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ORIGINAL ARTICLE

Virulence factors and antibiotic susceptibility patterns of Klebsiella pneumonia strains Histamine producing bacteria isolated from sputum

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ARTICLE INFORMATIONS

ABSTRACT Objective: Histamine- producing bacteria(HPB) may be associated with

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Klebsiella pneumoniae HPLC Antibiotic Histamine Producing Bacteria (HPB) Histadine decarboxylase hdc

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Hanaa k. Ibrahim Email: <u>hanaakhlil878@yahoo.com</u> Veterinary Medicine College Basra University Iraq. important pathogenic bacteria and is most frequently recovered from clinical specimens and can cause a classic form of primary pneumonia. **Methods:** Fifty-one samples of sputum collected from patients suffering from respiratory problem in the advisory clinic for chest and respiratory disease Basra city. Isolates of *Klebsiella pneumoniae* were identified by their morphological and biochemical characteristics. And The colony was cultured on Niven's agar medium and. purple colony on Niven's medium is indicator of Histamine Producing Bacteria (HBP) according to Niven's *et al.*, 1981. All the isolates of *Klebsiella pneumoniae* identified of *hdc* gene by PCR and gene expression *hdc* detection by HPLC technique and identified were subjected to antibiotic sensitivity testing by MIC the Bio Meraux company according to16 antibiotics resistance profile and study Factors affecting Histamine production.

much respiratory problem. Klebsiella pneumoniae is one of the most

Results: Fifty one(51) samples were collected from the patients sputum suffering from respiratory problem in the advisory clinic for chest and respiratory disease and two hundred and fifty five (255) isolates were tested using phenotypic classification based on chemical properties of bacteria. (50)strains of *Klebsiella pneumoniae* were identified by their morphology and biochemical characteristics. Out of above only eleven (11, 21.56%)strain of *Klebsiella pneumoniae* were positive for histamine production depending on the Niven's medium as a histamine producer indicator and has been confirmed in PCR for the presence of the gene histadine decarboxylase *hdc*. Histamine concentration was 46.06µg/ml that produced from *Klebsiella pneumoniae* was detected by HPLC technology. Majority of the strains HPB isolated were sensitive to Trimethoprim . The proportion concentration of histamine produced from *Klebsiella pneumoniae* in pH(6.5) was 29.04µg/ml and less at a NaCl 1mg was 2.75 µg/ml. **Conclusion:** The *K. pneumonia* have a new virulence factor it can be cause or enhanced pathogenicity and high sensitive

cause or enhanced pathogenicity and high sensitive against many antibiotics Trimethoprim, while the overall resistance pattern high resistance Ampicilin, Ambicillin/Sulbactam.

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INTRODUCTION

Klebsiella pneumoniae is one of the most important pathogenic bacteria. K. pneumoniae is a part of the normal flora of humans where they inhabit mucosal surfaces. Klebsiella pneumoniae is most frequently recovered from clinical specimens and can cause a classic form of primary pneumonia. Klebsiella *pneumonia*e can also cause a variety of extra pulmonary infections, including enteritis and meningitis in infants, urinary tract infections in children and adults and septicemia¹ and nosocomial infections by *K. pneumoniae* are still much more prevalent, and may be more dangerous due to the rapid development and spread of antimicrobial resistance in hospital settings^{2,3}. Multidrug resistant bacteria cause serious nosocomial and community acquired infections that are hard to eradicate by using available antibiotics. Moreover, extensive use of broad-spectrum antibiotics in hospitalized patients has led to both increased carriage of Klebsiella and the development of multidrug-resistant strains that produce extended–spectrum beta-lactamase^{4,5}. Studies have shown that antibiotic consumption leads to selective pressure increasing beta-lactam resistance in bacteria of the genus *Enterobacteriaceae*⁶. Pathogenicity of K. pneumoniae is due to the presence of many virulence genes which encode virulence factors that allow it to attack the immune system of mammalians and cause many kind of diseases. Some of these virulence factors are: biofilm formation, hypermucoviscosity, capsule synthesis, adhesions, iron uptake and lipopolysaccharides formation^{7,8}. K. pneumoniae has been found capable to resist many antibiotics especially third generation cephalosporins like cefotaxime, ceftriaxone and Ceftazidime9. Many clinical features of K. pneumoniae infections are related with virulence gens according to number and mode of action of these genes¹⁰. A hazardous level of histamine is produced by the microbial decarboxylation and Enteric bacteria have been reported to be the dominant histamine-producing bacteria (HPB) Klebsiella pneumoniae as the primary histamine-forming microorganism¹¹.

MATERIALS AND METHDS

Sample collection and identification

Fifty-one samples of sputum collected from patients suffering from respiratory problem in the advisory clinic for chest and respiratory disease Basra city. The samples were cultured by the streaking on blood agar and MacConkey agar and then incubated at a 37° C for 24 hours. The bacterial colonies was purified and identified according to colonies morphology, color and size .The colony was cultured on Niven's agar medium. The plate were incubated at 37° C for 48hrs. purple colony on Niven's medium is indicator of Histamine Producing Bacteria (HBP) according toNiven's *et al.*, 1981¹².

Identification of *hdc* gene

Bromophenol blue 3ml was added to the DNA solution. The bands of DNA examined under UV. The fragment of the *hdc* gene (709 bp) was amplified by Primers *f* -*hdc* (5'- TCH ATY ARY AAC TGY GGTGAC TGG RG -3') and *r* -*hdc* (5'- CCC ACA KCA TBA RWG GDG TRT GRC C -3')^{13,14}. Twenty five micro litter of master mix PCR (50U/ml) *Taq* DNA polymerase , 400 μ M of each of nucleotides (promega). Twenty nanogram of DNA and 75 Peg mole of reface and forward primers. The PCR processes were run for 40 cycles at 94°C for 1 min , 54 °C for 1 min and 72°C for 1 min (Theromcycal).Agarose 1% in 1× TBE buffer (89 mMtris-borate , 2 mM EDTA, (PH 8.3).The PCR products were run at 86 V for 1 hr . Ethidium bromide (0.3µg/ml) were added to gels to be visualized bunds and UV transit- laminator were used for gene bands imaging.

Gene expression *hdc* detection by HPLC technique

Histamine was estimated by High performance liquid chromatography technique HPLC that produced by bacterial isolates the concentration of histamine based on absorption and wavelength¹⁵.

Antibiotic Susceptibility Test

Antibiotic resistance of bacteria of *Klebsiella pneumonia* was determined by MIC the Bio Meraux company according to16 antibiotics resistance profile.

Factors affecting Histamine production

PH effect: was tested by using two different Niven's medium pH(6 and 6.5) . Each one inoculate with loopful of culture and Incubate at 37°C for 24 hrs. and then measured histamine production by HPLC system¹⁵. **NaCl affect:** was tested by using two different Niven's medium NaCl(0.25% and 1mg) . Each one inoculate with loopful of culture and Incubate at 37°C for 24 hrs then measured histamine production by HPLC system¹⁵.

Temperature affect: was tested by using two different Niven's medium temperature $(35^{\circ}C \text{ and } 40^{\circ}C)$ each one inoculate with loopful of culture and Incubate at $37^{\circ}C$ for 24 hrs then measured histamine production by HPLC system¹⁵.

RESULTS

Fifty-one samples of sputum collected from patients suffering from respiratory problem in the advisory clinic for chest and respiratory disease and 255 isolates, fifty (50) isolates were identified by their morphology and biochemical characteristics as Klebsiella pneumoniae. Morphology of Klebsiella pneumoniae identified were large, dome-shaped, mucoid colonies on blood agar and lactose fermenting colonies on MacConkey agar. Microscopically appear as gram-negative, short, plump, straight rods were seen. The biochemical characters identified were negative indole test, negative methyl red test and positive for Voges-Proskauer test, citrate utilization test, urease test, acid and abundant gas production from glucose, lactose, sucrose, maltose and mannitol sugar fermentation tests and only 11 of Klebsiella pneumoniae were positive histamine production and by 21.56%. Histamine producing bacteria HPB were detected according to colorimetric

change from green to violet color by Niven's medium as shown in Figure 1. The number of *Klebsiella pneumoniae* isolates as non- HPB 40 out of 51 isolates (78.43%).





Figure 1. *Klebsiella pneumoniae* isolated grown on Niven's media (a): Positive bacteria HPB, (b):Negative bacteria Non HPB.

The presence of histamine producing bacteria can be recognized by the formation of purple halo colonies on Niven's agar and has been confirmed in PCR for the presence of the gene histadine decarboxylase hdc as shown Figure 2 which has the ability to convert histidine to histamine.



Figure 2. Positive Histamine producing bacteria isolates of *Klebsiella pneumoniae* for 709pb gene bands.

The level of histamine was estimated by HPLC technique the concentration of histamine in the *Klebsiella pneumoniae* was 46.06 μ g/ml as shown in Figure 3.



Figure 3. HPLC analysis for histamine concentration detection of *K. pneumonia*.

Antibiotic Susceptibility Test by MIC

Results are commonly reported as the Minimal Inhibitory Concentration (MIC), which is the lowest concentration of drug that inhibits the growth of the organism. An antibiotic is a kind of ubiquitous contaminant in the aquatic environment with industrial effluents and sewage discharge. The bacterial isolates were exposed to 16 antibiotics for susceptibility testing.





Figure 4. Sensitivity and resistance of the *Klebsiella pneumonia* towards various antibiotic, a: Normal flora HPB b: Normal flora non- HPB.

Antibiotic sensitivity and resistance test of 16 confirmed K. pneumoniae. This study showed that Klebsiella pneumonia histamine production highest degree of sensitivity against to Cefepime, Imipenem, Tobramycin and Trimethoprim 100%, while the isolates had been multidrug resistance Ampicilin 58.82%, Ciprofloxacin and Nitrofurantion(29.41%); respectively; as shown in Figure 4 a .Klebsiella pneumonia Non- histamine production, all of these antibiotics were categorized into three categories on the basis of their sensitivity. Results of one group had strains which were susceptible to Ceftazidime 88.23% and Aminoglycosides 58.82%. The second group had strains which were intermediate to antibiotics (Tobramycin and Levofloxacin). The third group contained strains which were resistant to synthetic penicillins(Amoxicillin, piperacillin) to 100% shown as Figure 4 b.

Factors affecting Histamine production

There are factors that affect the ability to produce histamine (pH, NaCl and Temperature).

 Table 1. Factors affecting the concentration of histamine in histamine-producing bacteria

Isolates	PH		NaCl mg		Temperature	
HPB µg/ml	6	6.5	1	0.25	35°C	40°C
K.pneumoniae	16.14%	29.04%	2.7%	21.5%	20.4%	16.19%

The results showed in the Table 1 factors affecting the production of histamine in HPB. *K* .*pneumoniae* had been the highest histamine concentration 29.04% μ g/ml in the pH6.5 and , while in the NaCl 1 mg had been lower 2.75% μ g/ml shown as Figure 5.











Figure 5. Factors affecting the concentration of histamine by HPLC analysis in *K*. *pneumonia* A: Effect of(pH 6.5),B: Effect of(pH 6.), C: Effect salts Nacl(1 mg), D: Effect salts Nacl (0.25 mg),E: Effect temperature(40 °C) and F: Effect temperature35°C.

DISCUSSION

This results showed that the isolates can change the color of Niven's medium, because presence of Purple Bromocresol and PH change due to the activity of histidine decarboxylase enzyme that convert acidic histidine to alkaline histamine¹⁶ which exploit the change of pH due to histamine formation and consequently the change in the color of the medium¹⁷. Results were also agreed with Bjornsdottir, K. and *et al*¹⁸, which used primer to determine the histamine *hdc* gene and found its size was 709bp. It was also found that all the bacteria that gave positive results in the detection of the bacteria producing histamine were given as a result of gene *hdc* amplification that responsible for production of histamine.

Bacteria species belonging to the Enterobacteriaceae family play a role in the reproducibility of biogenic amines, may be this is part of the amines fed^{19.20}. The present study is agreed with the results of²¹ showed that bacterial histamine producing can be classified into two groups : bacteria have producing a huge quantities of histamine (> 100mg / 100 ml) as Klebsiella pneumonia and Morganella morganiiand others species that produce less amounts of histamine (< 25 mg / 100 ml) as E. coli and Citrobacter freundii. The present study agreed with many research²² the less concentration of some microorganisms like E.coli and Citrobacter *freundii* that histidine conversion into histamine depends on their enzyme action and the ability to grow in environment. Affecting factors that assist in producing histamine were studied like pH, NaCl and temperature. Acording Figure 4 a and b. In vitro data showed a wide range of beta-lactams, aminoglycosides, Flouroquinolcin and other antibiotics which are useful for treatment of *Klebsiella* infections 23,24,25 . Both Gram positive and Gram negative bacteria have cell walls which is composed of heavily cross-linked peptidoglycan layers which are stimulated by cell-wall transpeptidases also known as penicillin binding protein(PBP). β-lactam antibiotics disturb peptide bond formation by acting as competitive inhibitors to these PBPs. These result in formation of irreversible covalent bonded penicilloylwith complexes weak cross-linked enzyme peptidoglycans, thus ease bacteria lyses and death²⁶. The antibiotic treatments for K. pneumoniae infections are Beta-lactams as carbapenems and cephalosporins. aminoglycosides such as quinolones and gentamycin. The treatments are ineffective toward some isolates of K. pneumoniae that have resistance mechanisms²⁷. K. pneumoniae contain two resistance mechanisms: enzymes production and formation of biofilm. Resistance had been shown against beta-lactams, sulfa methoxazoles, carbapenems, fluoroquinolones, trimethoprim, and aminoglycosides²⁸. Research have shown that antibiotic abuse develop to selective increasing β -lactam resistance in bacteria of the genus Enterobacteriaceae²⁹. Plasmid encoded resistance to broad spectrum cephalosporins is becoming a widespread phenomenon in clinical medicine. These antibiotics are destroyed by an array of different

extended spectrum β -lactamases (ESBLs). It had developed by mutation of TEM/SHV type β -lactamases. Plasmids coding these enzymes has been encountered in many species of the enterobacteriaceae but are often harbored by *K. pneumoniae*³⁰.

The results³¹ has been confirm that stress of the oxidation–reduction potential the reason for production of amines to enhanced the conditions to the effort to produce histamine and effectiveness of the enzyme HDC discouraging the presence of oxygen, also amines formed is strongly affected by temperature between 20-37°Care typical for bacterial growth of most those containing the enzyme remover carboxylaes and found the low temperature leads to growth reduce.

The present results have agreed with many studies^{32,33} of that factors affect the growth of bacteria producing histamine and the effectiveness of the carboxyl groups enzyme and on the accumulation of histamine such as temperature incubation and pH of the medium factors. The increasment of NaCl concentration at least from histamine accumulation this agreed with³¹. A number of researchers found^{34,35,36} that the degree of mild temperatures can done for the bacterial growth of histamine production in less than 3-4 hrs, studies also showed that histamine cannot break degree cooking temperature. It was found that increasing the proportion of histamine at reduced pH also was found that reduced histamine concentration if the medium treated with 5% NaCl concentration. Histamine producing bacteria is able to grow in a range of temperature 37 .

CONCLUSIONS

The *K. pneumonia* have a new virulence factor it can be cause or enhanced pathogenicity. The presence of new virulence agents in the bacterial normal flora is their ability to produce histamine, making it one of the causes of the irritable bowel disease. *K .pneumonia* have high sensitive against many antibiotics Trimethoprim, while the overall resistance pattern high resistance Ampicilin, Ambicillin/Sulbactam.

REFERENCES

- Rahamathulla M.P., Harish B.N., Mataseje L. and Mulvey M.R. "Carbapenem resistance mechanisms among blood isolates of *Klebsiella pneumoniae* and Escherichia coli". Afr. J. Microbiol. Res. 2016, 10(2):45-53.
- Hadi M., Mohammad R. and Zohreh Arab-Halvaii. "High prevalence of extended spectrum beta lactamase producing *Klebsiella pneumoniae* in a tertiary care hospital in Tehran, Iran". The Journal of infection in developing countries. 2010, 4(3): 132-138.
- Sarathbabu R., Ramani T.V., Bhaskararao K. and Supriya Panda. "Antibiotic susceptibility pattern of Klebsiella pneumoniae isolated from sputum, urine and pus samples". Journal of Pharmacy and Biological Sciences. 2012, 1(2):04-09.
- Sikarwar A.S. and Batra H.V."Challenge to healthcare: Multidrug resistance in *Klebsiella pneumoniae*". International Conference on Food Engineering and BiotechnologyIPCBEE. 2011, 9: 130-134.

- Harbarth S., Balkhy H., Goossens H., Jarlier V., Kluytmans J., Laxminarayan R. and Pittet D. "Antimicrobial resistance: One world, one fight! Antimicrob Resist Infect Control Antimicrobial Resistance and Infection Control". Antimicrobial Resistance and Infection Control. 2015, 4(49). Doi:10.1186/s13756-015-0091-2.
- Tzouvelekis L.S., Markogiannakis A., Psichogiou M., Tassios P.T., and Daikos G.L. "Carbapenemases in *Klebsiella pneumoniae* and Other Enterobacteriaceae: an Evolving Crisis of Global Dimensions". Clinical Microbiology Reviews. 2012, 25(4): 682–707.
- Fertas-Aissani R., Messai Y., Alouache S. and Bakour R. "Virulence profiles and antibiotic susceptibility patterns of *Klebsiella pneumonia* strains isolated from different clinical specimens". Pathol. Biol. 2013, 61(5):209-216.
- Chung T.H., Karkey A., Pham T.D., Boinett C.J., Cain A.K. and Ellington M. "A high-resolution genomic analysis of multidrug resistant hospital outbreaks of *Klebsiella pneumoniae*". EMBO Mol. Med. 2015, 7(3):227-39.
- Yeh K.M., Kurup A., Siu L.K., Koh Y.L., Fung C.P., Lin J.C., Chen T.L., Chang F.Y. and Koh T.H. "Capsular serotype K1 or K2, rather than magA and rmpA, is a major virulence determinant for *Klebsiella pneumoniae* liver abscess in Singapore and Taiwan". J. Clin. Microbiol.2007, 45(2):466-471.
- Wiskur B.J., Hunt J.J., and Callegan M.C."Hypermucoviscosity as a virulence factor in experimental *Klebsiella pneumoniae* endophthalmitis". Invest. Ophthalmol. Vis. Sci. 2008, 49(11):4931-4938. doi: 10.1167/iovs.08-2276. Epub 2008 Jun 27.
- Becker K.K., Southwick J., Reardon R.B. and acCormack J.N. "Histamine poisoning associated with eating tuna burgers". JAMA. 2001, 285(10): 1327–1330.
- Niven C.F., Jeffrey M.B. and Corlett D.A. (1981). "Differential Plating Medium for Quantitative Detection of Histamine – Producing Bacteria". Appl. Environ. Microbiol. 1981, 41(1): 321-322.
- Takahashi H., Kimura B., Yoshikawa M. and Fujii T."Cloning and sequencing of the histidine decarboxylase genes of gram – negative, histamine –producing bacteria and their application in detection and identification of these organisms". Applied . Environ. Microbial. 2003, 69(5): 2568-2579.
- Koohdar V.A., Razavilar V., Motalebi A.A., Mosakhani F., and Valinassab T."Isolation and Identification of Histamine-forming bacteria infrozen Skipjack tuna (*Katsuwonuspelamis*)". Iran. J. Fisher. Sci. 2011, 10 (4): 678-688.
- Cinquina A.L., Longo F., Cali A., De Santis L., Baccelliere R., and Cozzani R."Validation and comparison of analytical methods for the determination of histamine in tuna fish samples". J. Chromatogr. A. 2004, 1032(1-2): 79–85.
- Bjornsdottir-Butler. "Development of molecular-based methods for determination of high histamine producing bacteria in fish". International Journal of Food Microbiology. 2010, 139(3): 161-167.
- Meena T., Anita G., Hirekudel S. and Binaya B.N. "Dominance of Enterobacteria among Histamine-Producing Bacteria Isolated from Indian Mackerel". Advances in Microbiology. 2013, 3: 537-542.
- Bjornsdottir K., Bolton G.E., Mcclellan-Green P.D., Jaykus L.A. and Green D.P. "Detection of Gram-Negative Histamine-

Producing Bacteria in Fish: A Comparative Study". J. Food Prot. 2009, 72(9):1987–1991.

- Bover-Cid S., Hugas M., Izquierdo-Pulido M. and Vidal-Carou M.C. "Reduction of biogenic amine formation using a negative amino acid-decarboxylase starter culture for fermentation of fuet sausage". J. Food Protect. 2000, 63(2):237–243.
- Cardozo M., Lima K.S.C., França T.C.C. and Lima A.L.S. "Biogenic Amines: A Public Health Problem". Rev. Virtual Quim. 2013, 5(2): 149-168.
- Koohdar V.A., Razavilar V., Motalebi A.A., Mosakhani F. and Valinassab T. "Isolation and Identification of Histamine-forming bacteria in frozen Skipjack tuna (*Katsuwonuspelamis*)". Iranian Journal of Fisheries Sciences. 2011, 10(4):678-688.
- Gonzalez C.J., Santos J.A., Garcia-Lopez M.L., Gonzalez N. and Otero A. "Mesophilic aeromonads in wild and aquacultured freshwater fish". J. Food Protect. 2001, 64(5):687–691.
- Weisenberg S.A., Morgan D.J., Witter E.R. and Larone D.H. "Clinical outcomes of patients with *Klebsiella pneumonia* carbapenemase – producing *K. pneumoniae* after treatment with imipenem or meropenem". Diagn.Microbiol Infect Dis. 2009, 9: 130-134.
- ChanY.R., Liu J.S., Pociask D.A., Zheng M., Mietzner T.A. and Berger T."Lipocalin 2 is required for pulmonary host defense against Klebsiella infection". J. Immunol. 2009,182(8): 4947-56.
- J.M. Adams-haduch,, B.A. Potoski ,H.E. Sidjabat , D.L. Paterson and Y. Doi. "Activity to Temocillin against KPC-Producing Klebsiellapneumonia and Escherichia coli", Antimicrob Agents Chemother. 2009, 53(6): 2700-2701.DOI:10.1128/AAC.00290-09
- M.S. Wilke, L. Andrew, L. Natalie and C. J. Strynadka. "Blactamantibiotic resistant : a current structural prospective" Current Opinionin Microbiology. 2005, 8:525-533.
- Qureshi, S. Klebsiella Infections Treatment & Management (M. Bronze, Ed.). Retrieved November 29, 2015, from <u>http://emedicine.medscape.com/article/219907-treatment</u>
- Kumar, V., Sun, P., Vamathevan, J., Li, Y., Ingraham, K., Palmer, L., Brown, J. R. Comparative Genomics of *Klebsiella pneumoniae* Strains with Different Antibiotic Resistance Profiles. *Antimicrobial Agents and Chemotherapy*. 2011, 55(9), 4267– 4276.
- Sedláková, Karel Urbánek , Vladimíra Vojtová , Hana Suchánková3 , Peter Imwensi1 and Milan Kolář. "Antibiotic consumption and its influence on the resistance in Enterobacteriaceae". *BMC Research Notes*. 2014, 7:454. doi: 10.1186/1756-0500-7-454.
- 30. G.A. Jacoby. "Genetics of extended spectrum beta-lactamases". Eur. J. Clin. Microbiol. Infect. Dis 1994., 13: 2-11.
- Karovičova , J. and Kohajdova , Z. "Biogenic Amines in Food". Chem. Pap. 2005, 59(1): 70-79.
- Novella-Rodriguez, S. ; Veciana-Nogues, M. T.; Izquierdo-Pulido, M. M. and Vidal-Carou, M.C."Distribution of biogenic amines and polyamines in cheese". J. Food Sci. 2003, 68:750– 755.
- Fernández M, Linares DM, Rodríguez A and Alvarez MA. "Factors affecting tyramineproduction in *Enterococcus durans*IPLA 655. Appl. Microbiol. Biotechnol.2007, 73(6):1400–1406.
- Ross, T. and Sanderson, K. "A risk assessment of selected seafoods in NSW–Final report. Hobart". School of Agricultural Science, University of Tasmania, 2000.

- Kim ,S.H.; Price, R.J.; Morrissey, M.T.; Field, K.G.; Wei, C.I. and An ,H. "Occurrence of histamine- forming bacteria in albacore and histamine accumulation in muscles at ambient temperature". J. Food Sci. 2002, 67(4):1515-1521.
- Massuri Mahamudin, SitiHasmah Mohtar and Rozila Alias. "Effect of different storage conditions towards the formation of histamine producing bacteria in Canned Tuna (*Thunnus SPP.*)". Indian Journal of Fundamental and Applied Life Sciences. 2016, 6 (1): 82-87.
- Pons-Sánchez-Cascado, S.; Vidal-Carou, M.C.; Mariné-Font, A. and Veciana-Nogués, M.T. "Influence of the freshness grade of raw fish on the formation of volatile and biogenic amines during the manufacture and storage of vinegar-marinated anchovies". J. Agric.Food Chem. 2005, 539 (22): 8586-8592