

Histopathological and Molecular Study of *Raillietina echinobothrida* of Domestic and Wild Pigeons in Al-Muthanna Province

Hussein J. Jasim¹, Ibrahim A. Abdulzahra², Katherine B. Faraj³, Saif M. Abed⁴

¹Department of Vet. Parasitology, College of Vet. Med., ²Department of Biology, College of Science, University of Al-Muthanna, Iraq; ³Department of Microbiology and Vet. Parasitology, College of Vet. Med., University of Basrah, Iraq; ⁴Department of Microbiology science, Samawah, Iraq.

ABSTRACT

This study was conducted to detect and distinguish *Raillietina echinobothrida* of domestic and wild infecting pigeons has been based on morphology, histopathological and molecular analysis. The present study was conducted on 115 pigeons (73 of *S. Decaecto* and 42 of *C. livia*) of different age groups and of both sexes. The results of the present study were revealed that only 86 out of 115 examined pigeons were infected by *Raillietina* spp at infection rate of 74.78%. Besides, the percentage of infection with *Raillietina* spp were recorded in *S. decaecto* is higher than *C. livia* in 44.34 % and 30.43% respectively. The most isolated parasites of examined birds were showed the cestoda *R. echinobothrida* with total number (168) adult worms from total number of examined pigeons infected (115). The histopathological examination of small intestine of pigeons infected with *R. echinobothrida* were showed different changes like, an infiltration of inflammatory cells in lamina propria and atrophy of villi of small intestine with vacuolation of muscularis externa and loss of mucous membrane. The result of PCR were showed that specific primers (3S-F, BD2-R) were successfully for amplifying 944 bp with flanking region of ITS2 to *R. echinobothrida*. Also, the PCR result were revealed that 61 (70.93%) out of 86 birds were positive to *R. echinobothrida*.

Keywords: *R. echinobothrida*, domestic and wild pigeons, Pathological, Molecular, Iraq

Introduction

Iraq has a wide range of geological diversity from the peaks of Kurdistan Mountains in the north to the deserts and semi-deserts in the west and the great marshes in the south, these contribute to the wide biological as well as faunal diversity¹.

In recent years, poultry farming has tremendously developed and become one of the most intensive forms of animal husbandry activities². The Domestic pigeon (*Columbia livia*) is one of the commonest birds and tend to be closely contact with humans and birds as a source of protein, hobby, and recently as laboratory animals tend to

be closely contact with humans and birds as a source of protein, hobby, and recently as laboratory animals^{3,4}.

Parasitic infections cause great economic losses to poultry in our country, the birds are under constant stress and are prone to parasitic infections⁵. Endoparasite of poultry are common in the tropical and sub-tropical where the standard of husbandry is poor and climatic conditions are favorable for their existence^{6,7}.

Raillietina echinobothrida is consider the most pathogenic and prevalent species infecting birds throughout the world^{8,9}. This parasite inhabits in the jejunum and ileum of the definitive host, from where it obtains nutrition from the digested food of the host, whereas their larval stage (cyticeroid) resides in various invertebrate intermediate host, such as ants, beetles, small mini-wasps, or termites for completion of its life cycle. *R. echinobothrida* is a hermaphrodite worm having both the male and female reproductive organs in its body^{10,11}.

Corresponding Author:

Hussein J. Jasim
Department of Vet. Parasitology,
College of Vet. Med., University of Al-muthanna, Iraq
Email: husseinjasem2014@gmail.com

R. echinobothrida is responsible for 'nodular tapeworm disease' in poultry and retarded growth of young birds, emaciation of adult and low egg production of hen, disrupt gastrointestinal tract digestion decrease in consumption of fodder and intestinal obstruction can occur in heavy infection¹². In addition, this tapeworm does not cause gross pathological damages on well-nourished bird in the light infection, but do compete for food when they large numbers of parasites. If the heavy infection becomes, marked severe lesions on the intestinal walls due to the scolex are penetrate the mucosal layer and causes characteristic hyperplastic enteritis associated with the formation of granuloma (nodules contain cheesy and calassified material) on the intestinal wall of the infected birds and diarrhea could arise, which ostensibly resulted in ill health^{11,13}.

The taxonomy of Cyclophyllidea traditionally depends upon the accurate descriptions of small differences in the size and shape of the scolex, rostellum (unarmed or armed with rows of hooks) suckers and gravid proglottids⁹. Nevertheless, with a rich resource of bourgeoning data the molecular approach is proving to be more rapid, sensible and reliable than that based on morphological data alone for phylogenetic analysis¹⁴. The objective of the present study was to study and recognize the histopathological changes result of the invasive R. echinobothrida in the small intestine, supplementing the morphological observation, also confirmed the diagnosis of R. echinobothrida in the birds by the molecular diagnostic PCR technique.

Materials and Method

Study Area and Sampling: A total number of 115 pigeon samples (73 of *Streptopelia Decaecto* and 42 of *Columba livia*) of different age groups and of both sexes were collected randomly selected from Samawah markets in Muthanna province, south of Iraq. The process of collecting pigeon samples were extended during the period from February to September 2017.

Samples Collection and Parasitological Examinations: Postmortem examinations were performed according to¹⁵. Occasionally, cestoda were very adherent to the intestines that have been cut off with the intestine wall then preserved them in hot water till passive separation¹⁶. The worms were preserved in 4% formalin according to¹⁷.

Histopathological Examinations and DNA Extraction: A pieces of intestines from the infected pigeons were

collected. A histological study was conducted according to¹⁸. The Mini Kit DNA QIAamp/QIAGEN/Germany were using to isolate genomic DNA from adult worm according to [19]. The PCR program were performed of 94C° at 2 min followed by 35 cycle of 94C° for 1min, 57C° for 1min and 72C° for 1min, this was followed by a final extension step at 72 °C for 7 min¹⁹.

Results

The results of the present study were revealed that only 86 out of 115 examined pigeons were infested by *Raillietina* spp during the study period from February to May 2017 at infestation rate of 74.78% (86/115). Also, the result were showed that both *S. adecaecto* and *C. livia* (males and females) were infested with different the species of *Raillietina*. The number of Cestoda which were isolated from each bird may be reach to 5-7 worms. Furthermore, the percentage of infection with *Raillietina* spp were recorded in *S. decaecto* is higher than *C. livia* in 44.34 % (51/115) and 30.43% (35/115) respectively.

The histopathological examination of small intestine of pigeons infected with *Raillietina echinobothrida* were characterized by varying degrees of degenerative changes to destruction and sloughing of epithelial layer of intestinal mucosa in heavy infections as showed in figure 3. Also, the results of this examination were showed different pathological changes like, desquamation, destruction and loss of mucous membrane of villi as showed in figure 4.



Fig. 1: *Raillietina echinobothrida* worm with alum carmine stain, Scolex

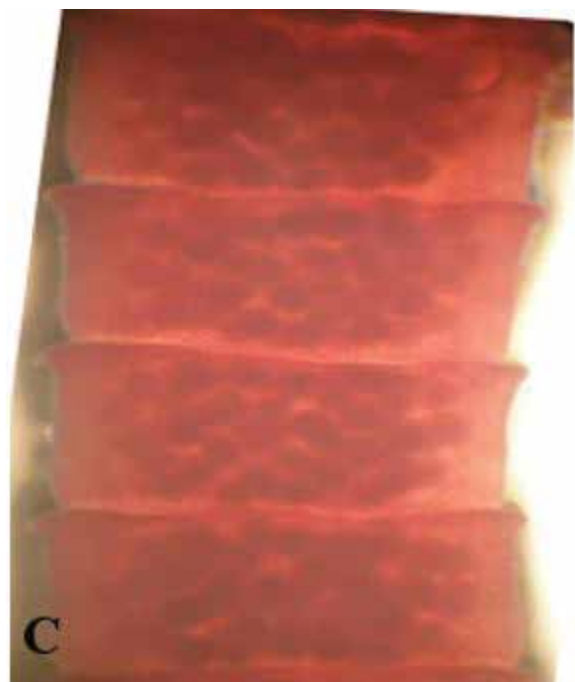


Fig. 2: *Raillietina echinobothrida* worm with alum carmine stain, Gravid segment with egg capsules

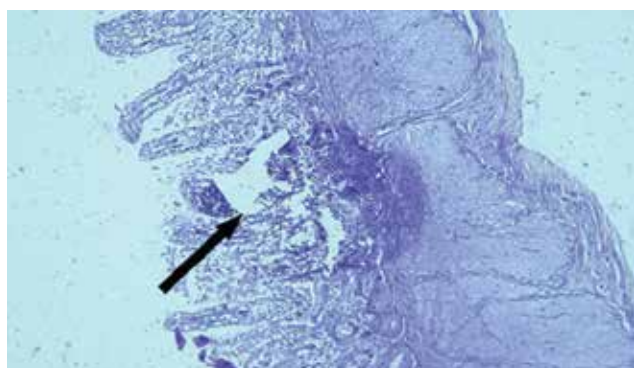


Fig. 3: Small intestine of pigeons infected with *Raillietina echinobothrida* showed destruction and sloughing of epithelial layer of intestinal mucosa in heavy infections

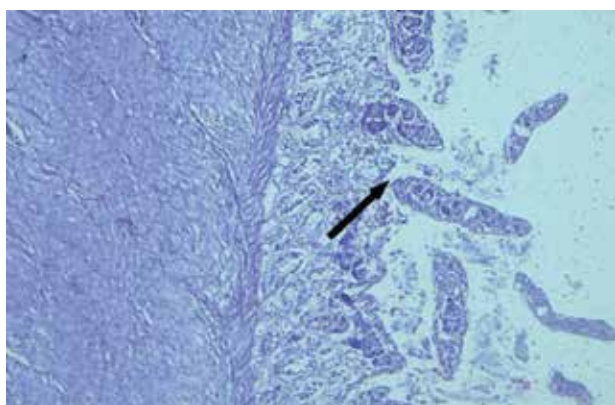


Fig. 4: Small intestine of pigeons infected with *Raillietina echinobothrida* showed destruction and desquamation of villi

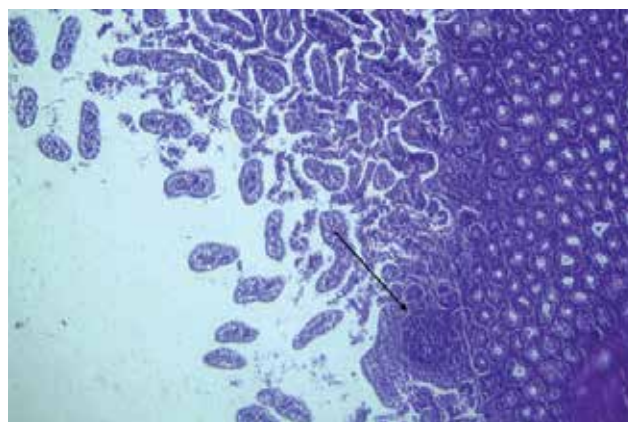


Fig. 5: small intestine of pigeons infected with *Raillietina echinobothrida* showed infiltration with heterophils throughout the mucosa, especially in the lamina propria

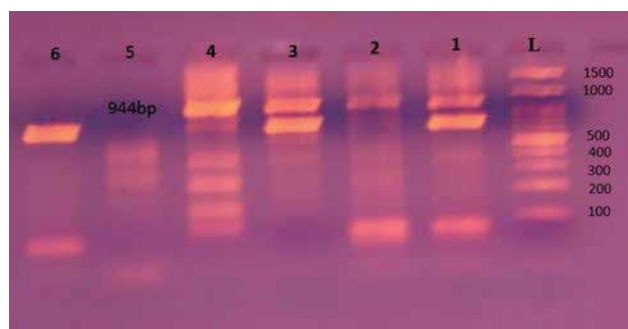


Fig. 6: Agarose gel electrophoresis of amplified DNA of *R. echinobothrida* by using primers (3S-F, BD2-R) region of region of 28S, Lane L DNA ladder, Lanes 1,2,3,4, positive to 944 bp.

Discussion

The result in the present study were revealed that only 86 out of 115 examined pigeons were infested by *Raillietina* spp during the study period from February to May 2017 at infestation rate of 74.78% (86/115). Also, the results in this study were recorded and confirm that *Raillietina* spp spread throughout the months of the year with high prevalence. This result agreed with most studies as [21,22, 23, 24]. But, this result was disagreed with [25, 26] who reported rate of infection 25.3%, and 17.4% respectively.

In addition, the spread of tapeworms and presence in all seasons of the year is the result of a natural of life cycle of the tapeworm that transmitted from birds to another through contamination feed and water with mature segment led to easily spread between birds^{24,27,28}. Finally, there are many studies confirm that changes in the climate factors like temperature or humidity had no directly effect

in incidence infection and severity, therefore *Raillietina* spp is found in all months the year^{24,29}.

The results were agreement with many previous studies^{31,32,33} and disagreement with³⁴. This agreement between the studies were recorded that the male's bird were more infected than females may be because consuming greater quantities of food as result to fly away from the nest and for long distance, While, the females remains near the nest near her eggs. Also, the females are concentrated in the food on the limestone material that need in building egg shells which reduce the handling of food^{31,32,35}.

The duodenum and jejunum cavities of *Raillietina* spp infested birds was congested and filled with greenish or yellowish feces with a very soft to liquid consistency and containing much mucous exudate. In cases with a heavy infection of parasites the mucosa was thickened and also revealed hemorrhagic spots. These finding are agreement with other studies^{24,26}. The explanation of results the gross examination in the present study is attributed to the adhesion site of adult worms in the small intestine by scolex and change his location in wall of intestine continuously and the worms migrated up and down the intestinal lumen away from the congestion to get nutrition when present in large numbers and also aggregate in the lower half of the intestine. Moreover, probably this lesion is results from anther secondary bacterial infection in the intestine due to destruction of intestinal epithelium by parasites, hemorrhage have been noticed in others may be due to irritation of heavy infection²⁸.

On the other side, the histopathological examination of the small intestine of pigeons infected with *Raillietina echinobothrida* were showed necrosis of villous epithelium, massive hemorrhages and infiltration of inflammatory cells in the lamina propria and sub mucosa. These results are agreement with previously studies^{24,28}. Each parasite have a degree of adverse impact on host and may be a change in texture of tissue, or take change in overall impact. The parasites occurs a damage to their hosts like; injurer and metabolic outcomes affecting enzymes and hormones of the host or consume food the host^{31,36}.

Also In the present study the reason to chosen these sets of primers may be attributed to the constancy and stability of the second internal transcribed spacer (ITS2) region of ribosomal DNA gene makes them a prominent target for species variation. In addition, the

gene sequences of these locations have been developed for molecular phylogenetic studies³⁸.

Acknowledgments

The authors are grateful to Departments of Veterinary Microbiology and Parasitology, College of Veterinary Medicine, Al-Muthanna University- Iraq for providing the facilities.

Conflict of Interest: This research is a personal non-profit work and there is no conflict of interest.

Source of Funding: None.

Ethical Clearance: Ethical clearance was obtained from the Faculty Scientific Committee (Department of Anatomy College of Veterinary Medicine/AL Muthanna University/Iraq,) to study the histopathological and molecular of *Raillietina echinobothrida* of domestic and wild pigeons in Al-Muthanna Province

REFERENCES

1. Anonymous, Iraqi fourth National Report to the Diversity. Ministry of Environment Republic of Iraqi. (2010). 160pp.
2. Puttalakshamma GC, Ananda KJ, Prathiush PR, Mamatha GS, Rao S. Prevalence of gastrointestinal parasites of poultry in and around Banglore. *Vet. World*, (2008). 1(7): 201–202.
3. Harlin RW. Pigeons. *Vet. Clin. N. Am. Small Anim. Pract*, (1994). 24: 157–173.
4. Radfar MH, Fathi S, Asl EN, Dehaghi MM, Seghinsara HR A survey of parasites of domestic pigeons (*Columba livia domestica*) in South Khorasan, Iran. *Vet. Res.*, (2011). 4(1): 18–23.
5. Clements, J. F. *Birds of the World: a Checklist*. Cornell University Press. (2000). p. 880.
6. Imura T, Suzuki Y, Ejiri H, Sato Y, Ishida K, Sumiyama D, Murata K, Yukawa M. Prevalence of avian haematozoa in wild birds in a high-altitude forest in Japan. *Vet. Parasitol.*, (2012). 183: 244–248.
7. Badparva E, Ezatpour B, Azami M, Badparva M. First report of birds' infection by intestinal parasites in Khorramabad, west Iran. *J. Parasit. Dis.*, (2015). 39(4): 720–724.

8. Hoberg EP, Mariaux J, Justine JL, Brooks DR, Weekes PJ. Phylogeny of the orders of the Eucestoda (Cercomeromorphae) based on comparative morphology: historical perspectives and a new working hypothesis. *J Parasitol.* (1997) 83:1128–1147.
9. Ramnath, Jyrwa, D.B., A.K. Dutta, B. Das and V. Tandon. Molecular characterization of the Indian poultry nodular tapeworm, *Raillietina echinobothrida* (Cestoda: Cyclophyllidea Davaineidae) based on rDNA internal transcribed spacer 2 region, *Journal of parasitic diseases: official organ of the Indian Society for Parasitology*, (2014). 38: 22-26.
10. Morishita, T.Y. and Schaul, J. C. Parasites of birds. In Baker DG. Flynn's Parasites of Laboratory Animals (2 Ed.). Iowa (US): Blackwell Publishers. (2008). pp. 238–239.
11. Waghmare S, Sherkhane AS, Chavan R, Gomase V. Redescription on *Raillietina echinobothrida* (Pasquale, 1890) (Cestoda: Davaineidae) and Study of Conserved Domain across Divergent Phylogenetic Lineages of Class Cestoda. *J Veterinar Sci Technol* (2014). 5: 187. doi:10.4172/2157-7579.1000187.
12. Radfar MH, Asl EN, Seghinsara HR, Dehaghi MM, Fathi S. Biodiversity and prevalence of parasites of domestic pigeons (*Columba livia domestica*) in a selected semiarid zone of South Khorasan, Iran. *Trop. Anim. Health Prod.*, (2012). 44: 225–229.
13. Small L. Internal parasites (worms) of poultry. *Agnote*, (1996). 669, 1-4.
14. Brooks DR, Hoberg EP. Parasite systematics in the 21st century: opportunities and obstacles. *Trends Parasitol* (2001). 17: 273–275.
15. Fowler NG. How to carry out a field investigation. In: Jordan ETW, Pallison M (eds) *Poultry diseases*, 4th ed. Saunders WB Company, London, (1996), pp422-456.
16. Charles, MH. *Diagnostic Veterinary Parasitology*. 2nd ed. Mosby, Elsevier.; (1998). pp:246-257
17. Soulsby, E. J. L. *Helminths, Arthropods and Protozoa of Domesticated animals*. 7th ed. Bailliere Tindall, London, UK. (1982)
18. Bancroft, J. D., M. Gamble. *Theory and Practice of Histological Techniques*. 5th ed., Harcourt Publ. Ltd., London. (2002). P125-131.
19. Butboonchool, P., Wongsawad, C., Rojanapaibul, A., and Jong-Yil, C Morphology and Molecular Phylogeny of *Raillietina* spp. (Cestoda: Cyclophyllidea: Davaineidae) from Domestic Chickens in Thailand, *Korean J Parasitol* (2016). Vol. 54, No. 6: 777-786.
20. Permin, A and Hansen, J, W. *The Epidemiology, Diagnosis and Control of Poultry Parasites: An FAO Handbook*. Food and Agriculture Organization of the United Nations, Rome, Italy, (2003). pp. 6-8.
21. Lalchhandama, K. On the structure of *Raillietina echinobothrida*, the tapeworm of domestic fowl. *Sci Vis* (2009). 9 (4), 174-182.
22. Fotedar, D. N., N. G. Khateeb Occurrence and seasonal variation of helminthes parasites of domestic fowl in Kashmir. *Indian J. Helminthol.* (1986). 38, 49-54.
23. Pandit, B. A., A. S. MIR, M. A. A. Banday, R. A. Shahardar Prevalence of helminth parasites in Indigenous fowls of Kashmir Valley. *Poult. Adv.* XXIV(X), (1991). 37-39.
24. Jasim, H. Z. *Diagnosis of Cestodes and Chewing Lice That Infest on Wild and Domestic Pigeons in Basrah Province*. M. Sc. Thesis., College of Veterinary Medicine, University of Basrah. (2017). Pp: 3-10.
25. Vazquez, B.; Esperon, F.; Neves, E.; Lopez J.; Ballesteros, C. and Munoz V. screening for several potential pathogens in feral pigeons (*Columba livia*) in Madrid. *Acta. Veterinaria Scandinavica.* (2010). 52:45-50.
26. Al-Marsomy A. W. and Al-Hamadaani S. H. Association of Cestoda *Raillietina echinobothrida* in Rock Pigeon *Columba livia* from Baghdad city of Iraq. *Baghdad Science Journal*, (2016). 13(3) Pp:463-468.
27. Klein, R.; Bartel, M.; Paulus, M.; Quack, M.; Tarricone, K.; Wagner, G.; Ball, M.; Ru"del, H.; and Schlu"ter, C. Pollution of urban industrial ecosystems in Germany—the use of bioindicators from different trophic levels. *Environ Bioindic* (2008). 3:W19.

28. Abed, A. A; Naji, A.H and, Rhyaf G.A. Investigation study of some parasites infected domestic pigeon (*Columba livia domestica*) in Al-Dewaniya city. IOSR Journal of Pharmacy and Biological Sciences, (2014). 9 (4), 13-20.
29. Al-Bayati, N. Y. A study on pigeons (*Columba livia*) Cestodes infection in Diyala Province. Diyala Agri. Sci. J. (2011). 3 (2):1-12.
30. El-Dakhly K.M, Mahrous L.N, Mabrouk G.A. Distribution pattern of intestinal helminths in domestic pigeons (*Columba livia domestica*) and turkeys (*Meleagris gallopavo*) in Beni-Suef province, Egypt. J. Vet. Med. Res. (2016); 23(1):112–120
31. Mustafa, F. A. Epidemiological study of some cestodes infected digestive system of pigeon. M. Sc. Thesis. Coll. of Sci., Univ. of Basrah. (1984). Pp: 113. (In Arabic).
32. Fedynich, A. M.; Finger, R.S.; Ballard, B. M.; Garvon, J.M. and Mayfield, M. J. Helminthes of Ross and greater white-fronted geese wintering in south Texas, U.S.A. Comp. Parasitol., (2005). 73:33-38.
33. Shubber, H. W. K. The parasitic helminths of the Digestive tract of the ducks *Nettarufina* and *Anas crecca*. M. Sc. Thesis, Univ. AL-Qadisiyah (in Arabic). (2006).
34. Al – Aloosi, J. A. Survey of alimentary canal helminth of two birds Gull (*Larus ridibundus*) and wood pigeon (*Columba palumbus*) from Baghdad and Baiji regions M. Sc. Thesis, Univ. of Baghdad Iraq. (1985). 124 pp
35. Rajvanshi, I. and Gupta, A. N. Qualitative and Quantitative Analysis of Digenetic Trematodes Fauna in Cattle Egret, *Bubulcus ibis coromandus*. Proceedings of the Indian Academy of Parasitology, (1983). 4(1/2):1-5.
36. Hungerford, T. G. Diseases of poultry including cage birds and pigeons. Section 9 internal parasites of poultry pp. 483-499. Angus and Robertson Ltd., 4th ed. (1969).
37. Ghobashy A. M. and Taeleb A.A. Molecular Characterization of *Raillietina*(r.) Spp. Ortlepp, 1938 (Cestoda: Cyclophyllidae: Davaineidae) Infecting Domestic and Wild Birds (*Columba livia* And *Columba livia domestica*). World Journal of Zoology (2015). 10 (2): 136-141.
38. Butboonchoo P, Wongsawad C. Occurrence and HAT-RAPD analysis of gastrointestinal helminths in domestic chickens (*Gallus gallus domesticus*) in Phayao province, northern Thailand. Saudi J Biol Sci (in press). (2015).