The Effect of Follicular Fluid Supplementation on Embryo **Development in Iraqi Buffalo**

Husamaldeen A. Alsalim^{a*}, Ihsan A. Habeeb^a, Ahzan K. Abdulameer^b

^aDepartment of Theriogenology, College of Veterinary Medicine, University of Basrah, Iraq ^bDepartment of Theriogenology, College of Veterinary Medicine, University of, kufa, Iraq husamaldeen1976@gmail.com

Abstract

The study was carried out to evaluate the effect of Keywords: Buffalo, follicular fluid, Embryo development supplementation of the buffalo follicular fluid (buFF) in culture medium (Ham's F-10) on the development of oocytes in vitro maturation (IVM) rate, oocytes in vitro fertilization (IVF) rates, and embryonic developmental rate. The experimental study extended from November 2019 to March 2020. Samples included (80) female ovary and (20) bull testes was a collection from Basrah center slaughterhouse after 20 minutes from slaughter. Harvested samples were transported by disinfected clean box at (37 °C) in less than 1 hrs to the lab. Only A and B grads quality oocytes were selected and incubated in a prepared maturation medium. Spermatozoa were obtained from slicing caudal of epididymal. Matured oocytes and capacitated sperms were incubated fertilization medium. Fertilize oocytes were re-incubated in fertilization medium with changing 50% of the medium every day followed by an embryonic developmental assessment for 4 days. Results illustrated a significant increase in oocytes maturation rate in Ham F-10 media with a 20% buFF group compared control group. On the other hand, this study showed a significant increase in fertilization rate in Ham's F-10 medium with a 20% buFF group compared with the control group. Moreover, a high significant difference in embryonic development (2, 4,8,16 cells) in Ham's F-10 media with 20% buffalo follicular fluid group compared with the control group. Thus, It has been concluded that supplementation of 20% buFF to media could be enhanced in vitro oocyte maturation, fertilization, and in vitro embryonic development in buffalo.

Introduction

The zoological scale of buffalo in the class (Mammalia) order (Artiodactyla), Ruminantia (subordinate), family (Bovidae), sub-family (Bovinae), gender (Bubalus), species (Bubalus bubalis). Buffaloes are a very important resource in the agricultural economy in Iraq and some other countries such as India, China, and Pakistan Egypt, and Ext^[1]. Riverine and Swamp buffalo are an irreplaceable milk producer, resistant to climate, parasite, stress, diseases, and the rustic and high ability for acclimation in climatic zones ^[2]. However, there are many problems in buffalos like delayed reproductive maturity, reproductive seasonality, calving interval long, silent estrus, low follicles, and poor super ovulatory response have been referred to low reproductive effecting of this species [3]. Therefore, the assisted reproductive technologies (ART) are very important to improve the genetic potential for this species ^[1].

Oocytes maturation is a very important step in the IVF process. Many studies were done to improve the in vitro maturation in mammal's oocytes ^[4]. Supplementation of in vitro maturation medium with hormones and different concentrations of serum plays an important role in embryo development ^[5]. Oocytes maturation consists of two steps: cytoplasm and nuclear maturation [6]. Bovine oocytes maturation is affected by several factors, such as time of oocytes collection, transportation temperature and follicular size [7], stage of oocyte development [8], the diameter of the

oocyte ^[9], media composition ^[10], hormonal supplementation ^[11] serum addition like fetal calve serum (FCS) or bovine serum albumin (BSA) [12].

Buffalo follicular fluid is one of the alternatives of macromolecules that can be used as the supplementation in the maturation media because it is easy to get and cheap. Follicular fluid is a liquid semi-viscous and yellow in color, which dictates the follicle and surrounding oocvtes [13]. Buffalo follicular fluid consists of many growth factors, follicle-stimulating hormone (FSH), luteinizing hormone (LH), several nutrients, and rich in hyaluronic acid [13]. Follicular fluid is a result of the transport of blood plasma components that cross the follicular barrier and secretory activity of granulosa and theca cells [14].

Analysis of the biochemical properties of the follicular fluid surrounding the eggs plays important role in determining the oocyte's quality and subsequent potential for fertilization and fetal development ^[15]. The analysis of the components of the follicular fluid provides information about metabolic changes in the blood serum, as the circulating biochemical environment may be reflected in the formation of the follicular fluid [16]. Several studies of the chemical components of follicular fluids have been grouped into the following categories hormones, growth factors in the TGFbeta family, other growth factors and interleukins, reactive oxygen species (ROS), proteins, amino acids, peptides, sugars, anti-apoptotic factors, and prostanoids [16]. Limited

The Effect of Follicular Fluid Supplementation on Embryo Development in Iraqi

Buffalo

studies were conducted to examine the effect of buffalo follicular fluid on in vitro fertilization in buffalo, so we designed this study to evaluate the effect of supplementation the buffalo follicular fluid on oocytes maturation, oocytes fertilization, and embryos production from buffalo oocytes.

Materials and methods

Media and reagents

All chemicals and media were obtained from Sigma Chemical Company (St. Louis, MO, USA) and Gibco (Invitrogen Corporation, Grand Island, NY, USA), respectively. All the experiments were carried out in the central laboratory, university of Basrah, college of veterinary medicine.

Recovery and in vitro maturation of buffalo oocytes

Immature buffalo cumulus oocytes complexes were aspirated from 0.8-1 mm follicles of slaughterhouse ovaries by using an 18-gauge needle connected with a syringe. The recovered oocytes were transferred into Ham's F-10 supplemented with 10% fetal calf serum (FCS). Oocytes with clear cytoplasm and with many layers of cumulus cells were selected for in vitro maturation ^[17] and incubated for 20 h in tissue culture medium at 38.5 °C, 5% CO2, and maximum humidity [18]. The oocytes divided into 2 groups, the first group (control group) was incubated inside in vitro maturation medium (Ham's F-10 plus 10% FCS) for 20 h at 38.5 °C, 5% CO2, and maximum humidity [18]. While the second group (treated group) was incubated inside in vitro maturation medium which supplemented with 20% buffalo follicular fluid for 20 h at 38.5 °C, 5% CO2, and maximum humidity. Oocyte maturation was recorded through the presence of cumulus cells expansion as well as the presence of the first polar body as described by ^[19].

Buffalo follicular fluid preparation

Buffalo follicular fluid (buFF) was collected by aspiration from pre-ovulatory follicles (1-2 cm in diameter) containing oocytes during the reproductive season. Follicular fluids (10– 20 ml) for each sample were centrifuged twice at 1000 g for 15 min to removed cellular debris and transported to a sterilized

glass beaker for temperature inactivation at 50 °C for 30 Moreover, the percert reated group with 2 sterilized by using syringe driven filter 0.22mm and then was kept in sterilization test tubes 1.5ml capacity at 20 oC for Table 1. The effect of buffalo follicular fluid on In vitro maturation (IVM) of buffalo occytes (IVM) of buffalo occytes (IVM) of buffalo occytes (IVM) of buffalo occettes (IVM) occettes (IVM) of buffalo occe

using within vitro maturation, in vitro fertilization, and in vitro culture medium development embryo ^[20].

Sperm maturation and Capacitation

Sperms was opined from slicing tail of epididymal of buffalo's bull testis. The sperm samples with 2ml of tissue culture media (TCM-199) with anti-fungal (Nystatin) and anti-bacterial (Penicillin) transported from petri dish to 10ml test tube , Samples incubated in (37C°) for 4 hrs. The descending protoplasmic droplet from tail of sperm was good signs for maturation ^[21]. The end step sperm capacitation was added 50 IU/ml heparin at the last 45minutes from time of sperm maturation ^[22].

In vitro fertilization and culture

Matured oocytes rinsed twice in Ham's F-10 medium before transmission to IVF Petri-dishes contain IVF medium (Ham's F-10) containing 20% buFF, 10% FCS, 100 IU/ml penicillin-streptomycin and100 IU/mL Nystatin [23]. While the control group was left without buFF. For in-vitro fertilization, approximately $1-2 \times 106$ capacitated sperms were transferred for each matured oocyte inside the IVF medium [23]. Petri- dishes containing capacitated sperms plus matured oocytes were incubated at 38.5 °C, 5% CO2, and maximum humidity ^[24, 25]. One day after, the fertilization rate was recorded. In the following days, 50% of the medium was changed daily for 4 days and during this time the cleavage rate and embryo development was recorded according to ^[26].

Statistical Analysis

Chi-square test was used to compare maturation and fertilization rates among different groups [27]. All data were presented as means \pm SEM and the differences were considered as significant at P < 0.05.

Results

Effect of buffalo follicular fluid on maturation of buffalo oocytes

The current study revealed that the oocyte maturation rate was increased significantly (P<0.05) in the treated group when supplemented with 20% buFF compared with the control group 77.78 \pm 4.06, 38.89 \pm 9.09 respectively (figure 1). Moreover, the percentage of non- matured oocytes in the treated group with 20% buFF was decreased significantly (P<0.05) in compared control group 22.22 \pm 4.06, 61.11 \pm 9.09 respectively (table 1).

Groups	Ovaries No.	Cultured oocytes No.	Matured oocytes	Non matured oocytes
Control group	40	90	35 (38.89 ± 9.09) a	55 (61.11 ± 9.09) a
Treated group	40	90	$70(77.78 \pm 4.06)$ b	20 (22.22 ± 4.06) b

Data are presented as number (percentage \pm SEM). ^{ab} Different letters within each column indicate significant difference (p < 0.05)

The effect buffalo follicular fluid on *In vitro* fertilization (IVF) in buffalo

The present study showed that the fertilized oocytes were increased significantly (P<0.05) in the treated group with 20% buFF in comparison with the control group 84.29±2.64, **Table 2.** The effect of buffalo follicular fluid on IVF in buffalo oocytes

45.44 \pm 6.72 respectively (figure 2, 3 and Table 2). Non fertilized oocytes were increased significantly (P<0.05) in the control media group in comparison with a treated group that supplemented with 20% buFF 54.55 \pm 6.72, 15.70 \pm 2.64 respectively. While the percentage of embryos development

increased significantly (P< 0.05) in the treated group with 20% buFF 37.50±4.95 in compared with the control group15.32±3.63 (table 2).

Groups	Matured oocytes Fertilized oocytes		Cleaved oocytes	Non fertilized oocytes					
Control group	35 (38.89±9.09) a	17 (45.44 ±6.72) a	3 (15.32±3.63) a	18 (54.55±6.72) a					
Treated group	70 (77.78±4.06) b	59 (84.29±2.64) b	22 (37.50±4.95) b	11 (15.70±2.64) b					

Data are presented as number (percentage \pm SEM). ^{ab} Different letters within each column indicate significant

difference (p < 0.05) The effect of buffalo follicular fluid on embryo

The Effect of Follicular Fluid Supplementation on Embryo Development in Iraqi

Buffalo

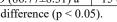
development in buffalo

The present study showed the embryonic development (2, 4,

8, and 16 cells) (figure 4, 5, 6, 7) significantly (P<0.05) higher in the treated group which supplemented with 20% buFF in compared with the control group (table 3).

Table 3. The effect of buffalo follicular fluid on embryos development in buffalo IVF.									
Groups	Fertilized oocytes	2-cell embryo	4-cell embryo	8-cell embryo	8-cell embryo				
Control group	17 (45.44±6.72) a	3 (15.32 ± 3.63) a	0 a	0 a	0 a				
Treated group	59 (84.29±2.64) b	22 (37.50±4.95) b	19 (86.77±8.31) a	15 (78.05±6.09) a	12 (82.14±8.98) a				
Data are present	ted as number (perce	entage ± SEM). ^{ab}	difference ($p < 0.0$)	5).					

Different letters within each column indicate significant



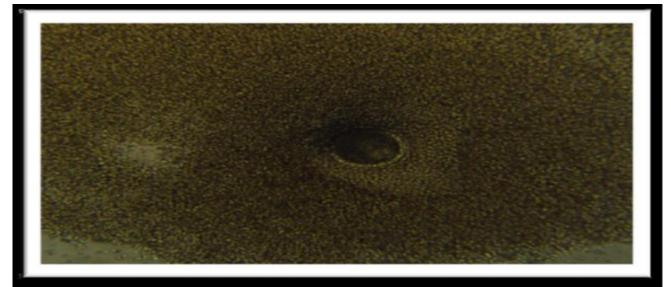


Figure 1. Matured Oocyte with cumulus cells expansions 10X

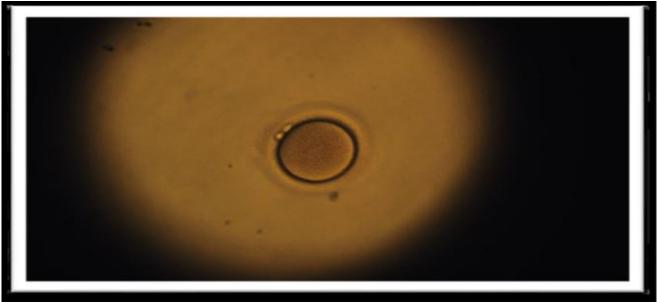


Figure 2. Fertilization Oocyte with two polar body - normal fertilization (10X)

The Effect of Follicular Fluid Supplementation on Embryo Development in Iraqi Buffalo

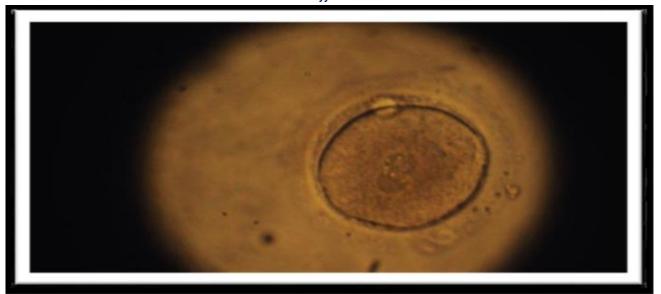


Figure 3. Fertilization Oocyte with two pronuclear - normal fertilization (10X)



Figure 4. 2-cells embryo resulted from IVF in buffalo (10X)

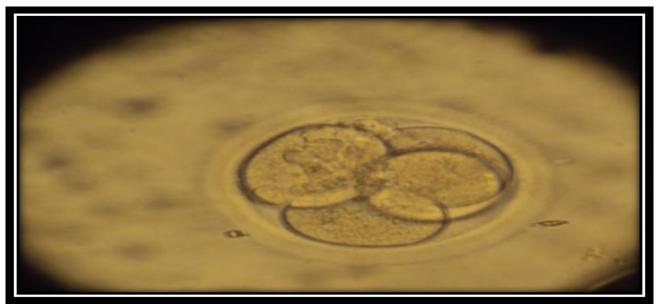


Figure 5. 4-cells embryo resulted from IVF in buffalo (10X)

The Effect of Follicular Fluid Supplementation on Embryo Development in Iraqi Buffalo

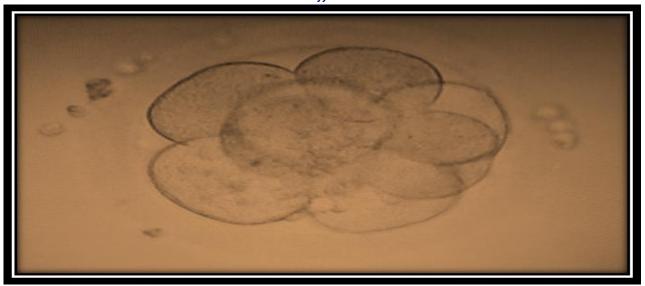


Figure 6. 8-cells embryo resulted from IVF in buffalo (10X)

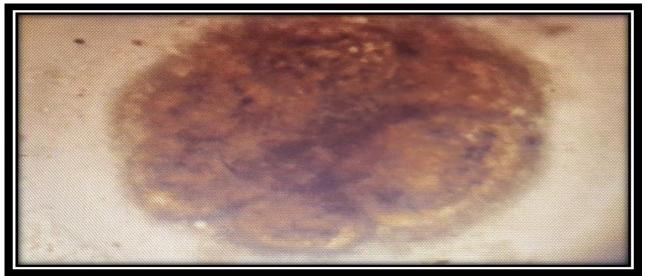


Figure 7. 16-cell embryo resulted from IVF in buffalo (10X)

Discussion

The current study showed that the maturation and fertilization rate of COCs with a buFF supplemented medium and improve the developmental rate of buffalo embryo by providing a positive action on the development of buffalo embryos from 2-cell to 16-cell. There is an important need to use the follicular fluid in the culture medium which contains various growth factors, hormones such as FSH, LH, moreover, it contains several nutrients and also rich with hyaluronic acid [13] that help in the process of development of embryos in buffalo IVF system. The study revealed that 20% of buffalo follicular fluid had a positive effect on oocytes maturation, fertilization, and subsequent embryo development in the treated group compared with the control group. This result is in agreement with previous studies which studied the effected of follicular fluid supplementation with IVM medium in other species.

In pigs, the use of a follicular fluid with a maturation medium enhanced oocytes maturation levels and promoted the formation rate of female and male nuclei ^[28]. The previous report have compared the effect of estrous cow serum (ECS) and follicular fluid in bovine IVF, it showed that presence of follicular fluid in IVM-IVF culture medium enhances the oocytes maturation rate, fertilization rate and improving the morula and blastocysts rate ^{[29][30]} Suggested

the beneficial effect of follicular fluid on the maturation of buffalo oocyte due to the existence of insulin-like growth factor, growth factors, hormones (FSH, LH, and estradiol) and intra-ovarian peptides in more physiological properties in follicular fluid. On the other hand ^[31, 32] Suggested that the buffalo follicular fluid in culture medium enhances cytoplasmic and nuclear maturation. ^[33] Revealed that the fluid from normal follicle contains a balanced proportion of estradiol/progesterone (EPR) and insulin-like growth factor binding proteins (IGFBPs) enhance oocytes maturation and fertilization rates.

Another report mentioned the positive effect of using buffalo follicular fluid with IVF medium which promotes sperm penetration as well as improves cumulus cell expansion and enhancing ATP levels in oocytes ^[34,35] Fixed that the follicular fluid contains (electrolytes, hormones, amino acids, growth factors, and other components) which used as a natural material for blocking the meiosis ^[36]. Another study has documented that maturation medium supplemented with follicular fluid supplied an appropriate environment for oocyte development ^[37]. It also increases the degree of cumulus cell expansion and enhances embryonic development ^[38]. The addition of follicular fluid to the maturation medium had beneficial effects for the maturation and development ability of bovine oocytes ^{[39]. [40]} Suggested

The Effect of Follicular Fluid Supplementation on Embryo Development in Iraqi

Buffalo

that buffalo follicular fluid catalyze the synthesis protein cell cycle in embryos which in turn supported their development, the presence of enzymes like lactate dehydrogenase, ATPase, transaminase, and alkaline phosphatase, glycosaminoglycan's, proteins and steroids, gonadotropins antioxidants, might have promoted the maturation oocytes, cleavage rates and development embryo [41,42].

Acknowledgement

Authors are thankful to meat inspectors at Basrah abattoir for their great and kind assistance in the collection of different samples.

References

- 1. Nandi S, Ravindranatha BM, Gupta PSP, Sarma PV. Timing of sequential changes in cumulus cells and first polar body extrusion during in vitro maturation of buffalo oocytes. Theriogenology. 2002; 57(3): 1151-1159.
- 2. Haldar A, Prakash BS. Effect of exogenous growth-hormone-releasing factor on blood metabolites and minerals in late maturing buffalo heifers (Bubalus bubalis). Journal of animal physiology and animal nutrition. 2007; 91(7-8): 326-332.
- Hufana-Duran D, Pedro PB, Venturina HV, Duran PG, Cruz LC. Full-term delivery of river buffalo calves (2n= 50) from in vitro-derived vitrified embryos by swamp buffalo recipients (2n= 48). Livestock Science. 2007; 107(2-3): 213-219.
- 4. Pawshe CH, Palanisamy A, Taneja M, Jain SK, Totey SM. Comparison of various maturation treatments on in vitro maturation of goat oocytes and their early embryonic development and cell numbers. Theriogenology. 1996; 46(6): 971-982.
- 5. Kharche SD, Sharma GT, Majumdar AC. In vitro maturation and fertilization of goat oocytes vitrified at the germinal vesicle stage. Small Ruminant Research. 2005; 57(1): 81-84.
- 6. Blondin P, Sirard MA. Oocyte and follicular morphology as determining characteristics for developmental competence in bovine oocytes. Molecular reproduction and development. 1995; 41(1): 54-62.
- 7. Pavlok A, Lucas-Hahn A, Niemann H. Fertilization and developmental competence of bovine oocytes derived from different categories of antral follicles. Molecular Reproduction and Development. 1992; 31(1): 63-67.
- Hagemann LJ, Beaumont SE, Berg M, Donnison MJ, Ledgard A, Peterson AJ, et al. Development during single IVP of bovine oocytes from dissected follicles: interactive effects of estrous cycle stage. Follicle size and atresia. Molecular Reproduction and Development: Incorporating Gamete Research. 1999; 53(4): 451-458.
- 9. Hyttel PFTCH, Fair T, Callesen H, Greve T. Oocyte growth, capacitation and final maturation in cattle. Theriogenology. 1997; 47(1): 23-32.
- Lonergan P, Khatir H, Carolan C, Mermillod P. Bovine blastocyst production in vitro following inhibition of oocyte meiotic resumption for 24 hours. Theriogenology; 1(47): 293.
- Zuelke KA, Bracketf BG. Luteinizing hormone-enhanced in vitro maturation of bovine oocytes with and without protein supplementation. Biology of reproduction. 1990; 43(5): 784-787.
- 12. Avery B, Bavister BD, Greve T. Development of bovine oocytes, in vitro matured in a chemically defined protein-free medium, supplemented with different amino acid formulations. Theriogenology. 1998; 1(49): 306.
- 13. Knight PG, Muttukrishna S, Groome NP. Development and application of a two-site enzyme immunoassay for the determination of total'activin-A concentrations in

serum and follicular fluid. Journal of Endocrinology. 1996; 148(2): 267-279.

- 14. Fortune JE. Ovarian follicular growth and development in mammals. Biology of reproduction. 1994 50(2): 225-232.
- 15. Carolan C, Lonergan P, Monget P, Monniaux D, Mermillod P. Effect of follicle size and quality on the ability of follicular fluid to support cytoplasmic maturation of bovine oocytes. Molecular Reproduction and Development: Incorporating Gamete Research. 1996; 43(4): 477-483.
- 16. Leroy JLMR, Vanholder T, Delanghe JR, Opsomer G, Van Soom A, Bols, PEJ, et al. Metabolic changes in follicular fluid of the dominant follicle in high-yielding dairy cows early post-partum. Theriogenology. 2004; 62(6): 1131-1143.
- Raza A, Samad HA, Rehman NU, Zia EUH. Studies on in vitro maturation and fertilization of Nili Ravi buffalo follicular oocytes. Int. J. Agric. Biol. 2001; 3(1): 503-506.
- Leibfried L, First NL. Characterization of bovine follicular oocytes and their ability to mature in vitro. Journal of Animal Science 1979; 48(1): 76-86.
- Hunter AG, Moor RM. Stage-dependent effects of inhibiting ribonucleic acids and protein synthesis on meiotic maturation of bovine oocytes in vitro. Journal of dairy science. 1987; 70(8): 1646-1651.
- 20. Dell'Aquila ME, Cho YS, Minoia P, Traina V, Lacalandra GM, Maritato F. Effects of follicular fluid supplementation of in-vitro maturation medium on the fertilization and development of equine oocytes after invitro fertilization or intracytoplasmic sperm injection. Human reproduction (Oxford, England). 1997; 12(12): 2766-2772.
- Gadella BM, Luna C. Cell biology and functional dynamics of the mammalian sperm surface. Theriogenology. 2014; 81(1): 74-84.
- 22. Manjunath P, Thérien I. Role of seminal plasma phospholipid-binding proteins in sperm membrane lipid modification that occurs during capacitation. Journal of reproductive immunology. (2002); 53(1-2): 109-119.
- 23. Galli C, Crotti G, Notari C, Turini P, Duchi R, Lazzari G. Embryo production by ovum pick up from live donors. Theriogenology. (2001); 55(6): 1341-1357.
- 24. Kobayashi K, Yamashita S, Hoshi H. Influence of epidermal growth factor and transforming growth factor- α on in vitro maturation of cumulus cell-enclosed bovine oocytes in a defined medium. Reproduction. (1994); 100(2): 439-446.
- 25. Anderiesz C, Ferraretti AP, Magli C, Fiorentino A, Fortini D, Gianaroli L, et al. Effect of recombinant human gonadotrophins on human, bovine and murine oocyte meiosis, fertilization and embryonic development in vitro. Human Reproduction. 2000; 15(5): 1140-1148.
- 26. Iritani A, Kasai M, Niwa K, Song HB. Fertilization in vitro of cattle follicular oocytes with ejaculated spermatozoa capacitated in a chemically defined medium. Reproduction. 1984; 70(2): 487-492.
- Schefler WC. (1979). Statistics for the Biological Sciences.Upper Saddle River, NJ: Addison-Wesley. 103-120.
- Naito K, Fukuda Y, Toyoda Y. Effects of porcine follicular fluid on male pronucleus formation in porcine oocytes matured in vitro. Gamete research. 1988; 21(3): 289-295.
- 29. Larocca C. Effect of follicular fluid and estrus cow serum on maturation, fertilization and development of the bovine oocyte in vitro. Theriogenology. 1993; 39(1): 253.
- 30. Cognié Y, Poulin N, Locatelli Y, Mermillod P. State-of-

The Effect of Follicular Fluid Supplementation on Embryo Development in Iragi

Buffalo

the-art production, conservation and transfer of in-vitroproduced embryos in small ruminants. Reproduction, Fertility and Development. 2004; 16(4): 437-445.

- 31. Chauhan MS, Palta P, Das SK, Katiyar PK, Madan ML. Replacement of serum and hormone additives with follicular fluid in the IVM medium: effects on maturation, fertilization and subsequent development of buffalo oocytes in vitro. Theriogenology. 1997; 48(3): 461-469.
- Aquino FP, Ocampo MB. In vitro maturation of bubaline oocytes using bubaline follicular fluid. European J Biotech BioSci. 2014; 2(1): 35-38.
- 33. Nandi S, Raghu HM. Ravindranatha, B. M., Gupta, P. S. P., & Sarma, P. V. In vitro development of buffalo oocytes in media-containing fluids from different size class follicles. Reproduction in domestic animals. 2004; 39(1): 33-38.
- 34. Somfai T, Inaba Y, Watanabe S, Geshi M, Nagai T. Follicular fluid supplementation during in vitro maturation promotes sperm penetration in bovine oocytes by enhancing cumulus expansion and increasing mitochondrial activity in oocytes. Reproduction, Fertility and Development. 2012; 24(5): 743-752.
- 35. Larocca C. Effect of follicular fluid and estrus cow serum on maturation, fertilization and development of the bovine oocyte in vitro. Theriogenology. 1993; 39(1): 253.
- 36. Aguilar JJ, Woods GL, Miragaya MH, Olsen LM, Vanderwall DK. Effect of homologous preovulatory follicular fluid on in vitro maturation of equine cumulusoocyte complexes. Theriogenology. 2001; 56(5): 745-758.
- 37. Romero-Arredondo A, Seidel GE. Effects of follicular fluid during in vitro maturation of bovine oocytes on in vitro fertilization and early embryonic development. Biology of Reproduction. 1996; 55(5): 1012-1016.
- Aguilar JJ, Woods GL, Miragaya MH, Olsen LM, Vanderwall DK. Effect of homologous preovulatory follicular fluid on in vitro maturation of equine cumulusoocyte complexes. Theriogenology. 2001; 56(5): 745-758.
- 39. Algriany O, Bevers M, Schoevers E, Colenbrander B, Dieleman S. Follicle size-dependent effects of sow follicular fluid on in vitro cumulus expansion, nuclear maturation and blastocyst formation of sow cumulus oocytes complexes. Theriogenology. 2004; 62(8): 1483-1497.
- 40. Chi HJ, Kim DH, Koo JJ, Chang SS. The suitability and efficiency of human follicular fluid as a protein supplement in human in vitro fertilization programs. Fertility and sterility. 1998; 70(5): 871-877.
- Caucig H, Friedrich F, Hager R, Golob E. Enzymes in follicular and cyst fluid in human ovaries. Acta Endocr. 1971; 66(1): 52.
- 42. Hussein HA, Abd Ellah MR, Derar DR. Antioxidant concentrations in serum, follicular fluid, and corpus luteum of cyclic buffalo cows. Comparative Clinical Pathology. 2013; 22(5): 829-833.