

Records of two gonad-infecting species of *Philometra* (Nematoda: Philometridae) from marine fishes off Iraq, including the description of *Philometra parabrevicollis* n. sp. from the bigeye snapper *Lutjanus lutjanus* Bloch (Pisces, Lutjanidae)

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Abstract Recent examinations of some marine fishes from off the southern coast of Iraq revealed the presence of two species of Philometra Costa, 1845 (Nematoda: Philometridae): P. parabrevicollis n. sp. (males and subgravid and nongravid females) from the ovary of the bigeye snapper Lutjanus lutjanus Bloch (Perciformes, Lutjanidae) and Philometra sp. (subgravid females) from the ovary of the bartail flathead Platycephalus indicus (Linnaeus) (Perciformes, Platycephalidae). Specimens of species are described and illustrated based on light and scanning electron microscopical examinations. Philometra parabrevi*collis* **n. sp.** is mainly characterised by the length of spicules (267-285 µm) and gubernaculum (159-168 μm), the gubernaculum/spicule length ratio (1:1.64–1.76), the structure of the gubernaculum distal

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J. A. Mizher · A. H. Ali Department of Fisheries and Marine Resources, College of Agriculture, University of Basrah, Basrah, Iraq tip and of the male caudal end, and the body length of males (4.03–4.90 mm). The description of this new species again confirms a high degree of host specificity of gonad-infecting species of *Philometra* in congeneric lutjanid hosts. Although *Philometra* sp. parasitising *P. indicus* in Iraqi waters was previously recorded, its subgravid females are described for the first time. A key to gonad-infecting species of *Philometra* parasitic in fishes of the family Lutjanidae is provided.

Introduction

The dracunculoid family Philometridae Baylis & Daubney, 1926 contains many species of nematode parasites of fishes. Of the 17 presently recognised philometrid genera (Moravec & Shamsi, 2017; Moravec et al., 2019), the great majority of described species belongs to Philometra Costa, 1845, that includes parasites of the body cavity and various body tissues of freshwater, brackish-water and marine fishes (Ivashkin et al., 1971; Moravec, 2006; Anderson et al., 2009; Moravec & de Buron, 2013). To date, 13 philometrid species of the genera Clavinemoides Moravec, Khosheghbal & Pazooki, 2013, Philometra and Philometroides Yamaguti, 1935 were recorded from marine fishes in the Arabian (= Persian) Gulf (Parukhin, 1976; Moravec & Ali, 2005, 2013, 2014; Moravec et al., 2012, 2013, 2016a; Khosheghbal et al., 2017); reports of two additional species, *Philometra* globiceps (Rudolphi, 1819) and P. lateolabracis (Yamaguti, 1935), in this region by Petter & Sey (1997) and Kardousha (1999), respectively, were based on misidentifications (Moravec & Ali, 2013). Also the finding of Philometra madai Quiazon, Yoshinaga & Ogawa, 2008, a specific parasite of the Sparidae in Japan (see Quiazon et al., 2008; Moravec et al., 2019), in Lutjanus monostigma (Cuvier) (Lutjanidae) in the Red Sea, off Saudi Arabia (Al-Jahdali, 2014) is evidently based on misidentification, because only unidentifiable nematode females were collected; these should be reported as *Philometra* sp. until conspecific males are obtained. Records of unidentified species of Philometra from different fish hosts reported by Parukhin (1976), El-Naffar et al. (1992), Mohamed et al. (2010), Hosseini et al. (2013), Ali et al. (2014) and Moravec et al. (2016a) indicate the presence of additional, not yet established philometrid species in marine fishes of the Arabian Gulf.

During recent helminthological investigations of some marine fishes in the Arabian Gulf, off the coast near Basrah, southern Iraq, specimens of *Philometra* were found in the ovaries of the bigeye snapper *Lutjanus* lutjanus Bloch (Perciformes, Lutjanidae) and the bartail flathead *Platycephalus indicus* (Linnaeus) (Perciformes, Platycephalidae). Results of their closer examination are described below.

Lutjanus lutjanus (maximum body length 35 cm) is a tropical marine, highly commercial fish, whereas *P. indicus* (maximum body length 100 cm) is a subtropical marine commercial fish and gamefish; both of them have an Indo-West Pacific distribution (Froese & Pauly, 2021).

Materials and methods

Fishes were caught using trawl and drift gill nets during October–December 2019 and January–May and August–September 2020. The nematodes obtained from fresh fishes were washed in physiological saline and fixed in hot 5% formalin. For light microscopy examination, the nematodes were cleared with glycerine. Drawings were made with the aid of a Zeiss drawing attachment. Specimens used for scanning electron microscopy (SEM) were postfixed in 1% osmium tetroxide (in phosphate buffer), dehydrated through a graded acetone series, critical-point-dried and sputter-coated with gold; they were examined using a JEOL JSM-7401F scanning electron microscope at an accelerating voltage of 4 kV (GB low mode). All measurements are in micrometres unless otherwise indicated. The fish nomenclature adopted follows Eschmeyer's Catalog of Fishes (Van der Laan et al., 2021). Ecological terms (prevalence and intensity of infection) are used in accordance with Bush et al. (1997).

Philometridae Baylis & Daubney, 1926 Philometra Costa, 1845

Type species: *Philometra reticulatum* Costa, 1845, by original designation [= syn. of *Philometra globiceps* (Rudolphi, 1816) Railliet, 1916]

Philometra parabrevicollis n. sp

Type-host: Lutjanus lutjanus Bloch (Perciformes: Lutjanidae), bigeye snapper.

Type-locality: Near Basrah, northwestern Arabian Gulf, Iraq (collected in May 2020).

Type-material: Male holotype, female allotype and 17 paratypes (4 males, 13 females) in the Helminthological Collection of the Institute of Parasitology, BC CAS, České Budějovice, Czech Republic (Cat. No. N– 1255).

Prevalence and intensity: 5% (3 fish infected/62 fish examined); 4–9 (mean 7) nematode specimens per fish.

Site in host: Ovary.

ZooBank registration: To comply with the regulations set out in article 8.5 of the amended 2012 version of the *International Code of Zoological Nomenclature* (ICZN, 2012), details of the new species have been submitted to ZooBank. The Life Science Identifier (LSID) for *Philometra parabrevicollis* **n. sp.** is urn:lsid:zoobank.org:act:9BB75C20-F52F-44AB-ADEE-62E2CA9389FF.

Etymology: The species name *parabrevicollis* is a Latin adjective composed of two parts, *par* (= match, alike, comparable with) and *brevicollis* (= the name of nematode species), and relates to the fact that this nematode is morphologically very similar to *Philometra brevicollis*.



Fig. 1 *Philometra parabrevicollis* **n. sp.** from *Lutjanus lutjanus*. A, B, Anterior end of male and nongravid female, respectively, lateral views; C, Anterior end of subgravid female, lateral view; D, E, Cephalic end of male and subgravid female, respectively, apical views; F, Posterior end of subgravid female, dorsoventral view; G, H, Posterior end of male, ventral and lateral views, respectively; I, J, Posterior end and vulva of nongravid female, lateral views, respectively; K, Gubernaculum, lateral view; L, M, Distal end of gubernaculum, dorsal and lateral views, respectively; N, Caudal end of male, apical view

Description

Male [Based on 8 specimens; measurements of holotype in parentheses; Figs. 1A, D, G, H, K–N, 2C–G, 3A–G] Body filiform, whitish, 4.03–4.90 (4.90) mm long, maximum width 60–84 (66); anterior end of body rounded, 45–54 (48) wide, with conspicuous

constriction at short distance posterior to cephalic extremity (Figs. 1A, 2E); width of body at this constriction 33–39 (36). Maximum width/body length ratio 1:53–74 (1:74). Cuticle smooth. Oral aperture small, circular. Cephalic papillae 14 in number, arranged in 2 circles; outer circle formed by 4 submedian pairs, inner circle consists of 4 submedian



Fig. 2 *Philometra parabrevicollis* **n. sp.** from *Lutjanus lutjanus*, scanning electron micrographs. A, B, Cephalic end of subgravid female, subventral and apical views, respectively; C, D, Cephalic end of male, lateral and apical views, respectively (arrow indicates amphid); E, Anterior body end of male, lateral view; F, G, Distal end of gubernaculum, lateral and subdorsal views, respectively. *Abbreviations*: a, submedian pair of outer cephalic papillae; b, inner submedian cephalic papilla; c, inner lateral cephalic papilla; g, gubernaculum; o, oral aperture; s, spicule

and 2 lateral papillae (Figs. 1D, 2C, D). Pair of small, slit-like lateral amphids somewhat posterior to lateral cephalic papillae (Figs. 1D, 2D). Oesophagus 462–594 (525) long, comprising 10–12% (11%) of body length, with slight inflation at anterior end measuring $36-51 \times 24-33$ (39×30); posterior part of muscular oesophagus overlapped by well-developed oesophageal gland with large cell nucleus; maximum

width of gland 24–30 (27). Ventriculus indistinct. Nerve-ring and oesophageal nucleus 183–228 and 309–426, respectively, from anterior extremity. Excretory pore 204–225 (255) from anterior end. Testis extending anteriorly approximately to level just posterior to nerve ring (Fig. 1A). Posterior end of body blunt, 36–45 (42) wide, provided with broad U-shaped mound situated laterally and dorsally to cloacal



Fig. 3 *Philometra parabrevicollis* **n. sp.** from *Lutjanus lutjanus*, scanning electron micrographs of male. A, B, Caudal end, sublateral and subventral views, respectively; C, Region of cloaca, ventrolateral view; D, Caudal end, apical view (arrow indicates phasmid); E, F, Posterior end of body, ventral and lateral views, respectively; G, Region of cloaca, subventral view. *Abbreviations*: d, caudal mound; e, postcloacal papilla-like protuberance; f, group of four adanal papillae; g, gubernaculum; s, spicule

opening (Figs. 1N, 3A, B, D–F). Four pairs of small, very flat, hardly visible adanal papillae present on caudal mound; 2 large papilla-like protuberances present posterior to cloacal aperture; pair of small sublateral phasmids present on caudal mound at level of postcloacal protuberances (Figs. 1N, 3A, B, D). Spicules needle-like, approximately equally long, with somewhat expanded proximal and sharply

pointed distal tips (Figs. 1G, H, 2F, G, 3A–G); length of spicules 267–285 (285), representing 5.5–6.7% (5.8%) of body length. Gubernaculum narrow, 159–168 (162) long, with anterior portion somewhat dorsally bent (Fig. 1H, K); length of anterior bent part 33–48 (48), comprising 20–30% (30%) of entire gubernaculum length; distal portion of gubernaculum tongue-shaped in dorsal view, rounded at tip, each dorsolateral margin with rounded transversely striated ridge; these ridges separated along their entire length by smooth depressed distally tapering field (Figs. 1L, M, 2F, G, 3A, C, F); ventral surface of this region with 2 longitudinal grooves (Fig. 3C, D, G). Length ratio of gubernaculum and spicules 1:1.64–1.76 (1:1.76). Spicules and gubernaculum well sclerotised, yellowish, anterior part of gubernaculum and proximal ends of spicules colourless.

Subgravid female [Based on 4 ovigerous specimens; measurements of allotype in parentheses; Figs. 1C, F, E, 2A, B] Body whitish, 17.03-27.88 (27.45) mm long; maximum width/body length ratio 1:78-124 (1:96). Cephalic end rounded, 87-123 (105) wide. Cephalic papillae very small, indistinct in lateral view (Fig. 1C). Oral aperture oval, surrounded by 14 small cephalic papillae arranged in 2 circles: 4 submedian pairs of papillae of outer circle and 6 single papillae (4 submedian and 2 lateral) of inner circle; amphids indistinct (Figs. 1E, 2A, B). Oesophagus 789-884 (870) long including its anterior bulbous inflation, representing 3-5% (3%) of body length; anterior inflation 75-90 (75) long and 81-90 (90) wide; maximum width of posterior part of oesophagus including well-developed dorsal gland 51-81 (81) (Fig. 1C). Small ventriculus present, 21 (21) long, 60-66 (66) wide. Nerve-ring and oesophageal nucleus 218-245 (245) and 503-598 (598), respectively, from anterior extremity. Intestine straight, ending blindly; posterior end of intestine atrophied, forming ligament 180-270 (180) long attached ventrally to body wall close to posterior extremity (Fig. 1F). Vulva and anus absent. Ovaries reflexed near body ends. Uterus filled with eggs. Posterior end rounded, 69-150 (95) wide, with pair of minute papilla-like caudal projections (Fig. 1F).

Nongravid female [Based on 7 specimens; Fig. 1B, I, J] Body whitish, 5.56–9.27 mm long, maximum width 60–177; maximum width/body length ratio 1:52–93. Cephalic end rounded, 39–69 wide. Cephalic papillae small, indistinct in lateral view (Fig. 1B). Oesophagus including anterior bulbous inflation 540–857 long, comprising 9–12% of body length; its anterior inflation 39–69 long and 27–66 wide. Posterior portion of oesophagus including oesophageal gland 30–54 wide (Fig. 1B). Nerve-ring and oesophageal nucleus 150–180 and 375–544, respectively, from anterior extremity. Ventriculus 6–24 long, 15–45 wide. Rudimentary vulva at 3.62–3.92 mm from anterior extremity (at 65–67% of entire body end) in small specimens 5.56–5.83 mm long (Fig. 1J), but vulva absent in larger specimen 9.26 mm long. Uterus empty. Posterior end rounded, 36–163 wide, without caudal projections (Fig. 1I).

Remarks

Gonad-infecting species of *Philometra* are known to exhibit a high degree of host specificity (e.g., Moravec & Manoharan, 2014a, b; Moravec et al., 2016b; Ghanmi et al., 2018; Moravec & Barton, 2018). To date, the following ten species of *Philometra* parasitic in the gonads of Lutjanus spp. with known males have been described, all in the regions of the Atlantic Ocean (Gulf of Mexico), Indian Ocean (Bay of Bengal) and South Pacific Ocean (off Australia and New Caledonia) (Moravec & Barton, 2018): P. arafurensis Moravec & Barton, 2018, P. argentimaculati Moravec & Manoharan, 2014, P. brevicollis Moravec & Justine, 2011, P. carponotati Moravec & Diggles, 2014, P. fulvi Moravec & Manoharan, 2014, P. latispicula Moravec. Bakenhaster & Fajer-Ávila, 2014, P. longispicula Moravec, Bakenhaster & Fajer-Avila, 2014, P. mawsonae Moravec & Barton, 2018, P. mira Moravec & Justine, 2011 and P. synagridis Moravec, Bakenhaster & Fajer-Ávila, 2014 (see Moravec & Justine, 2011; Moravec & Diggles, 2014; Moravec & Manoharan, 2014b; Moravec et al., 2014; Moravec & Barton, 2018).

Of these, the spicules of six species are either distinctly shorter (P. fulvi, P. latispicula, P. mira and P. synagridis) or distinctly longer (P. longispicula and *P. mawsonae*) than those in the new species. By the length of spicules, P. parabrevicollis n. sp. somewhat resembles P. arafurensis, P. argentimaculati, P. brevicollis and P. carponotati. Males of all these four species, as well as that of the new species, are characterised by a distinct "neck" on their anterior body ends, by the presence of a U-shaped caudal mound and an elongate tongue-shaped distal portion of the gubernaculum with numerous dorsolateral transverse lamella-like structures with an elongate smooth dorsal field between them; a pair of small caudal papilla-like projections observed in the subgravid females were also found in gravid or subgravid females of P. arafurensis, P. brevicollis and P. carponotati.



Fig. 4 *Philometra* sp. from *Platycephalus indicus*, subgravid female. A, Anterior end, lateral view; B, Posterior end, lateral view; C, Cephalic end, apical view

Of the four above-mentioned species with spicule lengths somewhat similar to those of the new species, only the spicules of *P. brevicollis* are approximately as long as those of P. parabrevicollis n. sp. (279-321 µm vs 267-285 µm), whereas those of P. arafurensis, P. argentimaculati and P. carponotati are somewhat shorter, not exceeding 264 µm. Also the gubernaculum of P. brevicollis is approximately as long as that of the new species (151–168 μ m vs 159–168 μ m), whereas the gubernaculum of P. arafurensis, P. argentimaculati and P. carponotati is shorter, not exceeding 147 um. However, P. brevicollis differs from P. parabrevicollis n. sp. in the gubernaculum/spicules length ratio (1:1.79–1.93 vs 1.64–1.76), relative length of spicules to the entire body length (5.0-5.3% vs)5.5–6.7%), body length (5.39–5.92 mm vs 4.03–4.90 mm) and in the absence (vs presence) of a pair of large postcloacal papilla-like protuberances. Philometra brevicollis also differs from the new species in the number (three vs four pairs) of observed adanal papillae, but this is an unrealible feature, because these papillae are usually difficult to observe in philometrids.

Distinction of *P. parabrevicollis* **n. sp.** from other gonad-infecting species of *Philometra* parasitising lutjanids is apparent from the key at the end of Discussion.

Philometra sp. 2 of Moravec et al., 2016

Host: Platycephalus indicus (Linnaeus) (Perciformes, Platycephalidae), bartail flathead.

Locality: Near Basrah, northwestern Arabian Gulf, Iraq (collected in February–March 2020).

Voucher specimens: Helminthological Collection of the Institute of Parasitology, BC CAS, České Budějovice, Czech Republic (Cat. No. N–1100).

Prevalence and intensity: 3% (3fish infected/118 fish examined; 1–4 (mean 2) nematodes per fish. *Site in host*: Ovary.

Description

Subgravid female ([Based on 2 complete and 3 incomplete specimens; Figs. 4A-C, 5A): Body of fixed specimens grey-coloured, 96-195 mm long, maximum width 938-1,251; maximum width/body length ratio 1:102-156. Cephalic end rounded, 258-340 wide. Cephalic papillae very small, indistinct in lateral view (Fig. 4A). Oral aperture oval, surrounded by 14 small cephalic papillae arranged in 2 circles: 4 submedian pairs of papillae of outer circle and 6 single papillae (4 submedian and 2 lateral) of inner circle; amphids indistinct (Figs. 4C, 5A). Oesophagus 1.29-1.56 mm long including its anterior bulbous inflation, representing 0.8-1.3% of body length; anterior inflation 150-177 long and 177 wide; maximum width of posterior part of oesophagus including well-developed dorsal gland 136-163 (Fig. 4A). Small ventriculus present, 27–68 long, 95



Fig. 5 *Philometra* sp. from *Platycephalus indicus*, scanning electron micrographs. A, Cephalic end of subgravid female from Iraq, subapical view; B, Cephalic end of gravid female from India, apical view. *Abbreviations*: a, submedian pair of outer cephalic papillae; b, inner submedian cephalic papilla; c, inner lateral cephalic papilla

wide. Nerve-ring and oesophageal nucleus 231–326 and 830–979, respectively, from anterior extremity. Intestine straight, ending blindly; posterior end of intestine atrophied, forming ligament 462–625 long attached ventrally to body wall close to posterior extremity (Fig. 4B). Vulva and anus absent. Ovaries coiled near body ends (Fig. 4A, B). Uterus filled with eggs. Posterior end rounded, 150–190 wide, without caudal projections (Fig. 4B).

Remarks

Two small nongravid and one incomplete (posterior part of body) gravid females from the ovary of *P. indicus* in Iraq were previously described as *Philometra* sp. 2 by Moravec et al. (2016a). Larger subgravid females of this not yet established species are described in this paper for the first time.

According to Moravec et al. (Moravec et al. 2016a, these nematodes are probably conspecific with those described by Moravec & Nagasawa (1989) from the gonads of P. indicus off Japan and misidentified as Philometra inimici Yamaguti, 1941. In 2013, the senior author (F.M.) of the present paper examined one gravid female of Philometra (Fig. 5B), 92 mm long, from the ovary of P. indicus of the Bay of Bengal, India, sent to him by Dr. Jayaraman Manoharan; this finding has not been published. It is highly probable that all these nematodes from the gonads of P. indicus collected in Iraq, India and Japan belong to the same, hitherto undescribed species of *Philometra*, whose distribution area coincides more or less with the Indian-West Pacific distribution of its host *P. indicus*. Since the female morphology of available specimens is similar to that of most other gonad-infecting congeners, the new species cannot be established until conspecicific males are collected and described.

Discussion

The present study shows again the importance of male morphology for the taxonomy of philometrids, because females of many species of Philometra are very similar and practically indistinguishable from each other by morphological features. As mentioned by Moravec et al. (2021), besides the shape and lengths of spicules, the most important taxonomic features of these parasites seem to be a detailed structure of the male caudal end and, in particular, the structure of the gubernaculum distal tip. During recent years, especially the latter feature made it possible to distinguish between some gonad-infecting species of Philometra with morphologically indistinguishable females (e.g., Moravec et al., 2016b; Ghanmi et al., 2018; Moravec & Barton, 2018). However, it is necessary to note that the details of the gubernaculum have to be studied by SEM.

Snappers (*Lutjanus* spp.) seem to be frequently parasitised by philometrid nematodes (Moravec et al.,

2014). Besides Dentiphilometra lutjani González-Solís, Moravec & Tuz Paredes, 2007 (a parasite of body musculature) and Philometra brevispicula Moravec a Bakenhaster, 2010 (a parasite of mouth tissues), both parasites of Lutjanus griseus (Linnaeus) in Florida Bay, USA and off the Caribbean coast of Mexico, respectively (González-Solís et al., 2007; Moravec & Bakenhaster, 2010) and D. malabarici Moravec & Barton, 2018 and P. papillicaudata Moravec & Barton, 2018 (both parasites of tissues behind gills) of Lutjanus malabaricus (Bloch et Schneider) and L. sebae (Cuvier), respectively, from off northern Australia (Moravec & Barton, 2018), all other remaining species are represented by the parasites of fish gonads. The males of all these 11 nominal gonad-infecting species of Philometra (see above) are known and only those of P. mira have not yet been studied by SEM.

The present description of an additional new gonadinfecting species of *Philometra* from *L. lutjanus* off Iraq confirms the high degree of host specificity of these nematodes previously observed in *Lutjanus* spp. in the Gulf of Mexico (Moravec et al., 2014), in the Bay of Bengal (Moravec & Manoharan, 2014b) and in northern Australian waters (Moravec & Barton, 2018).

Key to gonad-infecting species of *Philometra* parasitising fishes of the family Lutjanidae:

1a Spicules longer than 350 µm 2 2a Spicules 459-690 µm and 378-618 µm long; Longer spicule represents 13–19% of body length. Length ratio of gubernaculum and longer spicule 1:4.5-6.9. Gravid female 130-250 mm long. In Lutjanus campechanus and L. vivanus; Gulf of Mexico P. longispicula 2b Spicules 384-435 µm and 351-420 µm long; longer spicule represents 10-11% of body length. Length ratio of gubernaculum and longer spicule 1:2.1-2.4. Gravid female 70-111 mm long. In Lutjanus malabaricus; off AustraliaP. mawsonae 3a Spicules shorter than 150 µm 4 3b Spicules longer than 150 µm 6 4a Spicules 96–105 µm long. Length of gubernaculum 78–84 µm. Body length of male 3.47–3.66 mm. Female unknown. In Lutjanus vitta; off New Caledonia P. mira

4b Spicules longer than 120 µm5

5a Gubernaculum 69–93 µm long, its distal end with dorsal protuberance (in lateral view) and with numerous dorsal lamela-like structures arranged in 2 longitudinal rows separated by median groove (3 posteriormost lamellae undivided). Spicules 123–138 µm long. Body length of gravid female 230 mm. In *Lutjanus fulvus* (Lutjanidae); Indian Ocean (Bay of Bengal) *P. fulvi*

6a Body length of male 5.39-5.92 mm. Spicules 279–312 µm long. Length of gubernaculum 151–168 µm; length ratio of gubernaculum and spicules 1:1.79–1.93; distal end of gubernaculum with dorsal surface provided with 2 lateral rows of lamela-like structures and smooth longitudinal median field between them. In *Lutjanus vitta* (Lutjanidae); off New Caledonia*P. brevicollis*

6b Body length of male less than 5 mm7

7a Spicules 267–285 μ m long, length of gubernaculum 159–168 μ m; length ratio of gubernaculum and spicules 1:1.64–1.76; distal end of gubernaculum with dorsal surface provided with 2 lateral rows of lamelalike structures and smooth longitudinal median field between them. Gravid female not known. In *Lutjanus lutjanus*; off Iraq *P. parabrevicollis* **n. sp.**

9b Spicules usually shorter..... 10

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval All applicable institutional, national and international guidelines for the care and use of animals were followed.

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