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Laboratory diagnosis of novel species of *Theileria hirci*, *Eimeria caprovina* and *Eimeria pallida* in goats in Iraq

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Abstract

Laboratory examination of blood smears and fecal samples collected from 615 goats around Baghdad city revealed positive cases of *Theileria hirci* (208), *Eimeria caprovina* (94) and *Eimeria pallida* (48). These protozoa are novel species and have not been previously reported in Iraq. © 2002 Elsevier Science B.V. All rights reserved.

Keywords: Goats; *Theileria hirci*; *Eimeria caprovina*; *Eimeria pallida*; Iraq

1. Introduction

Theileriosis is one of the important diseases of farm animals in Iraq. In sheep, Little Wood (1915) reported the causative agent to be *Theileria hirci*. In cows, Machattie (1935) mentioned that the causative agent was *T. anulata*. In goats, theileriosis has only been suspected clinically by veterinarians without laboratory confirmation.

Mirza (1970) identified the following seven species of *Eimeria* in goats in Iraq: *E. crandalis* (*E. hirci*), *E. christenseni*, *E. parva* (*E. alijeri*), *E. fauri* (*E. apsheronica*), *E. ninakoholyakimovae*, *E. arlongi* and *E. granulosa* (*E. jolchijevi*).

The present investigation was carried out to identify some non-reported protozoa of goats in Iraq.

2. Materials and methods

Blood smears and fecal samples were collected randomly from 615 goats around Baghdad city (north, south, east, west) and from IPA station for goat breeding.

Blood smears were stained with Giemsa and examined under the oil-immersion lens (Adam et al., 1971). The parasitemia was calculated according to Schalm et al. (1975). Fecal sample of about 3–5 g were well mixed and filtered through four fold gauze, then treated with Sheather's sugar solution according to Dunn (1969).

3. Results

Microscopic examination of the 615 stained blood smear's for *T. hirci* revealed 208 (33.82%) positive cases, while examination of the 615 fecal samples

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Table 1
Number and percentage positive of *T. hirci*, *E. caprovina* and *E. pallida*

Protozoan	Number of positive animals	%
<i>T. hirci</i>	208	33.82
<i>E. caprovina</i>	94	15.28
<i>E. pallida</i>	48	7.8

revealed 94 (15.28%) and 48 (7.8%) positive cases for *E. caprovina* and *E. pallida* (Table 1).

The distribution of the positive cases for *T. hirci*, *E. caprovina* and *E. pallida* according to the directions

Table 2
Distribution of positive cases according to the directions and IPA station

Protozoan	North	South	East	West	IPA station	Total
<i>T. hirci</i>	17 (8.17%)	40 (19.23%)	66 (31.73%)	32 (15.38%)	53 (25.48%)	208
<i>E. caprovina</i>	10 (10.6%)	13 (13.8%)	33 (35.1%)	11 (11.7%)	27 (28.72%)	94
<i>E. pallida</i>	10 (20.83%)	14 (29.16%)	4 (8.3%)	9 (18.75%)	11 (22.91%)	48

Table 3
Description of *E. caprovina* and *E. pallida* oocysts in goats

Protozoan	Shape	Size (µm)	Micropyle	Micropolar cup	External layer of oocyst wall
<i>E. caprovina</i>	Broad oval or ellipsoidal	L: 27.72 (26.4-31.35), W: 22.4 (19.8-24.75)	Present	Not present	Colorless
<i>E. pallida</i>	Oval or ellipsoidal	L: 17.32 (16.5-18.5), W: 13.5 (13-14)	Present (±)	Not present	Pale yellow, green or colorless

and IPA station are shown in Table 2. *E. caprovina* and *E. pallida* descriptions of the oocysts are mentioned in Table 3. *T. hirci* was seen in many forms in blood smears. Fig. 1 shows *T. hirci* inside a red blood cell, while Figs. 2 and 3 show respectively the microschizonts and macroschizonts in the cytoplasm of a lymphocyte.

4. Discussion

T. hirci infection in general was 33.82% and according to the direction and IPA station, it ranged between 8.17 and 31.13%. These results are similar to Luo and

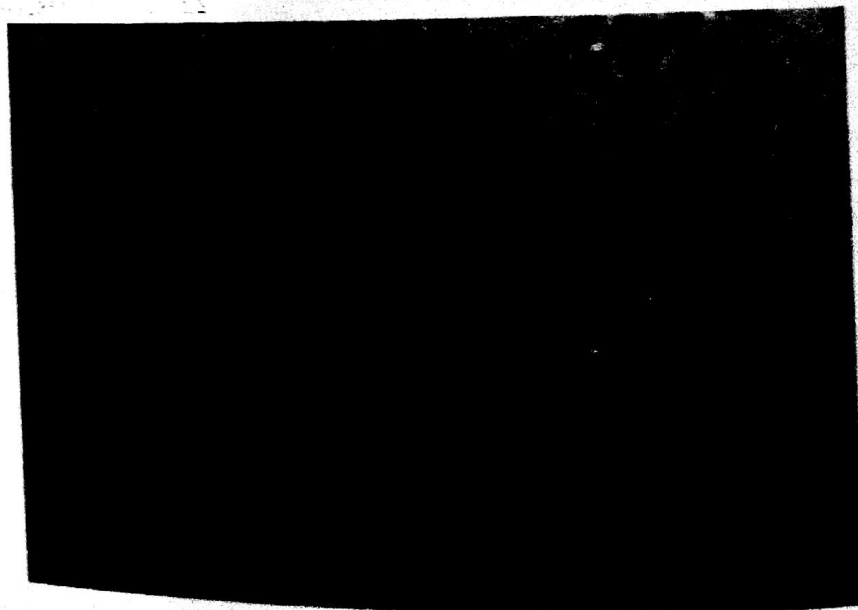


Fig. 1. *T. hirci* inside a red blood cell in a blood smear of a goat (1000×).



Fig. 2. Microschizonts of *T. hirci* inside a lymphocyte in a blood smear of a goat (1000×).

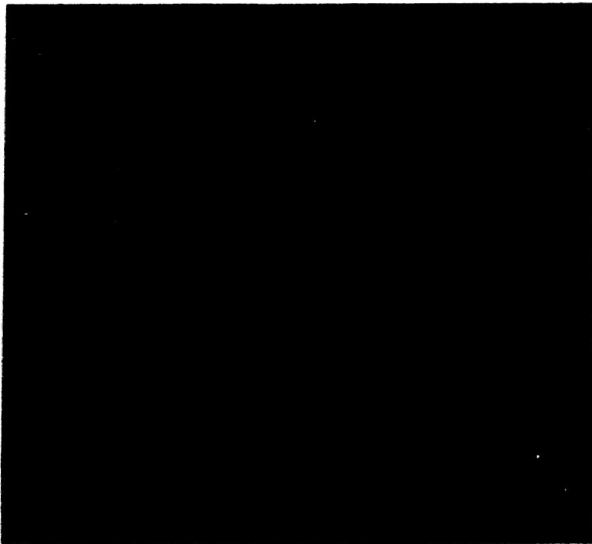


Fig. 3. Macroschizonts of *T. hirci* inside a lymphocyte in a blood smear of a goat (1000×).

Yin (1997) in China who found *T. hirci* infection in goats between 8.06 and 40%. Most positive cases of *T. hirci* of this investigation were apparently healthy. A single case showed enlargement of prescapular lymph nodes, pale conjunctiva, with (16%) parasitemia. Such finding is in accordance with Sasmal et al. (1983), who observed clinical signs of theileriosis in goats along with high parasitemias ranging between 18.25 and 31.36%. The shapes of *T. hirci* in examined blood smears were similar to Neitz (1957).

The 15–28% of *E. caprovina* infection of this investigation lies within the ranges mentioned by Norton (1986), O'callaghan (1989), Penzhorn et al. (1994), Kusiluka et al. (1996) Koudela and Bokova (1998) and Jalila et al. (1998) (4, 12, 29.4, 16, 6 and 12%, respectively). The description of the oocyst was similar to Lima (1980).

E. pallida infection of 7.8% is within the percentage ranges mentioned by Chhabra and Pandey (1991) and Jalila et al. (1998) (6.2 and 4%, respectively), while Woji et al. (1994) reported an infection percentage of 22%. The description of the oocyst was similar to Christensen (1938).

5. Conclusion

Due to the natural resistance of goats to infection by *T. hirci*, high parasitemias are required for obvious clinical signs.

References

- Adam, K.M.G., Paul, J., Zaman, V., 1971. Medical and Veterinary Protozoology. Churchill Livingstone, London, 200 pp.
- Chhabra, R.C., Pandey, V.S., 1991. Coccidia of goats in Zimbabwe. Vet. Parasitol. 39, 119–205.
- Christensen, J.F., 1938. Species differentiation in the coccidia from the domestic sheep. J. Parasitol. 24, 453–465.

- Dunn, A.M., 1969. Veterinary Helminthology. Maclehorse, R. and Co. Ltd., University Press, Glasgow, UK, pp. 275-283.
- Jalila, A.M., Dorny, P., Sani, R., Salim, N.B., Vercrugsse, J., 1998. Coccidial infections of goats in Selangor, Peninsular Malaysia. *Vet. Parasitol.* 74, 165-172.
- Koudela, B., Bokova, A., 1998. Coccidiosis in goats in the Czech Republic. *Vet. Parasitol.* 76, 261-267.
- Kusiluka, I.J.M., Kambarage, D.M., Matthewman, R.W., Harrison, L.J.S., Daborn, C.J., 1996. Coccidiosis of small ruminants in Tanzania. *Small Rumin. Res.* 21, 127-131.
- Lima, J.D., 1980. *Eimeria caprovina* spp. from domestic goat *Capra hircus*, from the US. *J. Protozool.* 27, 153-154.
- Little Wood, W., 1915. Annual Report for the Year 1914. Veterinary Services, Ministry of Agriculture, Government Press, Cairo. Cited from Neitz, W.O. (1957). Theileriosis, gonderiosis and cytoxozoonoses: a review. *Onderstepoort J. Vet. Res.* 27, 275-430
- Luo, J., Yin H., 1997. Theileriosis of sheep and goats in China. *Trop. Anim. Health Prod.* 29 88-105.
- Machattie, C., 1935. Theileriosis of young calves in Baghdad dairies. *Trans. R. Soc. Trop. Med. Hyg.* 28, 649-654.
- Mirza, Y.M., 1970. Incidence and distribution of coccidia (Sporozoa Eimeriidae) in mammals from Baghdad area. M.Sc. Thesis. College of Science, University of Baghdad, 195 pp.
- Neitz, W.O., 1957. Theileriosis, gonderiosis and cytoxozoonoses: a review. *Onderstepoort J. Vet. Res.* 27, 275-430.
- Norton, C.C., 1986. Coccidia of the domestic goat *Capra hircus* with notes on *Eimeria ovinoidallis*, *E. bakuensis* (Syn. *ovina*) from the sheep *Ovis aries*. *Parasitology* 92, 279-287.
- O'callaghan, M.G., 1989. Coccidia of domestic and feral goats in south Australia. *Vet. Parasitol.* 30, 267-272.
- Penzhorn, B., Rognlie, M.C., Hall, L.L., Knapp, S.E., 1994. Eimeria coccidia of Cashmere goats in south western Montana. *Vet. Parasitol.* 55, 137-142.
- Sasmal, N.K., Biswas, S.S., Brattacharyya, B., Banerjee, C., Maitra, D.N., 1983. *Theileria hirci*—studies on transmission from Sahabadi sheep to black Bengal goat. *Ind. Vet. J.* 60, 599-604.
- Schalm, W.O., Jain, N.C., Carroll, E.J., 1975. *Veterinary Hematology*, 3rd Edition. Lea and Febiger, Philadelphia, PA, 807 pp.
- Woji, A.Y., Little, D.A., Ikwuegbu, O.A., 1994. Prevalence of coccidial infections in the west Africa dwarf goat in the subhumid zone of Nigeria. *Trop. Anim. Health Prod.* 26, 1-10.