty acids group 65%, natural growth regulators 25 3%. potassium phosphor ppm, 2%,nitrogen 15%) with three levels (A1= 0, A2 = 2, A3= 4 ml /L), The solutions were sprayed once on the leaves in the early morning, every 20 days. At the end of experiment the plants were harvest and washed with distilled water then the morphological parameters were taken such us Plant height(cm), Number of shoots, number of leaves, stem diameter (mm), fresh and dry weight (gm). then dried by freeze dryer. Total chlorophyll was determined according to Goodwin (1976), total carbohydrates was determined according to Dobois et al., (1956), The shoot dry matter was grunden, Zn^{+2} and Fe^{+2} were determined by Atomic Absorption Spectrophotometer after digested the samples in HNO₃ and HCIO₄ as pointed out in A.O.A.C.(1984), the N content was determined by micro Kjeldahl method (Bremner and Mulvaney, 1982), k by flame photometer, and P by spectrophotometer, after digestion in H₂SO₄ and HCIO₄ (Jackson, 1958). antioxidant enzymes extraction was accomplished by weighing 0.3g from leaves were collected after 3 months were directly dry frozen. The frozen tissues were pestled in a cold mortar condition. A 6.0 mL solution containing 50 mM potassium phosphate buffer (pH 7.0), 0.1 mM EDTA, 4% polyvinyll phrolidone (PVP) and 0.2 mM ascorbic acid. The extract was centrifuged at 12,000 x g at 4 °C for 20 min for purification. The enzyme assays were represented in supernatant. The CAT activity was measured as the change in absorbance of the reaction mixture at 240 nm due to hydrogen peroxide reduction (Aebi, 1984). It was calculated by the coefficient for H₂O₂ at 240 nm (40 mM cm). Determination of peroxidase POD activity was determined according to Kar and Choudhuri (1987) method. Activity unit was calculated using the coefficient of absorbance for tetraguaiacol at 470 nm (22.6 mM). POD activity was expressed as enzyme units per gram fresh weight (U/g fw). Superoxide dismutase (SOD) activity was determined according to Giannopotitis & Ries (1977). The absorbance readings were taken at wave length of 560nm. One unit of SOD activity was defined as the enzyme activity that reduced the photo reduction of nitroblue tetrazolium to blue formazan by 50 %. SOD activity was expressed as enzyme units per gram fresh weight (U/g fw).

Soil separates	%	Texture
Sand	14.10	
Silt	48.40	Mixture
Clay	37.50	
Chemical characters	Units	Value
Electrical Conductivity(EC)	dS.m ⁻¹	2.8
рН	_	7.95
Organic matter	%	0.85
Soluble ions		
Soluble Ca ⁺⁺	gm.100gm ⁻¹	7.05
Mg ⁺⁺ Na ⁺	mmol.L ⁻¹	6.45
Na^+	$mmol \ .L^{-1}$	4.62
K ⁺	$mg. L^{-1}$	12.08
HCO3 ⁻	%	20.80
Cl	$mmol \ .L^{-1}$	7.32
SO4 ⁼	$mmol \ .L^{-1}$	9.86

 Table 1. Some physical and chemical characters for the studied soil.

Table 2. Some chemical characters for the studied irrigation water types .

	Units	(W1)	(W2)	(W3)	(W4)
pH	-	7.12	6.85	6.32	6.16
E.Ce	dS.m ⁻¹	1.2	5.2	6.2	8.1
Cl	mmol. L ⁻¹	4.71	13.15	19.23	23.16
Soluble Ca ⁺⁺	Mmol. L ⁻¹	4.91	14.82	21.78	24.64
Soluble Mg ⁺⁺	Mmol. L ⁻¹	7.85	22.57	25.97	32.85
Soluble K ⁺	Mmol. L ⁻¹	0.15	0.26	0.28	0.38
Soluble Na ⁺	Mmol. L ⁻¹	3.56	13.30	20.32	29.81

Results

Growth parameters: When evaluating the data of research recorded in Table (3), it is clear that different water types play an important role in plant growth with the four types of water. The results showed a significant decline in the average of growth parameters (plant height, number of shoots, number of leaves, stem diameter, fresh and dry weight). The decrease in growth parameters appeared with well water compared to tap water that having the highest value, and this effect was more obvious in the salt sensitive plant (citrus seedlings), while the Ziziphus seedlings were exceed in growth parameters values. On the other hand, there are many significant differences appeared when treated with seaweeds compared with untreated seedlings the highest value was represented by the treatment A3W1 which were 62.33 cm for plant height, 4.83 for number of shoots, 40.50 for number of leaves and 7.26 for stem diameter.

Biochemical attributes: The data of biochemical attributes are presented in Table 4 There were significant differences between treatments in total chlorophyll content and total carbohydrates, The highest value was found at treatment A3W1 for both plants which was 3.48 mg/g in the total chlorophyll and 73.82 mg /g in total carbohydrates, while the lowest at A1W4 which was 1.38 mg /g in the total chlorophyll and 42.92 mg / g in the total carbohydrates. On the other

hand, it is clear from the present results that the activities of the three important anti oxidative enzymes were increased significantly with increasing the salinity of irrigation water. It was observed that the activities of CAT, BOD and SOD were increased at well water, but the lowest value in tap Water. In addition, the seaweeds raised the activities of antioxidative enzymes in Ziziphus seedlings as compared with citrus seedlings. The treatment (A1W4) gave the enzymes activities CAT, highest SOD, BOD which was 24.10, 21.83 and 7.93 u / g / f.w respectively.

Minerals content: A negative relationship was appeared in N, P, k, Zn, Fe content in seedling leaves for both plants with increasing water salinity, While The interaction between concentrations the seaweeds and irrigation water types gave significant differences in minerals content. The highest value of minerals were at treatment A3W1 Table (5), which were 3.03%, 1.22%, 1.66%, 77.21µg/g, 60.39 µg/g for in N, P, k, Zn, Fe content respectively.

Discussion

high-quality plants can be produced only by using high-quality irrigation water. There are local differences in water characteristics with the source of the water, After analyzing the results from the experiment, the plants of this study responded significantly to the types of irrigation water, and indicated that the seedling growth was reduced at well water which has a high salinity comparing with tap water, this decrease due to the high ions concentration accumulation in the soil solution which may influenced on the enzymes activation inside the plant cells by modulating the active sites of enzymes also the H⁺-ATP ase pumps may stop working or the membranes may be impaired ,that affect the permeability as well as the salinity affected on the Photosynthesis, respiration processes and electron transport chains (Orcutt and Nilsen,2000). There is an imbalance of nutrients due to the factor involved in the salt-induced inhibition in photosynthesis and subsequently in plant growth reduction. We found that the type of water treatments play an important role in altering the mineral nutrient distribution and decreased the absorption rates, different researchers reported that the increasing salinity of water has effects on some traits of growth and yield (Hamaad et. al., 2013; Hirich et. al., 2014; Algosaibi et. al., 2015) and reduced Zn, Fe accumulation (Bhatt et. al., 2008). The antioxidant enzymes were raised positively with increasing of salt concentration in irrigation water and this activation may occur due to the excessive generation of reactive oxygen species (ROS) such as superoxide and hydroxyl radicals that are concede red as a one corollary of a biotic stress exactly as high salinity and cellular homeostasis (Mittler 2002). ROSs may have the ability to interact with many cellular components, and may result in significant damage to cell structures. When ROS increases. chain reaction start, in which superoxide dismutase (SOD) catalyzes the dismutation of O2 - radical to molecular O2 and H2O2 (Meloni et. al., 2003).

The H2O2 is then detoxified in the ascorbate-glutathione cycle (Mittler 2002), which involves the oxidation and re-reduction of ascorbate and glutathione through the ascorbate peroxidase (APX) and glutathion reductase (GR) enzymes (Noctor & Foyer 1998). If there is an imbalance in the cell compartment between the production of reactive oxygen species (ROS) and antioxidant defense, oxidative stress and damage occurs (Mittler 2002),many researchers reported that, (Maia *et. al.*, 2010; Kahrizi *et. al.*, 2012)

On the other hand, the interaction of irrigation water type and spraying with seaweeds gave the plants the ability to tolerate the salinity stress especially the plants which were irrigated with well water, compared with the untreated plants That maybe because the sea weeds could enrich the nutrients contents of the soil and is probably due to the presence of growth promoting hormones and nutrients in more quantities. The salinity of water stimulated the decreasing of leaf chlorophyll through embarrassment of chlorophyll synthesis or accelerated degradation has been well explained by Reddy and Vora (1986), and this reverberated on total carbohydrates content. Different researchers reported that the treatmentbyseaweed caused substantial increase in growth (Thirumaran et. al., 2009; Taha and Salih,2012; Abdul-Jabar et. al., 2012 ; Hamaad et. al., 2013;)An attention-grabbing observation that there is a relatively salt tolerance in Ziziphus seedlings more than Citrus seedling. It is well known that most crops are salt sensitive or hypersensitive (glycophytes) on the contrary to halophytes that have the capacity to accommodate severe salinity because of the very special anatomical and morphological adaptaavoidance mechanisms tions or (Flowers et al., 1986). These are relatively unique characteristics for which the genes are not likely to be introgressed easily into crop plant (Yokoi et al., 2002). From our study we can conclude that, the increasing of salinity of irrigation water over 5 dS.m⁻¹has a negative influence and reduction in growth parameters ,biochemical attributed and minerals contents. Also, the spraying with seaweeds act as a bio stimulator, reduce from the effects of salinity in irrigation water and encourage the plants to resist a bio stress like salinity, furthermore, type of plant play an important role in the tolerance toward salinity according to irrigation waters types the result shows that Ziziphus showed obvious tolerance compared with citrus seedling which is concedered a sensitive plant.

References

- A.O.A.C.(1984). Official Method of Analysis, 14 eds. Williams S, ed. Arlington, VI.
- Abdul-Jabar, A.S.; H. S. AL-Rashedy and M. A. AL-Ubeidi (2012). Effect of the Different Seaweed Extract (Seamino) Concentrationson Growth and Seed Chemical Composition of Two Wheat Varieties. Mesopotamia J. of Agric. Vol. 23 No. 1: 100 – 113.
- Aebi H (1984). Catalase *in Vitro*. Method Enzym 105: 121-126.
- Algosaibi, A. M., M.M. El-Garawany, A. E. Badran and A. M. Almadini (2015). Effect of Irrigation Water Salinity on the Growth of Quinoa Plant Seedlings. Journal of Agri-

cultural Science; Vol. 7, No.8.pp: 205-214.

- Beltran, J.M. (1999). Irrigation with saline water: Benefits and environmental impact. Agric. Water Manag., 40,183–194.
- Bhatt, M.J, Patel, A.D, Bhatti, P.M., Pandey A.N.(2008). Effect of soil salinity on growth, water status and nutrient accumulation inseedlings of *Ziziphus mauritiana* (Rhamnacea). – J. FruitOrnam. Plant Res. 16: 83-401.
- Boman, B.J. (1993). First-year response of 'Ruby Red' grapefruit on four root stocks to fertilization and salinity. Proceed. of the Florida– StateHort. Soc.., 106: 12–8.
- Bremner, J.M. and C.S. Mulvaney, (1982). Nitrogen-total. pp: 595-624 In A.L. Page *et al.* (ed.) Methods of soil analysis. Part 2. 2nd ed. Agron. Monograph 9. ASA and SSSA, Madison, WI.
- Dobois, M. K., K. A. Crilles, J. K. Hamiltor, D. A. Rebers and F. Smith (1956). Colorimetric method for determination of sugars and substance. Anal. Chem., 28: 350-365.
- Flowers, T.J, T.D. Colmer (2008). Salinity tolerance in halophytes. New Phytol,179:945-963.
- FlowersTJ, MA. Hajibagheri, NJW Clipson.(1986). Halophytes. The Quarterly Review of Biology 61, 313–337.
- Giannopotitis C. N. and S. K. Ries (1977). Superoxide dismutase in higher plants. Plant Physio 159: 309–314.
- Goodwin, T.W. (1976). Chemistry and Biochemistry of Plant Pigment. 2nd ed. Academic Press, London, New York., San Francisco, pp. 373.
- Hamaad,H.S., D.A. Mohammed and A. A. Obaid (2013).Effect of Irrigation water salinity, magnetic and soaking by the ascorbic acid and seaweeds (oligo-x) on germination

and seedling growth of hybrids Cucumber (Dalia)in protected environment. Diyala J. Agric. Sc., 5(2) 213-222.

- Hirich, A., A. Jelloul, R. Choukr-Allah and S.E. Jacobsen (2014). Saline water Irrigation of Quinoa and Chickpea: Seedling. Rate, Stomatal Conductance and Yield Responses J. Agro Crop Sci. 200 (378-389).
- Jackson, M. L.(1958). Soil chemical analysis. 1.ed. prentice-Hallinc, Englewood, Cliffs New Jersey.
- Kahrizi, S, M. Sedghi and O. Sofalian (2012). Effect of salt stress on proline and activity of antioxidant enzymes in ten durum wheat cultivarsAnnals of Biological Research, 2012, 3 (8):3870-3874.
- Kar P. K. and Choudhuri M. A. (1987). Possible mechanisms of light induced chlorophylleradication in senescencing leaves of hydrila veticillata. Physiol Plant 70:729–734.
- Maia O. M., E.L. Voigt, C. E. C. Macêdo, S. I. Ferreira-silva and J. A. G. Silveira (2010). Salt-induced changes in antioxidative enzyme activities in root tissues do not account for the differential salt tolerance of two cowpea cultivars.Braz. J. Plant Physiol., 22(1): 113-122.
- Maas, E.V., 1993. Salinity and citriculture. Tree Physiol., 12: 195–216.
- Meloni D. A., M. A. Oliva, C. A. Martinez and J. Cambraia (2003). Photosynthesis and activity of superoxide dismutase, peroxidase and glutathione reductase in cotton under saltstress. Environ Exp Bot 49:69-76.
- Mittler R., (2002). Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci.
- Murkute AA, Sharma S, Singh SK (2005). Citrus in terms of soil and water salinity: a review. Journal of

Scientific and Industrial Research 64:393 –402.

- Noctor, G. and C. H. Foyer(1998). Ascorbate and glutathione: Keeping active oxygen undercontrol. Annu Rev Plant Physiol Plant Mol Biol 49:249-279.
- Orcutt, D.M., and E.T. Nilsen (2000). The physiology of plants under stress. USA.
- Pitman MG. and A. Lauchli. (2002).
 Global impact of salinity and agricultural ecosystems. In: Salinity:
 Environment Plants Molecules,
 A. Lauchli and U. Lüttge (Eds.).
 Kluwer Academic Publishers,
 Dordrecht, pp. 3–20.
- Rathore S. (2009). Effect of seaweed extract on the growth, yield and nutrient uptake of soybean (Glycine max) under rainfed conditions, South African Journal of Botany,75;351-355.
- Reddy, M. P. and A. B. Vora (1986). Changes in pigment composition, hill reaction activity and saccharides metabolism in bajra (*Penni-setum typhoides* S & H) leaves under NaCl salinity; Photosynthetica; 20:50–55.
- Strick W.A.; M.S. Novak and J. Vanstaden. (2003). Cytokines in macro

algae. Plant Growth USA: Association of Official Analytical chemists. Regul.41:13-24.

- Taha, S. M. and L. M. Muhamad Salih (2012). Effect of Spry with Seaweed Extract (Matrix- 15) on Some Vegetative and Root Growth of two Strawberry Varieties (Fragaria X Ananasa Duch.) Journal of Kirkuk University for Agricultural Sciences. Vol. 3 (2):1-12.
- Thirumaran, G.; M. Arumugam; R. Arumugam and P. Anantharaman (2009). Effect of Seaweed Liquid Fertilizer on Growth and Pigment Concentration of *Cyamopsis tetragonoloba* (L)Taub. American-Eurasian Journal of Agronomy 2 (2): 50-56.
- Yokoi S., F.J. Quintero, B. Cubero, M.T. Ruiz, R.A. Bressan, P.M. Hasegawa, J.M. Pardo (2002). Differential expression and function of *Arabidopsis thaliana* NHX Na+/H+ antiporters in the salt stress response. Plant J., 30:529-539.
- Zodape,S.T(2001). Seaweedsasfertilizer. J. Sci. Ind.Res.60(5):378–382.

تأثير نوعية مياه الري والرش بمستخلص الأعشاب البحرية في النمو والصفات البايوكيميائية والمحتوى المعدني لشتلات السدر Ziziphus mauritiana والبرتقال المحلي Citrus sinensis

كريمة فاضل عباس ' ، خولة حمزة محمد ' وسلوى جمعة فاخر " لجامعة كربلاء ، كلية العلوم الطبية التطبيقية ، قسم الصحة البيئية . اجامعة البصرة ، كلية الزراعة ، قسم البستنة و هندسة الحدائق . حجامعة البصرة ، كلية الزراعة ، قسم التربة والموارد المائية .

الملخص

أجريت هذه الدر إسة لغرض معرفة تأثير نوعية مياه الري والرش بمستخلص الاعشاب البحرية في شتلات البر تقال Citrus sinensis والذي يعد من النباتات الحساسة للملوحة وشتلات السدر Ziziphus mauritiana من خلال مقاييس النمو الخضري والصفات البيوكيميائية والمحتوى المعدني ولقد طبقت اربع معاملات من مياه الري وهي ١- مياه الحنفية معامله $Ec = 5.2 \quad dS.m^{-1}$ ($Ec = 1.2 \quad dS.m^{-1}$ ($Ec = 1.2 \quad dS.m^{-1}$) ألمقارنة ($Ec = 1.2 \quad dS.m^{-1}$) مع ثلاث $Ec = 8.1 \text{ dS.m}^{-1}$ مياه البدر $Ec = 6.2 \text{ dS.m}^{-1}$ مع ثلاث - ۳ مستويات من مستخلص الأعشاب البحرية. (0, 2, 4 ml /L). وأظهرت النتائج حدوث انخفاض في جميع مقاييس النمو عند ارتفاع ملوحة مياه الري ولقد تمثلت هذه النتيجة في مياه البئر مقارنة متع باقي أنواع مياه الري والتي بلغت 42.94 سم في ارتفاع النبات و 00.2 في عدد الأفرع و 24.61 في عدد الأوراق و 5.34 في قطر الساق و 101.2 غم في الوزن الطري و 19.02 غم في الوزن الجاف وبينما اظهر التداخل مع مستخلص الأعشاب البحرية زيادة معنوية ملحوظة في مقاييس النمو والصفات البيوكيميائية والمحتوى المعدني، وأعطى أعلى زيادة عند المعاملة بـ 4) ml /L) وقد سجلت أعلى معنوية عند المعاملة A3W1 وبلغت 3.48 ملغم/غم في الكلوروفيل الكلي و73.82 ملغم/غم في الكربو هيدرات الكلية و بينما كانت الأدني عندA1W4 وبلغت 38. إملغم/ غم في الكلوروفيل الكلي و42.92 ملغم/غم في الكربو هيدرات الكلية. وقد أظهرت فعالية الأنزيمات(SOD) و superoxide dismutase و peroxidase (POD) و catalase (CAT) زيادة ملحوظة عند زيادة مستوى الملوحة في مياه الري وكذلك عمل المستخلص البحري على زيادة محتوى هذه الأنزيمات حيث أعطت المعاملة (A1W4) أعلى فعالية لأنزيم .CAT SOD, BOD وبلغت 24.10 و 21.83 و 7.93 وحدة/غرام/ وزن طرى وعلى التوالي.